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ROZPRAWA DOKTORSKA

Doświadczalne oraz farmakoekonomiczne aspekty polimerowych nośników leków

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Spis treści

Wykaz stosowanych skrótów

5-FdUMP	ang. 5-Fluorodeoxyuridine monophosphate, Monofosforan 5-fluorodeoksyurydyny
5-FU	5-fluorouracyl
ATR-FTIR	ang. Attenuated Total Reflectance Fourier Transform Infrared Spectroscopy, Spektroskopia osłabionego całkowitego odbicia w podczerwieni
CRC	ang. Colorectal Cancer, Rak jelita grubego
CT	ang. Computed Tomography, Tomografia komputerowa
DLS	ang. Dynamic Light Scattering, Dynamiczne Rozpraszanie Światła
DNA	ang. Deoxyribonucleic Acid, Kwas deoksyrybonukleinowy
DPD	ang. Dihydropyrimidine Dehydrogenase, Dehydrogenaza dihydropirymidynowa
DSC	ang. Differential Scanning Calorimetry, Skaningowa kalorymetria różnicowa
dTMP	ang. Thymidine Monophosphate, Monofosforan deoksytymidyny
dUMP	ang. Deoxyuridine Monophosphate, Monofosforan deoksyurydyny
EMA	ang. Europejska Agencja Leków, European Medicines Agency
FA	ang. Folic Acid, Kwas foliowy
HEA	Akrylan 2-hydroksyetylu
HT	ang. Hyperthermal Therapy, Terapia termiczna
LC	ang. Loading Capacity, Zawartość leku
LE	ang. Loading Efficiency, Wydajność procesu enkapsulacji
MRI	ang. Magnetic Resonance Imaging, Obrazowanie metodą rezonansu magnetycznego
MTHFR	ang. Methylenetetrahydrofolate Reductase, Reduktaza

	metylenotetrahydrofolianu
NIPAAm	N-izopropylokrylamid
NMR	ang. Nuclear Magnetic Resonance, Spektroskopia magnetycznego rezonansu jądrowego
RAFT/ MADIX	ang. Reverisble Addition-Fragemntation chain Transfer/ Macromolecular Design via Interchange of Xantathes, Odwracalne Addycyjno-Frgamentacyjne przeniesienie Łańcucha/ Projektowanie Makromolekularne poprzez Wymianę Ksantogenianów
RBC	ang. Red Blood Cells, Erytrocyty ludzkie
SEC	ang. Size Exclusion Chromatography, Chromatografia wykluczania / Chromatografia z rozdziałem wielkości
SEM	ang. Scaning Electron Microscopy, Skaningowy mikroskop elektronowy
SDDS	ang. Smart Drug Delivery Systems, Inteligentne systemy kontrolowanego dostarczania leków
PET	ang. Positron Emission Tomography, Pozytonowa tomografia emisyjna
PS	ang. Phosphatidylserine, Fosfatydyloseryna
PTD	ang. Photodynamic Therapy, Terapia fotodynamiczna
RGD / NLS	Sekwencja peptydowa składająca się z aminokwasów argininy, glicyny i kwasu asparaginowego (Arg-Gly-Asp, RGD) / ang. Nuclear Localization Sequences, Sekwencja sygnałowa
TEM	ang. Transmission Electron Microscopy, Transmisyjna mikroskopia elektronowa
TGA	ang. Thermogravimetric Analysis, Analiza termograwimetryczna
TS	ang. Thymidylate Synthase, Syntaza tymidylanowa
TT	ang. Target Therapy, Terapia celowana
UV–Vis	ang. Ultraviolet–Visible Spectroscopy

Rozdział 1. Wykaz publikacji będących podstawą rozprawy doktorskiej

Sumaryczny Impact Factor (IF) dla cyklu publikacji: **13.241**
Łączna liczba punktów według listy czasopism punktowanych Ministerstwa Edukacji i Nauki (MEiN): **280 punktów**.
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Wykaz publikacji będących podstawą rozprawy doktorskiej:

Publikacja I

Milewska Sylwia, Niemirowicz-Laskowska Katarzyna, Siemiaszko Gabriela, Nowicki Piotr, Wilczewska Agnieszka Zofia, Car Halina: Current Trends and Challenges in Pharmacoeconomic Aspects of Nanocarriers as Drug Delivery Systems for Cancer Treatment. International Journal of Nanomedicine, 2021: 16, s. 6593-6644, DOI: 10.2147/IJN.S323831, IF: **7.033**, MNiSW/MEiN: **140.0**. Cytowania: **10/12** (Web of Science/Scopus).

Publikacja II

Milewska Sylwia, Siemiaszko Gabriela, Wilczewska Agnieszka Zofia, Misztalewska-Turkowicz Iwona, Markiewicz Karolina Halina, Szymczuk Dawid, Sawicka Diana, Car Halina, Lazny Ryszard, Niemirowicz-Laskowska Katarzyna: Folic-acid-conjugated thermoresponsive polymeric particles for targeted delivery of 5-fluorouracil to CRC cells. International Journal of Molecular Sciences, 2023: 24(2), 1364, s. 1-25, DOI: 10.3390/ijms24021364, IF: **6.208**, MNiSW/MEiN: **140.0**.

Rozdział 2. Wprowadzenie

Nanotechnologia jest jedną z najbardziej dynamicznie rozwijających się dziedzin nauki i techniki, która łączy w sobie elementy innych nauk, w tym biologii, chemii, mechaniki oraz informatyki, co w konsekwencji nadaje jej interdyscyplinarny charakter dając podstawy do jej zastosowania w naukach medycznych. Odgrywa to znaczącą rolę w rozwoju medycyny, w tym terapii celowanej dedykowanej leczeniu nowotworów [1].

Produktami nanotechnologii są nanocząstki, które zgodnie z definicją są strukturami o wielkości do 100 nm przynajmniej w jednym wymiarze. Nanostruktury charakteryzują się unikalnymi parametrami fizykochemicznymi oraz właściwościami biologicznymi dzięki czemu ich zastosowanie w terapii (leczenie celowane, fotouczulanie, hipertermia), jak również w diagnostyce (detekcja, obrazowanie) różnych chorób, zwłaszcza chorób nowotworowych daje wiele nowych możliwości terapeutycznych. Na szczególną uwagę zasługuje użycie inteligentnych systemów kontrolowanego dostarczania leków (ang. Smart Drug Delivery Systems, SDDS) oraz terapia celowana (ang. Target Therapy, TT) jak również techniki diagnostyczne, tj. nowoczesne techniki obrazowania w czasie rzeczywistym [2-5].

Rak jelita grubego jest trzecim najczęściej diagnozowanym nowotworem na świecie. Na podstawie danych opublikowanych przez International Agency for Research on Cancer w 2020 r., w bazie GLOBOCAN 2020, wykazano, iż zdiagnozowano około 2 mln nowych przypadków raka jelita grubego, z czego większość –- ponad milion nowych przypadków (1065960) odnotowano u mężczyzn, zaś ponad 800 tys. nowych zachorowań stwierdzono u kobiet [6-7]. Ze względu na częstość występowania, rak jelita grubego znalazł się na drugim miejscu wśród kobiet i na trzecim miejscu wśród mężczyzn i w 2020 r. był odpowiedzialny za ponad 90 tysięcy zgonów [6-7]. Zgodnie z polskimi danymi epidemiologicznymi, rak jelita grubego jest drugim pod względem występowania i śmiertelności nowotworem, częstszy jest tylko rak płuca u mężczyzn i rak piersi u kobiet. Co roku jest diagnozowany u ponad 18 tys. osób, z czego 11 tys. pacjentów umiera [6-7]. Analizy predykcyjne umożliwiają prognozowanie i oszacowanie liczby nowych diagnozowanych przypadków nowotworów jelita grubego i wskazują, że w Polsce liczba ta będzie stale wzrastać aż do 2040 r. [8]. Szacuje się, że dynamika wzrostu względem 2018 r. w 2040 r. (13271 w 2018 r. i 17925 w 2040 r.) wyniesie ok. 35% [9].

W terapii pacjentów z zdiagnozowanym nowotworem jelita grubego lekiem z wyboru jest 5-fluorouracyl (5-FU) stosowany zarówno w monoterapii, w oparciu o schematy lekowe, a także w terapiach innowacyjnych, takich jak programy lekowe czy medycyna

spersonalizowana. Powyższe wynika z faktu, iż bazując na danych pochodzących z analiz cech fenotypowych, transkryptomicznych i metabolomicznych tkanek, 5-FU ma udowodnione badaniami naukowymi wyższe powinowactwo do tkanek okrężnicy niż do pozostałych organów w ciele człowieka [10]. Z chemicznego punktu widzenia, 5-FU jest fluorową pochodną uracylu należącą do antymetabolitów pirymidyn, których mechanizm działania polega na inhibicji syntazy tymidylanowej (ang. Thymidylate Synthase, TS) [11-12]. Podanie pacjentowi 5-FU prowadzi do inaktywacji większości cząsteczek leku przez dehydrogenazę dihydropirymidynową (ang. Dihydropyrimidine Dehydrogenase, DPD), a następnie pozostała część jest przekształcana do monofosforanu 5-fluorodeoksyurydyny (5-FdUMP), która hamuje aktywność syntazy tymidylanowej, w konsekwencji uniemożliwiając metylację monofosforanu deoksyurydyny (ang. Deoxyuridine Monophosphate, dUMP) i monofosforanu deoksytymidyny (ang. Thymidine Monophosphate, dTMP) [12]. Wyżej scharakteryzowane procesy powodują znaczne zmniejszenie zdolności komórek do replikacji i naprawy DNA, co interferuje w proces ich podziału. Wynikiem terapii jest zahamowanie proliferacji komórek nowotworowych i skierowanie ich na szlak apoptozy bądź nekrozy [12-13].

Ocena skuteczności leczenia w oparciu o 5-FU dowodzi, że wskaźniki odpowiedzi pacjentów na leczenie są niskie, co ma bezpośredni związek z predyspozycjami do rozwoju wrodzonej lub nabytej oporności na ten lek. Powyższe może doprowadzić do poważnych ograniczeń w efektywnym leczeniu pacjentów i w rezultacie osiągnięciu niewielkich korzyści terapeutycznych [14]. Co więcej, należy podkreślić, że terapia 5-FU obciążona jest ryzykiem wystąpienia znacznych efektów ubocznych, a ich nasilenie może być większe, jeżeli lek będzie podawany w skojarzeniu z innymi chemioterapeutykami. Działania niepożądane można podzielić na te, które występują bardzo często, często, rzadko oraz bardzo rzadko. W terapii 5-FU bardzo często występują zaburzenia hematologiczne objawiające się zaburzeniem ilości, jak i funkcji komórek krwi i szpiku kostnego, widocznymi jako leukopenia, agranulocytoza, małopłytkowość oraz niedokrwistość. Do działań niepożądanych, które występują często można zaliczyć: nudności, wymioty, biegunkę oraz jadłowstręt. W przebiegu leczenia mogą wystąpić również inne działania niepożądane takie jak: gorączka, przetrwałe zapalenie błon śluzowych jamy ustnej, trudne do leczenia owrzodzenia i krwawienia z całego odcinka przewodu pokarmowego. Możliwe są również reakcje nadwrażliwości takie jak świąd, zmiany skórne czy przemijające zmiany w obrębie paznokci, a także sporadycznie odnotowuje się niebezpieczne reakcje anafilaktyczne. Podczas terapii 5-FU u pacjentów odnotowano znaczną kardiotoksyczność manifestującą się jako wstrząs

kardiogenny, nagłe zatrzymanie krążenia, zawał mięśnia sercowego. Stwierdzono także związek terapii 5-FU z występowaniem niedokrwiennego udaru mózgu, a także martwicy wątroby [15-16].

Warto zwrócić uwagę na fakt, iż w leczeniu raka jelita grubego osiągnięto pewne postępy, ale pacjenci w stadium przerzutowym, którzy nie kwalifikują się do intensywnej terapii wciąż mają do dyspozycji niewiele opcji terapeutycznych. Jeżeli choroba jest rozpoznana w zaawansowanym stadium to część pacjentów jest w średnim lub złym stanie ogólnym. U takich pacjentów nie stosuje się agresywnego leczenia, a opcji terapeutycznych jest zdecydowanie mniej. Wprowadzenie metody leczenia opartej na nanotechnologii daje ogromne możliwości w pokonaniu ograniczeń konwencjonalnej terapii przeciwnowotworowej [11]. Opracowuje się rożne formy i struktury nanomateriałów, które mogą być w sposób znamienny wykorzystywane do leczenia nowotworów nie tylko za pomocą chemioterapii, ale także radioterapii (promieniowanie Gamma, Beta, X) [17-19] oraz terapii fotodynamicznej (ang. Photodynamic Therapy, PTD) [20-22] lub termicznej (ang. Hyperthermal Therapy, HT) [23-25].

Dane literaturowe wskazują, iż 5-FU podawany w postaci iniekcji pozwala uzyskać obiektywną odpowiedź u 20% pacjentów, zaś podawany w ciągłym wlewie dożylnym zwiększa odsetek odpowiedzi, ale nie wywiera wpływu na czas przeżycia chorych. Udowodniono, że leczenie uzupełniające oparte na 5-FU i folinianie wapnia zmniejsza względne ryzyko wznowy nowotworu o 45%, a zgonu o 33%. Takie rozwiązanie również prowadzi do zwiększenia odsetka trzyletnich przeżyć wolnych od choroby prawie o 20% (z 44% do 62%), a przeżyć całkowitych o ponad 10% (z 64% do 76%). Innym chemioterapeutykiem wykazującym podobną skuteczność jest kapecytabina, której stosowanie nie wymaga konieczności przebywania pacjentów na oddziałach czy w ambulatoriach szpitalnych. Dodanie oksaliplatyny do powyższych leków zwiększa skuteczność leczenia tylko w niewielkim stopniu, a nasila objawy niepożądane, takie jak ciężka polineuropatia [26].

Eksperci podkreślają, że zaangażowane nowego rodzaju rozwiązań terapeutycznych i metod diagnostycznych pozwala na wydłużenie przeżycia pacjenta ze zdiagnozowanym nowotworem jelita grubego nawet trzykrotnie w porównaniu do poprzednich dziesięcioleci - wówczas można było przedłużyć życie pacjenta z rozsianym rakiem jelita grubego o 12 miesięcy [27].

Rosnąca liczba doniesień literaturowych potwierdza, iż zastosowanie rozwiązań z dziedziny nanotechnologii w porównaniu ze standardowym schematem leczeniem znacznie

ogranicza występowanie działań niepożądanych [25, 28]. Niewątpliwe, posiadanie możliwości uwolnienia leku w obrębie nowotworu jest znacznie korzystniejsze od ogólnoustrojowej dystrybucji cytostatyku [29-31]. Powyższe związane jest ogólną ideą zastosowania inteligentnych, wrażliwych na bodźce nośników leków jako systemów do selektywnego i ukierunkowanego dostarczenia substancji aktywnej do docelowych komórek i tkanek, a następnie uwolnienia jej w sposób kontrolowany, pomijając przy tym zdrowe narządy i części ciała [5]. Kolejnym istotnym aspektem związanym z osiągnięciem przez nanonośnik punktu docelowego jest jego funkcjonalizacja tj. zastosowanie tzw. ligandów nakierowujących. Ich obecność nadaje nośnikom leków charakter multifunkcjonalny, co stanowi kluczowy element w nowoczesnej terapii celowanej. Powyższa idea opiera się na tzw. bio-targetingu czyli podłączeniu do powierzchni nanotransportera zawierającego w sobie cząsteczki leku, charakterystycznych dla danego nowotworu ligandów receptorowych, np. cząsteczek kwasu foliowego, peptydów integrynowych o sekwencji RGD / NLS lub cząsteczek aktywnych błonowo, np. peptydów bądź ich mimetyków sterydowych. Osiągnięcie wysokiego powinowactwa i specyficzności w stosunku do komórek guza z pominięciem aktywności w stosunku do komórek niezmienionych nowotworowo opiera się zarówno na unikalnych właściwościach fizykochemicznych i morfologicznych nanoukładów oraz ich właściwościach farmakologicznych [25, 32-33].

Jednymi z najbardziej obiecujących kandydatów rozpatrywanych jako potencjalne nośniki leków są nośniki polimerowe. Ze względu na wysoką biokompatybilność z komórkami gospodarza, zdolności regeneracyjne i immunomodulujące nanocząstek polimerowych oraz możliwość poprawy parametrów farmakokinetycznych stosowanych obecnie chemioterapeutyków, istnieje możliwość zastosowania tego typu nanonośników jako komponentów bądź też jako nowych form leków w nowoczesnej terapii przeciwnowotworowej [5, 11]. W związku z powyższym, projektowane są tzw. inteligentne nośniki polimerowe [34]. Ich nazwa związana jest z faktem, iż w odpowiedzi na zmianę warunków środowiska, tj. pH, temperatury, obecność pola magnetycznego są one zdolne do uwolnienia enkapsulowanej substancji aktywnej w sposób bodźco-zależny. Cząsteczka leku może być uwięziona w nośniku polimerowym w wyniku utworzenia wiązań kowalencyjnych lub poprzez oddziaływania niekowalencyjne. W idealnych warunkach substancje czynne będą zakotwiczone w nośniku o dużej ładowności z zapewnioną zdolnością do uwalniania ich w sposób kontrolowany [33].

Zastosowanie nośników leków na bazie polimerów termowrażliwych sfunkcjonalizowanych ligandem nakierowującym zdolnych do skompleksowania substancji

aktywnej może przyczynić się do zniesienia problemu lekooporności komórek nowotworowych [35], definiowanej jako ich niewrażliwość na leczenie dedykowanym chemioterapeutykiem. W skutek lekooporności zastosowanie założonego w wytycznych postępowania terapeutycznego jest w dużym stopniu ograniczone, uniemożliwiając tym samym efektywną terapię [36]. Wyróżnia się różne mechanizmy odpowiedzialne za wytworzenie lekooporności przez komórki nowotworowe. Należą do nich m. in.: zdolność komórek zmienionych nowotworowo do hamowania apoptozy, co ma związek z rozregulowaniem szlaku apoptozy; nadekspresją białek transportowych, które są odpowiedzialne za usuwanie chemioterapeutyku z komórek; aktywnością enzymów i / lub białek docelowych, która jest zależna od zwiększenia ich ekspresji w komórkach zmienionych chorobowo lub zmniejszenia powinowactwa względem leku, co ostatecznie wpływa na zaburzony targeting; zmieniona adhezja komórek objętych procesem nowotworowym; nasilenie procesów naprawczych uszkodzonego DNA w wyniku działania produktów leczniczych [36]. W przypadku 5-FU znaczącą rolę w powstawaniu zjawiska lekooporności ma zmieniony metabolizm tego leku.

Przeprowadzone przez nas badania poparte doniesieniami literaturowym potwierdzają, iż zastosowanie swoistych, wrażliwych na bodźce układów polimerowych jak i nanohybryd polimerowo-magnetycznych jako nośników leków bądź też składowych terapii kombinowanej może uwrażliwić komórki nowotworowe na dany chemioterapeutyk [28]. Powyższe może być zrealizowane kilkoma drogami. Pierwsza z nich bazuje na mechanizmie wzajemnego oddziaływania pomiędzy nośnikiem, a komórką neoplastyczną. Nośnik ten może zmienić drogę transportu leku do wnętrza komórki znosząc tym samym lekooporność o charakterze błonowym [28, 35]. Drugi mechanizm opiera się na zdolności nośników do uwolnienia substancji aktywnej pod wpływem bodźców zewnętrznych, np. w środowisku o obniżonym pH lub podwyższonej temperaturze. W związku z powyższym, właściwości nanomateriałów mogą być szczególnie wykorzystywane do leczenia nowotworów nie tylko za pomocą chemioterapii, ale także w metodzie leczenia wykorzystującej terapię fotodynamiczną lub hipertermię [28, 31].

Powyższe ma też znaczenie w aspekcie farmakoekonomicznym, ze względu na fakt, iż zastosowanie nośników w terapii nowotworów może przyczynić się do racjonalizacji farmakoterapii chorób nowotworowych, zwłaszcza w zakresie wydatków ponoszonych na leczenie, co w przyszłości może ułatwić decydentom ochrony zdrowia identyfikację potrzeb pacjentów onkologicznych [11, 37-45]. Co więcej, warto zauważyć, że w 2018 r. średni koszt terapii rocznej leków finansowanych w ramach programów lekowych w przeliczeniu na

pacjenta na podstawie realnej wartości refundacji rozliczonych mg leku wynosił nieco ponad 40 tys. zł [46]. Analiza skuteczności leczenia przy zastosowaniu 5-FU pokazuje, że wskaźniki odpowiedzi pacjentów na leczenie są niskie, co jest bezpośrednio związane ze skłonnością do rozwoju wrodzonej lub nabytej oporności na ten lek [12]. Może to skutkować poważnymi ograniczeniami w efektywnym leczeniu pacjentów i niewielkimi korzyściami terapeutycznymi [47-48]. Należy podkreślić, iż dzięki przeprowadzeniu różnego rodzaju analiz farmakoekonomicznych możliwe jest oszacowanie redukcji w kosztach przyszłego leczenia oraz wzrostu produktywności, wynikającego z mniejszej chorobowości i wydłużonego przeżycia pacjentów onkologicznych [37-45]. W tym celu stosuje się tzw. wskaźniki efektywności diagnostyki onkologicznej i leczenia onkologicznego, np. wskaźnik przeżyć 5-letnich, unikanie niepotrzebnej hospitalizacji w trakcie chemioterapii lub czas od przeprowadzenia konsylium do rozpoczęcia pierwszego etapu leczenia raka [49].

Podsumowując, zastosowanie inteligentnych systemów nanotargetingu jakimi są polimerowe nośniki leków możne przyczynić się do znacznej poprawy efektywności dostarczania substancji czynnej zawartej w nośniku bezpośrednio do miejsca zmienionego chorobowego. Ma to również przełożenie na poprawę skuteczności leczenia oraz redukcji i nasilenia działań niepożądanych takich jak, np. immunosupresja szpiku kostnego, ostra kardiotoksyczność oraz przewlekłe zapalenie błony śluzowej jelit. W związku z powyższym, podczas projektowania systemów dostarczania leków należy rozważyć wiele składowych, w tym drogę podawania, formulację nośników i późniejszy transport przez błony biologiczne, profil molekularny nowotworu, obecność receptorów powierzchniowych jak również szczególne warunki, jakie panują w miejscu zmienionym chorobowo (pH, temperatura). W konsekwencji, należy wykorzystać rozwiązania syntetyczne pozwalające na otrzymanie materiałów wrażliwych na bodźce oraz podatnych na dalszą funkcjonalizację powierzchniową, aby kontrola procesu dostarczania oraz uwalniania leku była osiągnięta w sposób ukierunkowany w miejscach objętych procesem patologicznym, w odpowiedzi na określone warunki endogenne lub przy zastosowaniu bodźców zewnętrznych [50]. Z farmakoekonomicznego punktu widzenia, wdrożenie nośników leków może w konsekwencji wpłynąć na obniżenie kosztów bezpośrednich i pośrednich procesu leczenia poprzez poprawę *compliance* oraz jakości życia pacjentów, u których zdiagnozowano nowotwór [37].

Rozdział 3. Cel pracy

Terapia celowana (TT) realizowana poprzez zastosowanie inteligentnych transporterów leków (SDDS) to metody dostarczania leków stosowane w nanofarmakologii, których celem jest poprawa parametrów farmakokinetycznych, zwiększenie efektywności leczenia przeciwnowotworowego, jak również ograniczenie liczby działań niepożądanych towarzyszących standardowym schematom leczenia. Obecnie istotnym wyzwaniem w praktyce klinicznej raka jelita grubego jest znalezienie sposobu na poprawę skuteczności terapii przy zastosowaniu 5-FU. Efektywna farmakoterapia, może przyczynić się do obniżenia kosztów bezpośrednich i pośrednich procesu leczenia oraz może mieć wpływ na poprawę *compliance* pacjenta.

W ramach niniejszej rozprawy doktorskiej dokonano przeglądu literatury (**Publikacja I**) dotyczącej nowych możliwości oraz wyzwań w leczeniu nowotworów w tym jelita grubego z wykorzystaniem osiągnięć nanotechnologii oraz medycyny spersonalizowanej. W pracy podniesiono także aspekt farmakoekonomiczny stosowania systemów dostarczania leków w terapii przeciwnowotworowej uwzględniając ogromny potencjał nanomedycyny w zakresie doskonalenia i rozwoju systemu ochrony zdrowia.

Celem rozprawy doktorskiej było rozpoznanie możliwości zastosowania nowych termo wrażliwych nanocząstek polimerowych sfunkcjonalizowanych kwasem foliowym jako nośników 5-FU w terapii raka jelita grubego w modelu *in vitro* wykorzystującym linie komórkowe o zmiennej wrażliwości na chemioterapeutyk (**Publikacja II**). Nośniki polimerowe na bazie akrylanu 2-hydroksyetylu i N-izopropylokrylamidu scharakteryzowane z wykorzystaniem szeregu metod fizykochemicznych obejmujących analizy spektralne, chromatograficzne oraz mikroskopowe oceniono jako transportery 5-FU. Ocenę aktywności biologicznej obejmującą hemokompatybilność oraz cytotoksyczność przeprowadzono w stosunku do reprezentantów komórek gospodarza, tj: erytrocytów, monocytów/makrofagów, kardiomiocytów, fibroblastów, jak również do przedstawicieli komórek raka jelita grubego charakteryzujących się zróżnicowanym profilem molekularnym i wrażliwością na 5-FU. W dalszym etapie przeprowadzono analizę wpływu badanych nośników na procesy proliferacji komórek nowotworowych oraz ich zdolności do indukcji apoptozy bądź nekrozy. Na podstawie otrzymanych wyników dokonano oceny wpływu zastosowania rozwiązań opartych na dynamicznie rozwijającej się dziedzinie jaką jest nanotechnologia na aspekty doświadczalne i farmakoekonomiczne leczenia nowotworu jelita grubego z wykorzystaniem polimerowych nośników leków.

Rozdział 4. Realizacja celów naukowych – materiały, metody badawcze, wyniki badań

4.1. Materiał i metody

4.1.1. Polimerowe nośniki leków

4.1.2. Synteza i charakterystyka właściwości fizykochemicznych

Synteza i charakterystyka właściwości fizykochemicznych oraz cech morfologicznych ocenianych w ramach niniejszej rozprawy doktorskiej polimerowych nośników 5-FU zostały wykonane w ramach współpracy Zakładu Farmakologii Doświadczalnej Uniwersytetu Medycznego w Białymstoku, a Zakładem Polimerów i Syntezy Organicznej Uniwersytetu w Białymstoku i były wykonane przez zespół kierowany przez Panią Prof. Agnieszkę Zofię Wilczewską pod nadzorem promotora pomocniczego – Pani dr n. chem. Iwony Misztalewskiej-Turkowicz, w których również uczestniczyłam.

Nośniki leków były otrzymane w wyniku reakcji kontrolowanej polimeryzacji wolnorodnikowej RAFT/MADIX (ang. Reverisble Addition-Fragemntation chain Transfer/Macromolecular Design via Interchange of Xantathes). Kopolimery (PHEA-*b*-PNIPAAm) na bazie akrylanu 2-hydroksyetylu (HEA) i N-izopropylokrylamidu (NIPAAm) (różniące się długością bloku PHEA) były następnie modyfikowane kwasem foliowym (ang. Folic Acid, FA). Otrzymano kopolimery o podobnej zawartości cząsteczek kwasu foliowego (PTF-1, PTF-2, PTF-3) różniące się ilością wolnych grup hydroksylowych. Nośniki te wykorzystano do enkapsulacji 5-fluorouracylu. Oceniono wydajność procesu enkapsulacji (ang. Loading Efficiency, LE) i zawartości leku (ang. Loading Capacity, LC) w nośnikach.

Przed ewaluacją biologiczną, nośniki zostały dokładnie scharakteryzowane pod kątem właściwości fizykochemicznych oraz cech morfologicznych. W powyższym celu zastosowane zostały następujące techniki i metody badawcze:

- chromatograficzne: SEC (ang. Size Exclusion Chromatography),
- techniki spektralne: NMR (ang. Nuclear Magnetic Resonance), ATR-FTIR (ang. Attenuated Total Reflectance Fourier Transform Infrared Spectroscopy), UV–Vis (ang. Ultraviolet–Visible Spectroscopy), DLS (ang. Dynamic Light Scattering), z-potencjał - potencjał zeta (potencjał ζ),
- Analizy termalne: TGA (ang. Thermogravimetric Analysis), DSC (ang. Differential Scanning Calorimetry),

- Analizy mikroskopowe: SEM (ang. Scaning Electron Microscopy), TEM (ang. Transmission Electron Microscopy).

4.1.3. Ocena właściwości biologicznych

Właściwości biologiczne otrzymanych nośników na poziomie *in vitro* przeprowadzono poprzez ocenę nośników w zakresie biokompatybilności z komórkami prawidłowymi wykorzystując: erytrocyty ludzkie (ang. Red Blood Cells, RBC), komórki monocytów linii THP-1, fibroblastów linii CRL-1475 oraz linii CCD-112 CoN oraz kardiomiocytów linii H9C2 (2-1). Potencjał przeciwnowotworowy nośników zbadano w stosunku do komórek raka jelita grubego linii DLD-1, Caco-2 i HT-29, różniących się profilem molekularnym oraz wrażliwością na chemioterapeutyk (5-FU).

Hemokompatybilność badanych nośników oceniono stosując test hemolizy erytrocytów po 1 h inkubacji z nośnikami bez leku oraz z nośnikami z enkapsulowanym 5-FU w stężeniu (0,5 mg/mL). Aktywność metaboliczną korelującą z przeżywalnością badanych komórek monocytów THP-1 oceniono testem z użyciem soli tetrazolowej MTS. Przeżywalność komórek prawidłowych linii CRL-1475, CCD-112 CoN oraz H9C2 (2-1), oraz nowotworowych linii DLD-1, Caco-2 i HT-29 zbadano wykorzystując test czerwieni obojętnej. Wpływ badanych związków na proces proliferacji komórek nowotworowych przeprowadzono za pomocą analizy fluorescencyjnej testem resazuryny. Powyższe analizy przeprowadzono po 24 h inkubacji, w obecności otrzymanych nośników leków z oraz bez enkapsulowanego chemioterapeutyku (stężenia 0,1 i 0,5 mg/mL) w porównaniu do chemioterapeutyku w formie wolnej (5 i 25 μg/mL). Celem zbadania mechanizmu odpowiedzialnego za redukcję liczby żywych komórek po inkubacji z badanymi związkami, przeprowadzono test multipleksowy obejmujący ocenę zdolności nośników do indukcji apoptozy i/lub nekrozy w komórkach raka jelita grubego wykorzystując technikę luminometryczną oraz fluorymetryczną – zestaw RealTime-Glo™ Annexin V Apoptosis and Necrosis Assay.

4.2.Wyniki

Nośniki leków były otrzymane w wyniku reakcji kontrolowanej polimeryzacji wolnorodnikowej RAFT/MADIX. Sfunkcjonalizowane kwasem foliowym kopolimery blokowe (PHEA-*b*-PNIPAAm) z postmodyfikacją cząsteczką kwasu foliowego (FA) wykorzystano do enkapsulacji 5-fluorouracylu. Wydajność procesu enkapsulacji (LE) wnosiła ~ 53%, a zawartości leku (LC) w nośnikach klasyfikowała się w granicach 9-12%. Na tle dotychczas opublikowanych danych, otrzymane wyniki wydajności enkapsulacji jak i zawartości leku w nośniku są jak najbardziej zadowalające, gdyż zazwyczaj zawartość nie przekracza granicy 10% [51]. Przeprowadzone w kolejnym etapie analizy spektralne, termalne i mikroskopowe potwierdziły strukturę oraz morfologię otrzymanych nośników. Dokładna charakterystyka kandydatów na nośniki leków, pod kątem ich właściwości fizykochemicznych ma ogromne znaczenie z względu na fakt, iż to one determinują właściwości biologiczne otrzymanych materiałów. Ponadto, potwierdzenie na podstawie powyższych analiz jednorodności i homogenności badanych kopolimerów wywiera znaczący wpływ na powtarzalność i odtwarzalność otrzymywanych wyników jak i na bezpieczeństwo ich stosowania [52-53].

Nośniki sfunkcjonalizowane kwasem foliowym na bazie kopolimerów akrylanu 2-hydroksyetylu (HEA) i N-izopropylokrylamidu (NIPAAm) różniące się długością bloku PHEA oceniono pod kątem właściwości biologicznych na poziomie *in vitro*. Przeprowadzono ocenę nośników w zakresie biokompatybilności z komórkami prawidłowymi angażując w tym celu reprezentantów komórek gospodarza - erytrocyty ludzkie (RBC) oraz komercyjnie dostępne linie komórkowe: monocytów (THP-1), fibroblastów skóry (CRL-1475), i okrężnicy (CCD-112 CoN) oraz kardiomiocytów (H9C2 (2-1)). Zbadano także potencjał cytotoksyczny w stosunku do komórek nowotworowych trzech linii raka jelita grubego (DLD-1, Caco-2 i HT-29).

Badania rozpoczęto od oceny hemokompatybilności nośników. Analiza ta jest jednym z kluczowych parametrów determinujących kliniczne zastosowanie materiałów mających kontakt z krwią. Powyższe jest istotne ze względu na fakt, iż przy braku zgodności nośników z komórkami gospodarza, konsekwencją interakcji w przypadku nośników leków podanych droga dożylną może być m. in. zagrażająca życiu indukcja reakcji hemolizy wewnątrznaczyniowej. W tym celu zastosowano test hemolizy bazujący na ocenie spektrofotometrycznej uwalniania hemoglobiny z komórek krwinek czerwonych inkubowanych z nośnikiem przez okres 1 h. Pomiary aktywności hemolitycznej testowanych

nośników wykazały, że niezależnie od obecności 5-FU wszystkie badane kopolimery wykazują aktywność hemolityczną w granicach 1-2%. Bazując na danych opublikowanych przez Webera i wsp. [54], dedykowanych biomateriałom implantacyjnym, materiały powodujące hemolizę < 2% klasyfikuje się jako biozgodne, przez co badane przez nas kopolimery można uznać jako niehemolityczne.

Podczas chemioterapii 5-FU, jednymi z częściej występujących skutków toksycznych są hematologiczne działania niepożądane obejmujące między innymi ciężką mielosupresję. Stan ten prowadzi do zaburzeń w układzie immunologicznym, czego konsekwencją jest zwiększenie podatności pacjentów onkologicznych na zagrażające życiu zakażenia oportunistyczne [55]. Toksyczność hematologiczna związana z terapią 5-FU prowadzi do zmniejszenia liczby wszystkich rodzajów komórek krwi obwodowej, objawiający się neutropenią, monocytopenią i trombocytopenią. W obecnym badaniu poddano ocenie wpływ zsyntetyzowanych nośników polimerowych jako potencjalnych czynników redukujących toksyczność hematologiczną wywołaną przez 5-FU. W tym celu oceniono wpływ 24 godzinnej inkubacji nośników z i bez obecności 5-FU z komórkami monocytów linii THP-1. Należy podkreślić, iż komórki linii THP-1 są powszechnie stosowanym modelem odwzorowującym funkcje ludzkich komórek monocytów/makrofagów [56]. Ich wybór był także podyktowany faktem, iż zmniejszenie liczby monocytów na wczesnym etapie chemioterapii, jest ważnym czynnikiem predykcyjnym wystąpienia ciężkiej neutropenii [57].

Otrzymane wyniki wykazały, że po inkubacji komórek z 5-FU w formie wolnej odnotowano około 30-40% zmniejszenie aktywności metabolicznej badanych komórek, co z kolei jest bezpośrednio związane ze zmniejszeniem proliferacji i żywotności komórek. Traktowanie komórek THP-1 zsyntetyzowanymi nośnikami bez enkapsulowanego cytostatyku w stężeniu 0,5 mg/mL spowodowało znamienne statystycznie zwiększenie aktywności metabolicznej badanych komórek. Powyższe jest charakterystyczne w przypadku inkubacji komórek fizjologicznych z nośnikami polimerowymi na bazie NIPAAm i świadczy o ich kompatybilności i zdolności do proliferacji. Niezwykle istotnym był wynik otrzymany dla kopolimerów z enkapsulowanym 5-FU, dla których nie odnotowano znamiennego statystycznie obniżenia aktywności metabolicznej badanych komórek, niezależnie od zastosowanego stężenia. Wskazuje to na korzystny, cytoprotekcyjny wpływ nośników, który zmniejsza cytotoksyczne działanie badanego antymetabolitu.

Kardiotoksyczność manifestująca się u pacjentów poddanych terapii 5-FU jako bóle wieńcowe, zawał mięśnia sercowego, a w konsekwencji śmierć jest jednym z najcięższych efektów niepożądanych chemioterapii, z którymi borykają się pacjenci onkologiczni. W

związku z powyższym, w kolejnym etapie przeprowadzono badania kompatybilności w stosunku do reprezentantów komórek mięśnia sercowego, kardiomiocytów linii (H9C2 (2-1)). Otrzymane wyniki wskazują, że nośniki polimerowe bez jak i z ekapsulowanym 5-FU stosowane w obu badanych stężeniach (0,1 mg/mL oraz 0,5 mg/mL) nie zmniejszały żywotności kardiomiocytów w porównaniu do kontroli. W przypadku inkubacji komórek z 5-FU w stężeniach 5 i 25 μg/mLw formie wolnej zaobserwowano znamienne statystycznie zmniejszenie ich żywotności – o ponad 60%. Należy również podkreślić, że nośniki kopolimerowe z enkapsulowanym 5-FU wywierają statystycznie istotnie słabszy efekt cytotoksyczny niż 5-FU w postaci wolnej. Powyższe wskazuje na potencjał tych nośników do zapobiegania rozwojowi kardiotoksyczności wywołanej chemioterapią u pacjentów z nowotworem jelita grubego leczonych schematem opartym na 5-FU.

5-fluorouracyl jest silnym inhibitorem proliferacji fibroblastów, komórek macierzy zewnątrzkomórkowej odpowiedzialnych za procesy regeneracji oraz nadawanie skórze wytrzymałości i sprężystości. Pacjenci poddawani chemioterapii opartej na 5-FU wykazywali znaczące problemy z prawidłowym gojeniem się ran [58]. W związku z powyższym, w badaniach ocenialiśmy również wpływ nanonośników polimerowych sfunkcjonalizowanych kwasem foliowym jako systemów dostarczania 5-FU na żywotność komórek fibroblastów skóry i okrężnicy. Otrzymane wyniki wykazały, że leczenie komórek fibroblastów skóry za pomocą 5-FU w postaci wolnej spowodowało znaczne zmniejszenie ich żywotności (poniżej 10%), zaś w przypadku fibroblastów okrężnicy było to około 40%. Zastosowanie zsyntetyzowanych nośników jak transporterów 5-FU skutkowało statystycznie istotnym mniejszym efektem cytotoksycznym, w porównaniu do traktowania 5-FU w formie wolnej. W przypadku fibroblastów skóry, żywotność była 5-krotnie wyższa, gdy komórki były traktowane 5-FU w postaci enkapsulowanej. Natomiast w przypadku traktowania fibroblastów okrężnicy zaobserwowano 2-krotny wzrost żywotności komórek po inkubacji z użyciem nośników zmodyfikowanych kwasem foliowym wraz z enkapsulowanym 5-FU w stężeniu 0,1 mg/mL. Uzyskane wyniki sugerują, że zastosowanie nośnika polimerowego do dostarczania cytostatyków może być pomocne w redukcji działań niepożądanych i toksycznych skutków ubocznych związanych z leczeniem przeciwnowotworowym.

Ostatnim etapem naszych badań była ocena potencjału przeciwnowotworowego zsyntetyzowanych kopolimerów. W tym celu oceniono cytotoksyczność badanych nośników z i bez enkapsulowanego 5-FU (0,1 i 0,5 mg/mL) w porównaniu do 5-FU w formie wolnej (5 i 25 μg/mL), w stosunku do trzech linii komórkowych raka jelita grubego: DLD-1, CaCo-2 oraz HT-29, różniących się między sobą profilem molekularnym oraz wrażliwością na

stosowany antymetabolit. Wszystkie z badanych linii komórkowych posiadają potwierdzoną obecność receptorów dla kwasu foliowego głównie typu αFR [59-62].W wyniku 24-godzinnej inkubacji zauważono, że traktowanie wszystkich rodzajów linii komórkowych (DLD-1, Caco-2 i HT-29) cytostatykiem w stężeniu 5 μg/mL w postaci wolnej nie wpłynęło znamiennie na zmniejszenie odsetka żywych komórek, zwiększenie stężenia do 25 μg/mL spowodowało, iż żywotność komórek linii DLD-1 i HT-29 została obniżona odpowiednio do 80% i 60%, natomiast w przypadku linii Caco-2 pozostała bez zmian. Zastosowanie 5-FU w formie enkapsulowanej wywarło zróżnicowany efekt cytotoksyczny na badane linie komórkowe. W przypadku linii DLD-1 oraz Caco-2, które są uznane jako oporne na leczenie 5-FU, zastosowanie nośników leków spowodowało zmniejszenie odsetka komórek żywych o ok. 40% w porównaniu do kontroli. Wzrost działania cytotoksycznego 5-FU zaaplikowanego w postaci nośnika zaobserwowano również w przypadku komórek HT-29, określanych jako średniowrażliwe na działanie 5-FU. W efekcie można stwierdzić, że oceniane komórki nowotworowe były znacznie bardziej wrażliwe na leczenie oparte na 5-FU w formie enkapsulowanej niż na lek w formie wolnej.

Lekooporność nowotworów wynika z wielu mechanizmów, do których można zaliczyć: mechanizmy związane z upośledzonym wychwytem/transportem leku. Obecnością pomp typu efflux i zmianami miejsc docelowych. W oparciu o antymetabolity, sposób działania indukuje cytotoksyczność poprzez zakłócanie biosyntezy kwasów nukleinowych. Analizując otrzymane wyniki, wskazujące na potencjał cytotoksyczny 5-FU w formie enkapsulowanej, założono, iż zastosowanie zsyntetyzowanych cząstek polimerowych sfunkcjonalizowanych kwasem foliowym może prowadzić do przezwyciężenia oporności na 5-FU. Powyższe może być osiągnięte poprzez interakcję nośników z receptorami dla FA. W celu lepszego wyjaśnienia proponowanego sposobu działania wykorzystano dwie linie komórkowe raka jelita grubego: DLD-1 i HT-29 [61, 63]. Wybrane linie komórkowe posiadają odmienne profile molekularne i genetyczne, w tym status RER i mutacje w obrębie TGFbIIR, a także zróżnicowany poziom ekspresji receptorów dla kwasu foliowego [64].

Ocenę wpływu zsyntetyzowanych nośników na aktywność metaboliczną i proliferację komórek nowotworowych przeprowadzono przy użyciu metod fluorescencyjnych, stosując test z resazuryną. Otrzymane wyniki wskazują, że cząstki polimerowe sfunkcjonalizowane FA znacząco hamują podział komórek w porównaniu do kontroli. W przypadku komórek linii DLD-1 po traktowaniu nośnikami z enkapsulowanym 5-FU, ponad 50% komórek wykazywało znacznie zmniejszoną aktywność metaboliczną, podczas gdy aplikacja 5-FU w formie wolnej skutkowała brakiem wpływu hamującego na proliferację komórek. Inkubacja

komórek linii HT-29 skutkowała podobną aktywnością inhibicyjną dla obu form leku (wolnej i enkapsulowanej). Należy podkreślić, że w obu leczonych liniach komórkowych zaobserwowano zahamowanie proliferacji komórek po zastosowaniu nośników leków bez enkapsulowanego leku, co sugeruje, że niektóre cytotoksyczne mechanizmy działania mogą być charakterystyczne dla nośników samych w sobie. Dodatkowe zwiększenie aktywności cytotoksycznej po zastosowaniu nośnika z enkapsulowanym cytostatykiem może wynikać z jego obecności bądź też wzajemnej interakcji zachodzącej pomiędzy nośnikiem a lekiem.

Przy wykorzystaniu testu multiplexowego zbadano także zdolność zsyntetyzowanych nośników do indukowania apoptozy i nekrozy. Po 24-godzinnej ekspozycji komórek jelita grubego linii DLD-1 i HT-29 na nośniki z i bez 5-FU oraz 5-FU w formie wolnej, przeprowadzono test bioluminescencyjny, który ocenia ekspozycję fosfatydyloseryny (ang. Phosphatidylserine, PS) na zewnętrznej błonie komórkowej, co jest charakterystycznym markerem podczas procesu apoptozy. Wyniki wykazały, że w przypadku komórek DLD-1 obciążone nośniki powodowały apoptozę na poziomie podobnym do 5-FU zastosowanego w stężeniu 20 μg/mL w formie wolnej. Traktowanie komórek HT-29 nośnikami polimerowymi z enkapsulowanym 5-FU powodowało 2-krotnie zwiększoną apoptozę w porównaniu do apoptozy odnotowanej w przypadku zaaplikowania cytostatyku w formie wolnej w stężeniu 20 μg/mL. W dalszym etapie w celu dokładniejszego określenia mechanizmu molekularnego zaangażowanego w obserwowaną aktywność cytotoksyczną zbadano czy i w jakim stopniu następuje indukcja procesu nekrozy w traktowanych komórkach. W tym celu zastosowano technikę opartą na pomiarze fluorescencji oceniającym stopień utraty integralności błony. Otrzymane wyniki wskazują, iż zsyntetyzowane nośniki, zarówno samodzielnie jak i z enkapsulowanym 5-FU indukują nekrozę w traktowanych komórkach około 2,5-krotnie skuteczniej niż lek w formie wolnej, niezależnie od zastosowanego stężenia (20 lub 100 μg/mL).

Farmakoekonomiczny aspekt zastosowania nośników leków w optymalizacji farmakoterapii chorób nowotworowych i racjonalizacji wydatków ponoszonych na leczenie, przeprowadzono w oparciu o dane pozyskane z baz danych takich jak PubMed, Embase czy Cochrane oraz z danymi pochodzącymi zarówno z polsko- jak i anglojęzycznych baz danych badań klinicznych takich jak INFARMA, Europejska Agencja Leków (ang. European Medicines Agency, EMA) czy U.S. National Library of Medicine. Został przeanalizowany wpływ terapii standardowej *vs* terapii opartej na rozwiązaniach z dziedziny nanotechnologii na perspektywy leczenia pacjentów (uwzględniając badania na poziomie in vitro, in vivo oraz kliniczne) ze zdiagnozowanym nowotworem, w tym rakiem jelita grubego.

Celem powyższej analizy było dostarczenie informacji o tym, która z dostępnych aktualnie technologii medycznych (terapia standardowa *vs* terapia oparta na nanotechnologii) pozwoli na uzyskanie najlepszych efektów zdrowotnych przy możliwie najniższych kosztach.

Z obecnie przeanalizowanych prac, które służyły napisaniu pierwszej oraz drugiej pracy wchodzących w skład niniejszej rozprawy doktorskiej wynika, iż:

- rozwiązania oparte na nanotechnologii, które są wykorzystywane w diagnostyce oraz leczeniu nowotworów są bardziej kosztoefektywne (redukcja kosztów przyszłego leczenia, poprawa *compliance* pacjenta, poprawa jakości i długości życia) w porównaniu ze standardową chemioterapią,
- nanoformulacje wykazują wysoką skuteczność terapeutyczną a także ograniczają działania niepożądane cytostatyków,
- zastosowanie nanonośników wiąże się z krótszym czasem hospitalizacji pacjenta, co ma również wpływ na koszty jego leczenia (np. bezpośrednie koszty medyczne, takie jak wykonane procedury medyczne czy opieka pielęgniarska),
- rozwiązania z dziedziny nanotechnologii umożliwiają zmniejszenie liczby działań niepożądanych, co może skutkować spadkiem liczby wykonywanych procedur medycznych, a także prowadzić do obniżenia kosztów osobowych, dawać większe szanse na remisję oraz umożliwić pacjentom powrót zarówno do życia prywatnego, jak i zawodowego.

Niewątpliwie do perspektyw dalszego rozwoju tematu należą:

- aspekt farmakoekonomiczny i nanomedyczny w innych typach badań, np. w badaniach mikrobiologicznych, badaniach obrazowych, szczepionkach [65-75],
- aspekt farmakoekonomiczny i nanomedyczny w konkretnych typach chorób, takich jak:

- cukrzyca: nieinwazyjne i bezbolesne monitorowanie poziomu glukozy we krwi [65-68],
- choroby sercowo-naczyniowe: inteligentne, nietoksyczne, biodegradowalne lub bioaktywne materiały i urządzenia stosowane do przeszczepów naczyniowych czy stentów uwalniających leki o zwiększonej biokompatybilności [65-67, 74],
- rozwinięcie tematu chorób nowotworowych: środki kontrastowe stosowane do skutecznego i wczesnego diagnozowania, prognozowania i monitorowania postępu choroby oraz do oceny ilościowej środków terapeutycznych i jednoczesnego obrazowania [65-67, 70-71],
- choroby neurodegeneracyjne (choroba Alzheimera, choroba Parkinsona): urządzenia diagnostyczne i biosensory oparte o nanosystemy; środki kontrastowe, konkurencyjne pod

względem ekonomicznym, stosowane do wzmocnienia konwencjonalnych metod obrazowania (MRI, obrazowanie molekularne, PET, itp.) [65-67, 72],
- choroby infekcyjne (HIV/AIDS): kontrolowane uwalnianie antybiotyków poprzez miejscowe dostarczanie leków, co prowadzi do bardziej efektywnego wykorzystania antybiotyków i kontroli dawki przyjętej przez pacjenta; narzędzia do diagnostyki przesiewowej o wysokiej skuteczności, z wykorzystaniem nanostruktur, o wysokiej czułości i krótkim czasie odpowiedzi [65-67, 73],
- inne choroby, np. choroba zwyrodnieniowa stawów, choroby skóry oraz leczenie ran: nanobiomateriały do unieruchamiania komórek macierzystych w miejscu urazu czy nanocząstki lipidowe do transportowania cząstek biologicznych w procesie gojenia ran [65-67, 74].

Analizując wartości kosztów jakie są ponoszone przez pacjentów onkologicznych, można zauważyć, że działania prewencyjne są najmniej kosztochłonne, droższa jest diagnostyka, zaś leczenie wymaga największych nakładów finansowych. Prawidłowe działania profilaktyczne, takie jak badania przesiewowe oraz kampanie edukacyjne skierowane nie tylko do osób obciążonych genetycznie, ale również do dzieci i młodzieży, umożliwiłyby wcześniejszą diagnostykę nowotworu. Powszechnie wiadomo, że im wcześniej nowotwór zostanie wykryty, tym większa jest szansa na jego wyleczenie. Pacjent, u którego późno zdiagnozowano nowotwór będzie miał mniejsze szanse na remisję, a jego leczenie będzie kosztochłonne. Co więcej, to wszystko przekłada się na aspekt farmakoekonomiczny ze względu na fakt, iż pacjent onkologiczny jest poniekąd obciążeniem dla systemu ochrony zdrowia, gdyż jego niemożność do wypełniania ról zawodowych i społecznych będzie bezpośrednio odzwierciedlona w kosztach pośrednich takich jak absencja chorobowa (zwolnienia lekarskie) [46].

Osiągnięcia nanomedycyny otwierają szerokie możliwości rozwoju spersonalizowanej terapii, w której wykrywanie raka, diagnostyka i terapia są dostosowane indywidualnie w zależności od molekularnego profilu nowotworu oraz cech osobniczych pacjenta [34, 76]. Udokumentowany spadek liczby działań niepożądanych, poprawa biodostępności czy wzrost *compliance* pacjentów, to tylko niektóre z licznych zalet jakimi charakteryzują się nanoformulacje. Zastosowanie powyższych rozwiązań znajduje również odzwierciedlenie w aspekcie farmakoekonomicznym. Zauważalne jest znaczne obniżenie kosztów bezpośrednich, czyli kosztów związanych z całym procesem leczenia (np. koszt sprzętu, leków, wykonanych badań diagnostycznych i procedur medycznych, jak również wynagrodzenia personelu

medycznego) oraz kosztów pośrednich, czyli czynności pozamedycznych (np. wyżywienie w szpitalu czy transport chorego) [37].

W związku z powyższym, niezbędne są dalsze, pogłębione badania oceniające toksyczność i bezpieczeństwo stosowania nanocząstek w terapii przeciwnowotworowej oraz szczegółowa analiza farmakoekonomiczna, która umożliwiłaby oszacowanie opłacalności i efektywności nowych rozwiązań [37, 65-67, 75].

Podsumowując, należy podkreślić, iż wciąż przed standardowymi schematami leczenia onkologicznego stoi wiele wyzwań, takich jak poszukiwanie nowych form leków oraz modyfikacja już obecnych, których potencjał terapeutyczny jest efektywny zarówno w aspekcie kosztów, jak i osiąganych efektów. Terapie celowane oparte na medycynie spersonalizowanej są kluczowymi i przyszłościowymi kierunkami badań. Niewątpliwie ich zastosowanie wiąże się z poprawą skuteczności leczenia oraz zmniejszeniem liczby działań niepożądanych. Wykorzystanie rozwiązań opartych na nanotechnologii w odniesieniu do onkologii ma na celu redukcję ujemnych aspektów chemioterapii w tym *compliance* pacjentów.

Rozdział 5. Wnioski

1. Nośniki na bazie kopolimerów akrylanu 2-hydroksyetylu (HEA) i N-izopropylokrylamidu (NIPAAm) sfunkcjonalizowane powierzchniowo ligandem nakierowującym, tj. cząsteczkami kwasu foliowego charakteryzują dobrze zdefiniowane właściwości fizykochemiczne i cechy morfologiczne, a także biozgodność z komórkami gospodarza, przez co są obiecującym materiałem do kreowania nanosystemów, których zadaniem jest selektywny transport leku oraz jego kontrolowane uwalnianie w miejscu docelowego działania.
2. Enkapsulacja 5-fluorouracylu w polimerowym nośniku w sposób istotny moduluje jego cytotoksyczność, poprawia kompatybilność cytostatyku w stosunku do komórek niezmienionych nowotworowo.
3. Zastosowanie systemów dostarczania 5-FU wykazuje działanie protekcyjne na komórki prawidłowe chroniąc przed szkodliwym działaniem cytostatyku, co w konsekwencji może przełożyć się na redukcję działań niepożądanych chemioterapii, takich jak np. immunosupresja szpiku kostnego związana z monocytopenią, ostra kardiotoksyczność oraz przewlekłe zapalenie błony śluzowej jelit.
4. Nośniki z enkapsulowanym 5-FU wykazują silniejsze działanie przeciwnowotworowe niż 5-FU zastosowany w formie wolnej oraz wykazują zdolność do uwrażliwienia komórek nowotworowych na 5-FU, pomimo potwierdzonej oporności na ten chemioterapeutyk.
5. Nanosystemy zawierające enkapsulowany chemioterapeutyk mają zdolność do indukcji apoptozy i nekrozy w traktowanych komórkach nowotworowych, w tym w komórkach charakteryzujących się fenotypem lekooporności.
6. W przypadku przełożenia otrzymanych wyników na warunki kliniczne, przyczyniłoby się to do zwiększenia efektywności terapii, poprawy jakości życia, a w efekcie do wydłużenia całkowitego czasu przeżycia pacjentów.
7. Wprowadzenie nowych substancji i produktów leczniczych na rynek farmaceutyczny takich jak nanosystemy, mogłoby przyczynić się do zapewnienia tańszej i efektywniejszej opieki medycznej, a wdrożenie metody leczenia opartej na nanotechnologii może być przyszłościowym rozwiązaniem umożliwiającym pokonanie ograniczeń konwencjonalnej terapii przeciwnowotworowej.

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Rozdział 7. Streszczenie w języku polskim

Rak jelita grubego jest trzecią najczęściej diagnozowaną postacią nowotworu. Obecnie ten nowotwór jest drugim nowotworem w Polsce pod względem występowania i śmiertelności, częstszy jest tylko rak płuca u mężczyzn i rak piersi u kobiet. Co roku jest diagnozowany u ponad 18 tys. osób, z czego 11 tys. pacjentów umiera.

Nanotechnologia jest interdyscyplinarną, nowoczesną i przyszłościową dziedziną wiedzy przez co znalazła zastosowanie w wielu obszarach, w tym w medycynie. Farmakoekonomika jest częścią ekonomiki zdrowia, której głównym celem jest ocena zastosowanej farmakoterapii z uwzględnieniem perspektywy ekonomicznej. Podobnie jak nanotechnologia jest zaliczana do dziedzin interdyscyplinarnych poprzez łączenie ze sobą nauk, medycznych i około medycznych, takich jak: medycyna, farmakologia, statystyka medyczna czy też ekonomia. Takie połączenie umożliwia obiektywną ocenę i analizę opłacalności zastosowanych środków farmakologicznych. Służy to określeniu zależności pomiędzy kosztem leczenia a osiągniętym dzięki niemu efektom zdrowotnym.

Celem niniejszej rozprawy doktorskiej było rozpoznanie wpływu zastosowania rozwiązań opartych na dynamicznie rozwijającej się dziedzinie jaką jest nanotechnologia na aspekty doświadczalne i farmakoekonomiczne leczenia nowotworu jelita grubego z wykorzystaniem nowych inteligentnych nośników leków na bazie polimerów termowrażliwych.

Niniejsza rozprawa doktorska powstała w oparciu o cykl spójnie tematycznych publikacji, z których pierwsza jest pracą przeglądową odzwierciedlającą nowe możliwości oraz wyzwania leczenia nowotworu jelita grubego z wykorzystaniem nanotechnologii oraz medycyny spersonalizowanej, jak również przybliżeniem aspektów farmakoekonomicznych takich rozwiązań, czyli kosztów towarzyszących całemu procesowi leczenia. Druga praca jest pracą oryginalną, w której zostały ocenione możliwości zastosowania nowych nośników polimerowych sfunkcjonalizowanych kwasem foliowym do ukierunkowanego dostarczania 5-fluorouracylu (5-FU) w terapii nowotworów jelita grubego w modelu in vitro.

Badania wykazały, iż enkapsulacja 5-fluorouracylu w polimerowym nośniku w sposób istotny moduluje jego cytotoksyczność, poprawia kompatybilność cytostatyku w stosunku do komórek niezmienionych nowotworowo. Powyższe skłania do wysnucia hipotezy, iż zastosowanie systemów dostarczania 5-FU wykazuje działanie protekcyjne na komórki

prawidłowe chroniąc przed szkodliwym działaniem cytostatyku, co w konsekwencji może przełożyć się na redukcję działań niepożądanych chemioterapii, takich jak np. immunosupresja szpiku kostnego związana z monocytopenią, ostra kardiotoksyczność oraz przewlekłe zapalenie błony śluzowej jelit. Z kolei, podczas oceny przeżywalności komórek zmienionych nowotworowo po zastosowaniu zsyntetyzowanych nanosystemów wykazano silniejsze działanie przeciwnowotworowe niż 5-FU zastosowany w formie wolnej oraz zdolność do uwrażliwienia komórek nowotworowych na 5-FU, pomimo potwierdzonej oporności na ten chemioterapeutyk. Ponadto, otrzymane nanosystemy zawierające enkapsulowany chemioterapeutyk mają zdolność do indukcji apoptozy i nekrozy w traktowanych komórkach nowotworowych, w tym w komórkach charakteryzujących się fenotypem lekooporności.

Podsumowując, wdrożenie nowych form leków na rynek farmaceutyczny z wykorzystaniem rozwiązań z dziedziny nanotechnologii oraz leczenie pacjentów w oparciu o terapię spersonalizowaną może w konsekwencji wpłynąć na obniżenie kosztów bezpośrednich i pośrednich procesu leczenia poprzez zwiększenie efektywności terapii oraz poprawę *compliance* pacjentów, u których zdiagnozowano nowotwór.

Rozdział 8. Streszczenie w języku angielskim

Colorectal cancer is the third most frequently diagnosed form of cancer. Currently, colorectal cancer is the second most common cancer in Poland in terms of incidence and mortality, with only lung cancer in men and breast cancer in women being more common. Every year it is diagnosed in more than 18,000 people, of whom 11,000 patients die.

Nanotechnology is an interdisciplinary, modern and forward-looking field of knowledge and due to this might be applied in many areas, including medicine. Pharmacoeconomics is a part of health economics, and the main purpose is to evaluate applied pharmacotherapy from an economic perspective. Like nanotechnology, it is categorized as an interdisciplinary field by combining sciences, medical and peri-medical, such as medicine, pharmacology, medical statistics or economics. The aforementioned combination makes it possible to objectively evaluate and analyze the cost-effectiveness of the pharmacological agents used. This serves to determine the relationship between the cost of treatment and the health effects achieved through it.

The purpose of this doctoral thesis was to recognize the impact of the use of solutions based on the dynamically developing field which is nanotechnology on the experimental and pharmacoeconomic aspects of colorectal cancer treatment using new smart drug carriers based on thermoresponsive polymers.

This dissertation is based on a series of coherently thematic publications, the first of which is a review paper reflecting the new opportunities and challenges of colorectal cancer treatment using nanotechnology and personalized medicine, as well as an approximation of the pharmacoeconomic aspects of such solutions, for instance the costs accompanying the entire treatment process. The second paper is an original work, in which the possibilities of using novel polymeric carriers functionalized with folic acid for targeted delivery of 5-fluorouracil (5-FU) in the treatment of colorectal cancer in an in vitro model were evaluated.

Studies have shown that encapsulation of 5-fluorouracil in a polymeric carrier significantly modulates its cytotoxicity via improving the compatibility of the cytostatic agents against non-tumorigenic cells. The above leads to the hypothesis that the use of 5-FU delivery systems exhibits a protective effect on normal cells, which might consequently translate into a reduction of chemotherapy side effects, such as bone marrow immunosuppression associated with monocytopenia, acute cardiotoxicity and chronic intestinal mucositis. In turn, the evaluation of the survival of neopastic cells after application of the synthesized nanosystems showed a more potent antitumor effect than 5-FU applied in

free form, as well as the ability to sensitize tumor cells to 5-FU, despite confirmed resistance to this chemotherapeutic. In addition, the obtained nanosystems containing the encapsulated chemotherapeutic agent possess the ability to induce apoptosis and necrosis in treated cancer cells, including those characterized by a drug-resistant phenotype.

In conclusion, the implementation of new forms of drugs into the pharmaceutical market using nanotechnology solutions and the treatment of patients based on personalized therapy might reduce the direct and indirect costs of the treatment process by increasing the efficiency of therapy and improving the compliance of patients diagnosed with cancer.

Rozdział 9. Current Trends and Challenges in Pharmacoeconomic Aspects of Nanocarriers as Drug Delivery Systems for Cancer Treatment

Milewska Sylwia, Niemirowicz-Laskowska Katarzyna, Siemiaszko Gabriela, Nowicki Piotr, Wilczewska Agnieszka Zofia, Car Halina: Current Trends and Challenges in Pharmacoeconomic Aspects of Nanocarriers as Drug Delivery Systems for Cancer Treatment. International Journal of Nanomedicine, 2021: 16, s. 6593-6644, DOI: 10.2147/IJN.S323831, IF: **7.033**, MNiSW/MEiN: **140.0**. Cytowania: **10/12** (Web of Science/Scopus).

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REVIEW

Current Trends and Challenges in Pharmacoeconomic Aspects of Nanocarriers as Drug Delivery Systems for Cancer Treatment

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Abstract: Nanotherapy is a part of nanomedicine that involves nanoparticles as carriers to deliver drugs to target locations. This novel targeting approach has been found to resolve various problems, especially those associated with cancer treatment. In nanotherapy, the carrier plays a crucial role in handling many of the existing challenges, including drug protection before early-stage degradations of active substances, allowing them to reach targeted cells and overcome cell resistance mechanisms. The present review comprises the following sections: the first part presents the introduction of pharmacoeconomics as a branch of healthcare economics, the second part covers various beneficial aspects of the use of nanocarriers for in vitro, in vivo, and pre- and clinical studies, as well as discussion on drug resistance problem and present solutions to overcome it. In the third part, progress in drug manufacturing and optimization of the process of nanoparticle synthesis were discussed. Finally, pharmacokinetic and toxicological properties of nanoformulations due to up-to-date studies were summarized. In this review, the most recent developments in the field of nanotechnology's economic impact, particularly beneficial applications in medicine were presented. Primarily focus on cancer treatment, but also discussion on other fields of application, which are strongly associated with cancer epidemiology and treatment, was made. In addition, the current limitations of nanomedicine and its huge potential to improve

and develop the health care system were presented.

Keywords: nanotechnology, pharmacology, pharmacoeconomic analysis, pharmacoeconomics, nanomaterials synthesis, clinical trials

Introduction

Nowadays, increasing evidence indicates that nanomedicine might have revolutionized therapeutic and diagnostic procedures, especially cancer treatment. This new technology provides a new toolset impacting the prevention of diseases by applying novel molecular diagnostic disease markers, early diagnosis of the neoplastic lesions in molecular imaging, and the treatment by enabling precise and effective therapies based on a personalized medication regimen.[1,2] Furthermore, there is evidence suggesting that combining nanomedicine with pharmacoeconomic evaluations could help reduce costs in managing cancer patients, for instance, by shortening the time of hospitalization or bringing down the number of necessary tests to be carried out. Another important fact worth mentioning is that the efficacy of drugs used with nanocarriers may substantially reduce cytotoxicity, preventing the occurrence of side effects by dose reduction and lower accumulation of therapeutic

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compounds in healthy body sites.[3,4] The considerations above provide a sound basis for holding nanotechnology in future medical developments capable of delivering highly efficacious and safe products. These new approaches should be available at reasonable costs and help restrict healthcare costs while maintaining clinical efficacy.[3,5]

From a pharmacoeconomic point of view, the development of new drug substances and products such as nanosystems and their introduction into the pharmaceutical market could contribute to more affordable care. Specifically, the potential for reducing adverse events plays a significant role in new encapsulated therapeutics, which results in fewer medical procedures and leads to the reduction in personnel costs. It also gives greater chances of remission and allows patients to return to professional life.[5,6]

Moreover, it should be emphasized that the application of nanotechnology in the medical field has many advantages since nanoparticles make a significant contribution as drug delivery systems due to their unique properties like the small size and large surface area.[7–9] The nanoformulation of drugs increases efficacy by enhancing the drug's cellular uptake in the cellular targets; hence, it achieves better biodistribution. Nanosized formulations, in comparison with conventional forms of drugs, exhibit better control of drug release kinetics, which lead to an increased active concentration and bioavailability. Another important factor is that the nanodrugs could induce a marked suppression of tumor growth, prolongation of total survival time in cancer patients, and targeted delivery, which might enhance cytotoxic effects on neoplastic cells and restrict adverse effects in the whole body.[10,11] All the above advantages make nanotechnology much cheaper than conventional therapies, which can also be reflected in the pharmacoeconomic aspect as the reduction or total avoidance of costs associated with medical (hospitalization, medical devices, monitoring therapy), and non-medical procedures (accommodation, transportation or the informal care).

It is worth noting that a broad literature review was undertaken. This paper presents existing evidence available regarding the effectiveness and expected pharmacoeconomic benefits of the alternative options of commonly used chemotherapeutic drugs to treat different types of cancers. Some factors may influence the results of the treatment regimen applied, such as patients' age, stage of the disease, therapy onset, benefit duration, and also time to recurrence. Pharmacoeconomic analyses of alternative therapy options will improve decision-making and will help to optimize the use of already limited health care resources allocated to the care of cancer patients.[12] This paper aims to identify potential benefits from applying pharmacoeconomic to the rapidly evolving area of nanotechnology, especially in the domain of drug development for cancer treatment, which is presented in Figure 1.

Pharmacoeconomics – a Use Case of Nanocarriers Evaluation

Pharmacoeconomics is considered as a branch of health economics, which identifies, measures, and compares the costs and consequences of drug therapy for healthcare systems and society.[13–15] Moreover, it provides essential guidance on the management of limited healthcare resources and medical practice. Given the limited financial resources, health economics, particularly pharmacoeconomic analysis, is becoming a frequently used criterion for decision-making in modern healthcare policy.[16,17] Therefore, searching for novel therapeutic options characterized by high efficacy with restricted side effects remains a highly desired goal.[18]

Pharmacoeconomics applies the principles of health economics to the field of pharmaceutical policy. Also, it uses a broad range of techniques for health economic evaluation in the specific context of drug management.[19,20] In effect, the introduction of novel forms of drugs, such as those encapsulated in carriers, lies in pharmacoeconomic purposes.

If think about conducting a pharmacoeconomic analysis, we should follow a clearly defined stepwise approach: a) Define the pharmacoeconomic problem – we should state the problem and select the objectives; b) Identify the perspective of the study – the most popular are: patients, provider, payer, and society; c) Identify the relevant interventions – we need to answer a significant question: "Have all relevant interventions been identified (including non-drug interventions)?"; Use decision trees or treatment models; d) Select the appropriate pharmacoeconomic method – CEA, CMA, CUA, CBA; e) Select the primary data source and study design – retrospective/prospective clinical trial data, economic (naturalistic) trial data; f) Select the secondary data sources – such as databases, literature, clinical expertise; g) Select appropriate analysis technique – modeling, meta-analysis; h) Identify the measures and the outcomes of alternative

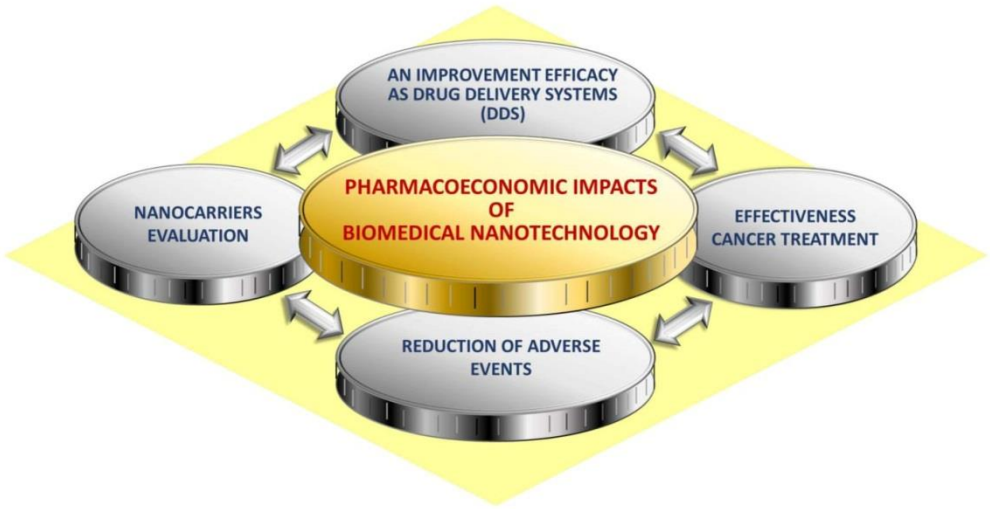

Figure 1 Pharmacoeconomic impacts in biomedical nanotechnology.

interventions – health outcomes and resource outcomes for beneficial as well as adverse effects; i) Use analytical methods – to establish the probability of outcome events and to answer the research question – such as efficacy rates, the incidence of adverse drug reactions, and decision trees; j) Estimate costs and effectiveness – reduce costs and outcomes; perform incremental cost analysis; k) Perform sensitivity analysis – determine the effect of varying uncertain variables over a range of results/assumptions; l) Interpret and present results – describe assumptions, methods, data sources; study limitations including significant omissions stated; interpret results.[21]

There are four most popular analyses to estimate the outcomes, and each of the methods is associated with a different type of pharmacoeconomic analysis, see Table 1.

In pharmacoeconomic analysis, costs are crucial elements that should be taken into consideration. They can be classified as direct (medical and non-medical), indirect and intangible costs. Financial costs relate to monetary payments associated with the price of a good or service traded on the market. Economic costs match the broader concept of resource consumption, irrespective of whether such resources are traded in the market.[13,24] In Table 2, we summarize and specify the types of costs that are considered in pharmacoeconomics. These costs together with the expected pharmacoeconomic efficacy measure when applying nanocarriers in cancer treatment are shown in Table 3.

For any pharmacoeconomic analysis, the perspective is critical since it determines what costs and benefits will be measured: 1. Societal – all costs and consequences that occurred during the treatment, 2. Third-party payer–payers are represented by insurance companies, employers, or the government; the direct costs are included, but also indirect costs can also be included, 3. Hospital/physician (healthcare providers) – providers include hospitals, private-practice physicians, or managed-care organizations; from this perspective, direct medical costs are included, 4. Patient – all costs borne by the patient for any product or services and are not covered by any insurance; there are direct, indirect, and intangible costs (out of pocket). According to the aforementioned, those costs/analysis should be taken into consideration if we are thinking about the safe application of nanocarriers in modern therapy.[13,25]

The Cancer Burden in the World

The National Cancer Institute defines cancer as a set of diseases in which abnormal cells divide without control and can spread to various tissues. Cancers can manifest in different parts of the body – leading to a range of different cancer types.[26] Based on the available data, it is assessed that cancer is one of the leading causes of death. In 2018, 9.6 million people were estimated to have died of various

Table 1 Types of Pharmacoeconomic Studies.[21-24]

Pharmacoeconomic Study	Description	Use Case	Example
Cost-minimization analysis (CMA)	**To select the least costly among several similar interventions.** • Applied when there is a need to compare multiple drugs of equal efficacy and equal tolerability. • Is performed when the outcomes are the same for the two interventions. • It cannot be used to evaluate programs or therapies that lead to different outcomes.	To identify the least costly option when outcomes/consequences are identical.	Compare costs of Drug A and Drug B (equal efficacy for a given condition and safety).
Cost-benefit analysis (CBA)	**To identify, measure, and compare the benefits and costs of a program or treatment alternative.** • The most comprehensive and the most difficult of all economic evaluation techniques. The benefits are assigned a monetary value so that costs and benefits can be easily compared. Different interventions can be compared - is a useful tool (like CUA) for resource allocation by policy-makers. • CBA should be employed when comparing treatment alternatives in which the costs and benefits do not occur at the same time. • Can be used to compare programs with different objectives - because all benefits are converted into currency and to evaluate a single program or compare various programs.	To compare programs or agents with different objectives or one program against a return on investment benchmark.	Clinical pharmacy service vs another institutional service.
Cost-effectiveness analysis (CEA)	**To assist decision-makers in identifying the preferred choice among possible alternatives.** • Used to evaluate multiple drug interventions for the same condition. The cost of drug treatments is weighed against the effectiveness of the drug. • The costs of drug treatments consist of acquisition costs, physician engagement, and nursing costs for administration of the drug. • The effectiveness of drug treatment is measured by the duration of treatment, length of hospital stay, and mortality rate. • The key measure of these evaluations is the incremental cost-effectiveness ratio (ICER).	To compare treatment alternatives for a given condition that differ in outcomes and costs.	Osteoporosis: Drug A vs Drug B on fracture risk reduction ($/fractures avoided).
Cost-utility analysis (CUA)	**To compare medications or interventions with different outcomes.** • Compare cost, quality, and the number of patient-years. • Used when programs and treatment alternatives should be compared. • CUA is **applied less frequently** than other economic evaluations because of the lack of standardization of measurement utilities, eg, difficulty comparing QALYs (quality-adjusted life-years) across patients and populations and difficulty quantifying patient preferences.	The same as CEA, useful when treatment extends life and/or affects the quality of life.	Compare cancer chemotherapy regimens.

Table 2 Type of Costs in Pharmacoeconomic Analysis.[13,22,24]

Costs		
Direct	**Medical**	1[a]. Hospitalization 1[b]. Outpatient visits (to primary care providers) 1[c]. Procedures and tests (laboratory tests, surgical interventions, USG) 1[d]. Medical devices 1[e]. Homecare 1[f]. Nursing care 1[g]. Medications 1[h]. Monitoring therapy 1[i]. Adverse events management 1[j]. Medical staff costs 1[k]. Administrative costs
	Non-medical	2[a]. Accommodation 2[b]. Transportation 2[c]. Non-medical services (home helper, meals on wheels, social assistance) 2[d]. Devices and investments 2[e]. The informal care
Indirect		3[a]. Sick leave or absences (short term disability) 3[b]. Reduced productivity at work (productivity loss) 3[c]. Early retirement due to illness (long term disability) 3[d]. The premature death
Intangible		Costs which are difficult to assess; a patient or their family might feel: 4[a]. Anxiety 4[b]. Pain 4[c]. Suffering

forms of cancer. Globally, WHO roughly estimates that 1 in 6 deaths is due to cancer. Considering the income – approximately 70% of deaths from cancer occur in low- and middle-income countries. The most common cancers, in terms of frequency and number of deaths, are lung, breast, and colorectal.[27]

Cancer burden is associated with risk factors belonging to three main groups, which are: socio-economical, lifestyle, and genetic/health predisposition comprising prolonging and chronic inflammation caused by the existence of microbial infections. Besides the fact that microbes might induce chronic inflammation, it was evidenced that they are able to produce carcinogenic bacterial metabolites, which caused mutation of genetic materials.[28] It means that disturbance in one of these groups triggers a cascade of processes leading to the development of cancer. Researchers have found several risk factors that may increase the chance of getting lung, breast, and colorectal cancer (Figure 2).

In the case of lung cancer, the number one risk factor is smoking. People who smoke cigarettes are about 15, even up to 30 times more likely to get or die of lung cancer than people who do not smoke. Smoking only a few cigarettes a day or occasionally increases the risk of developing lung cancer. The longer a person smokes, and the more cigarettes are smoked each day, the more risk becomes apparent. It is a misleading belief that smoking can only cause lung cancer. Smoking also causes several other neoplasms, such as cancer of the mouth and throat, esophagus, stomach, colon and rectum, liver, pancreas, kidney, urinary bladder, and even acute myeloid leukemia.[29] Moreover, it should be emphasized that tuberculosis, pneumonia, and chronic bronchitis are examples of pathology, which have a profound role in the emergence of cancer. In effect, in the case of lung cancer, prolonging microbial infections are major inflammation-inducing factors, which is known to be the cause of cancer development.[30]

Risk factors for breast cancer can be divided into modifiable and non-modifiable.[31] To have a lower risk of getting breast cancer, every woman should be physically active and keep the body weight normal, if possible – avoid taking contraceptives and hormone replacement therapy, have the first pregnancy before age 30, breastfeed, and have a full-term pregnancy. Smoking, being exposed to chemicals, drinking alcohol, and having changes in other hormones due to night shift working may also increase breast cancer risk.[31] Non-modifiable risk factors include age, genetic mutations, reproductive history, dense breasts, personal and family history of breast cancer, and previous treatment using radiotherapy. Important is the fact that there is evidence linking chronic inflammation, which might be caused by microbial infection, to breast cancer risk, development and progression.[32] For instance, it is established that breast cancer was one example of among other 15 incident cancer, in which the risk of developing one year after *Staphylococcus aureus* bacteremia (SAB) was significantly increased compared to the general population.[33] Screening for this aspect in cancers in populations with developed SAB infection might allow for earlier disease detection. Additionally, the presence of chronic infection also affects the human microbiota. Recent studies have found that people who have a good response to immunotherapy to treat their cancer appear to

Table 3 Efficacy of Selected Drugs and Expected Pharmacoeconomic Benefits Due to Their Nanoformulations.[115,140-167]

Nanocarrier		Efficacy	Expected Pharmaco-Economic Efficacy*
NP	**Type of NPs**	**Doxorubicin**	
Chitosan-dextran conjugate NPs	P	Reduction of tumor size; Prolongation of survival.	1^a-1^k,2^a-2^e 3^a-3^d,4^a-4^c
Dox-loaded chitosan NPs	P	Marked inhibition of tumor growth; Prolongation of survival.	1^a-1^k,2^a-2^e 3^a-3^d,4^a-4^c
Dendrimer-Dox conjugates NPs	P	A single dose can cure mice with *s.c.* implanted colon cancer; The 100% survival of the tumor-bearing mice; A lower weight loss.	1^a-1^k,2^a-2^e 3^a-3^d,4^a-4^c
Peptide-Dox conjugates NPs	Pp	Marked inhibition of tumor cells in vitro; An effectiveness against a Dox-resistant neuroblastoma cell line.	1^a-1^k,2^a-2^e 3^a-3^d,4^a-4^c
Dox-loaded polymeric NPs	P	The greatest inhibition of primary human liver tumors implanted *s.c.*; Reduction in tumor growth; greater tumor inhibition and tumor necrosis; A marked reduction in the tumor collagen levels; A little to no toxicity to the mice.	1^a-1^k,2^a-2^e 3^a-3^d,4^a-4^c
PCMB-Dox NPs	P	Prolongation of survival; Suppression of tumor growth by about 80%; No toxic effects evidenced by histology, blood chemistries, and body weight.	1^a-1^k,2^a-2^e 3^a-3^d,4^a-4^c
Dox-loaded exosomes	E	Significant inhibition of tumors; No cardiotoxicity.	1^a-1^k,2^a-2^e 3^a-3^d.4^a-4^c
Paramagnetic NPs	M	A greater killing of cancer cells.	1^a-1^k,2^a-2^e 3^a-3^d.4^a-4^c
		5-Fluorouracil	
SLNs	L	An improvement of the uptake of anticancer drugs inside colon tumors; Superior anticancer activity; Enhanced cytotoxic effects.	1^a-1^k,2^a-2^e 3^a-3^d,4^a-4^c
Chitosan-based NPs	P	Minimization of the toxic effects on healthy cells; An improvement of localization of the drug at the colon region; A decrease in drug-induced toxicity; A reduction of dose frequency and drug administration; A provision of better targeting efficiency and the accumulation of the drug.	1^a-1^k,2^a-2^e 3^a-3^d,4^a-4^c
PLGA NPs	P	The rate of cell lysis was about 80%; A prominent exhibition of an effect on target CRC cells.	1^a-1^k,2^a-2^e 3^a-3^d,4^a-4^c
FA and PLGA conjugates	P	An enhancement of anticancer activity; The lowest cell viability.	1^a-1^k,2^a-2^e 3^a-3^d,4^a-4^c
Eudragit S100 coated CPNs	P	A better targeting efficiency; An exhibition of drug release over a prolonged period.	1^a-1^k,2^a-2^e 3^a-3^d,4^a-4^c
SiNPs	M	An enhancement of cellular uptake; An improvement of cytotoxic effects.	1^a-1^k,2^a-2^e, 3^a-3^d,4^a-4^c

		Paclitaxel	
PLGA-NPs	P	Minimal systemic toxicity; Significantly better tumor growth inhibition effect with transplantable liver tumors; Facilitation of drug cell uptake; An increase in cellular association; An enhancement of cytotoxicity; Inhibition of intimal proliferation in a rabbit vascular injury model; A significant prolongation of survival; Improvement of drug encapsulation efficiency; Better control of drug release kinetics; An enhancement of cellular uptake; Better antitumor efficacy.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}

(*Continued*)

Table 3 (Continued).

Nanocarrier		Efficacy	Expected Pharmaco-Economic Efficacy*
NP	**Type of NPs**	**Doxorubicin**	
PLA NPs	P	Significant antitumor efficacy; More drug accumulation in tumors.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}
PCL NPs	P	An enhancement of cytotoxicity; A remarkable tumor growth inhibition; An enhancement of antitumor efficacy; No acute toxicity; An increase in cellular uptake; An enhancement of toxicity; An improvement of tumor inhibitory activity.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}
PEG-PCL NPs	P	An improvement of the pharmacokinetic profile; An increase in the mean survival time; Better drug loading profile; An improvement in entrapment efficiency to 98%; Significantly greater tumor accumulation.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}
PVP-*b*-PCL or PCL-*g*-PVA	P	Significantly superior antitumor efficacy; An exhibition in reduction of drug release rate profiles; Better antitumor activity.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}
HO-GC	P	Faster cellular uptake; Better therapeutic efficacy; An enhancement of the aqueous solubility; Achievement of a higher drug loading up to 20%; Achievement of maximum entrapment efficiency of 97%.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}
mPEG-CHO-chitosan NPs	P	Significantly slower tumor growth rate; An improvement of life span.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}
LyP-1-Abraxane NPs	Pp	A significant improvement of antitumor efficacy.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}
BSA NPs	Pp	High stability; Surface properties which specifically targeted to human prostate cancer cells.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}
OSA NPs	Pp	An improvement of the lipophilicity of albumin; Higher drug entrapment efficiency; Greater stability.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}
HA-NPs	P	A superior antitumor efficacy; An achievement of the drug loading up to 20.7%.	1^{a}-1^{k}, 2^{a}-2^{e} 3^{a}-3^{d}, 4^{a}-4^{c}
PBCA-NPs, (HA)-PBCA-NPs	P	A gradual drug release up to 80% within 96 h; Reduction of the initial burst release of the drug; A decrease in the cytotoxicity; An enhancement by cellular uptake; More potent antitumor inhibition activity.	1^{a}-1^{k}, 2^{a}-2^{e} 3^{a}-3^{d}, 4^{a}-4^{c}
HPG-C10-PEG, PEI-C18-HPG	P	Drug release up to 80%; Better tolerance; A significant exhibition and improvement of antitumor efficacy; A decrease in cytotoxicity.	1^{a}-1^{k}, 2^{a}-2^{e} 3^{a}-3^{d}, 4^{a}-4^{c}
PEG-PE NPs	P	Better antitumor activity; An improvement of antitumor efficacy.	1^{a}-1^{k}, 2^{a}-2^{e} 3^{a}-3^{d}, 4^{a}-4^{c}
Gelatin NPs	P	A significant improvement of antitumor activity.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}
NK 105	P	A significant better antitumor efficacy; Dramatically lower neurocytotoxicity.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}

Liposomes	L	A significant better antitumor efficacy; Greater tumor uptake; Reduction of toxicity; Significantly smaller tumor volumes; Inhibition of metastasis.	1[a]-1[k],2[a]-2[e] 3[a]-3[d],4[a]-4[c]
SLNs	L	Increased cellular uptake; Optimization of the drug entrapment efficiency; A significant enhancement of toxicity; An increase in brain uptake; Slower tumor growth rate; Potential to overcome P-gp-mediated MDR.	1[a]-1[k],2[a]-2[e] 3[a]-3[d],4[a]-4[c]

(*Continued*)

Table 3 (Continued).

Nanocarrier		Efficacy	Expected Pharmaco-Economic Efficacy*
NP	**Type of NPs**	**Doxorubicin**	
Lipid Nanocapsules	L	A significant increase in the life span; Potential to overcome P-gp-mediated MDR; An increase in drug cell uptake and retention; An increase in drug loading and entrapment efficiency; Prolonged and sustained in vitro release; An exhibition of better antitumor efficacy.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}
PTX Fatty Acid-Prodrug Lipid-Based NPs	L	Tumor growth inhibition; Antitumor activity; Less toxic; A significant improvement of drug loading efficiency; A superior anti-tumor efficacy.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}
Micro- and Nano-Emulsions	L	Much better tolerance; A significant improvement of antitumor efficacy; An increase in the life span; An extended release; Greater bioavailability.	1^{a}-1^{k},2^{a}-2^{e} 3^{a}-3^{d},4^{a}-4^{c}
Drug-Polymer Conjugates	P	A significantly better antitumor efficacy; A remarkable enhancement of tumor inhibitory activity; Low toxicity; Superior antitumor activity; Complete elimination of tumors (in some cases); Prolonged circulation time.	1^{a}-1^{k}, 2^{a}-2^{e} 3^{a}-3^{d}, 4^{a}-4^{c}
MNPs	M	An enhancement of cell inhibition activity; Low toxicity.	1^{a}-1^{k}, 2^{a}-2^{e} 3^{a}-3^{d}, 4^{a}-4^{c}
CNTs	C	A significant improvement of antitumor activity; An increase in drug loading; A significant increase in cell death; Non-toxicity.	1^{a}-1^{k}, 2^{a}-2^{e} 3^{a}-3^{d}, 4^{a}-4^{c}
CD NPs	P	Low haemolysis and cytotoxicity.	1^{a}-1^{k}, 2^{a}-2^{e} 3^{a}-3^{d}, 4^{a}-4^{c}
Nanogel	P	A significant improvement of antitumor efficacy.	1^{a}-1^{k}, 2^{a}-2^{e} 3^{a}-3^{d}, 4^{a}-4^{c}
ANG 1005	Pp	Better antitumor efficacy; An increase in survival time.	1^{a}-1^{k}, 2^{a}-2^{e} 3^{a}-3^{d}, 4^{a}-4^{c}

Note: *Based on Table 2.

Abbreviations: ABCB1 gene, ATP Binding Cassette Subfamily B Member 1; ANG 1005, Angiopep-2 Paclitaxel Conjugate; BSA, Bovine Serum Albumin; C, carbon-based nanoparticles; CD NPs, Cyclodextrin Nanoparticles; CNTs, Carbon Nanotubes; CRC, Colorectal Cancer; Dox, Doxorubicin; E, exosomes-based nanoparticles; Eudragit S100, Anionic Copolymers based on Methacrylic Acid and Methyl Methacrylate; HA, Hyaluronic Acid; (HA)-PBCA-NPs, Hyaluronic acid coated poly(butyl cyanoacrylate) nanoparticles; HO-GC, Hydrotropic Oligomer-Conjugated Glycol Chitosan; HPG, Hyperbranched Polyglycerol; HPG-C10-PEG, Hyperbranched polyglycerol-C10- poly (ethylene glycol); *i.v.*, intra venosa; L, lipid-based nanoparticles; LyP, 1-Abraxane - type of peptide; M, metallic-based nanoparticles; MNPs, Magnetic NPs; MDR, Multidrug Resistance; mPEG-CHO-chitosan NPs, Methoxypoly(ethylene glycol) conjugated Chitosan Nanoparticles; NK 105, Paclitaxel-incorporating Micellar Nanoparticle Formulation; PEG, polyaspartate micellar NPs; NPs, Nanoparticles; *s.c.*, subcutaneously; OSA, Octyl-modified bovine Serum Albumin; SiNP, Silica Nanoparticles; SLNs, Solid Lipid Nanoparticles; P, polymeric-based nanoparticles; PBCA, Poly(butyl cyanoacrylate); PCL, Poly(ε-caprolactone); PCMB-Dox NPs, PEGylated Carborane-Conjugated Amphiphilic Copolymer Doxorubicin Nanoparticles; PEI-C18-HPG, Polyethyleneimine (PEI)-C18-HPG; PEG-PCL, Poly(ethylene glycol)-Poly(ε-caprolactone); PEG-PE Nps, Poly(Ethylene Glycol)-Phosphatidyl Ethanolamine Nanoparticles; P-gp, Permeability glycoprotein; PLA, Polylactide; PLGA, Poly(lactic-*co*-glycolic acid); PLD, PEGylated Liposomal Doxorubicin; PLMB-Dox NPs, Doxorubicin-Loaded Carborane-Conjugated Polymeric Nanoparticles; Pp, peptide-based nanoparticles.

have a different microbiome composition than those who do not respond that well.[34]

The risk of getting colorectal cancer increases as the patient gets older.[36] About 90% of cases occur in people in their 50s or older. Other risk factors include inflammatory bowel disease (Crohn's disease, ulcerative colitis), a personal or family history of colorectal cancer or colorectal polyps, and a genetic syndrome, such as hereditary non-polyposis colorectal cancer (Lynch syndrome).[36] Moreover, in recent decades, there has been accumulating information in the published literature about the link between CRC and microbial infection. It has been announced that both viruses and bacteria can cause CRC via prolonged infection and accompanying inflammation, as well as induction of mutagenesis that leads to uncontrolled epithelial cell proliferation. Based on data from clinical and laboratory trials, among the aforementioned microbial agents, a crucial role was noted for

Figure 2 The burden of cancer: risk factors and the frequency of diagnosed cases and deaths (in the center[35]).

Streptococcus bovis, Helicobacter pylori, Escherichia coli, Klebsiella pneumoniae, and *Fusobacterium*.[37]

It should be kept in mind that lifestyle factors may also contribute to an increased risk of colorectal cancer, such as lack of regular physical activity, low amount of fruit and vegetables in the diet, a low-fiber and high-fat diet or a diet high in processed meat, overweight and obesity, alcohol consumption and tobacco use.[36]

Noteworthy is the fact that between 30% and 50% of cancers can be prevented by avoiding risk factors and implementing existing evidence-based prevention strategies. Cancer burden can also be reduced by early detection of cancer and the management of patients who develop cancer. Many cancers are curable if they are diagnosed early and treated properly.[38] Additionally, it should be emphasized that inflammation is often associated with cancer development and progression.[39] The triggering of chronic inflammation that increases cancer risk includes bacterial infections. In effect, the application of nanotechnology products that possess proved antimicrobial properties might have important implications for cancer preventions (Table 4).

Adequate prevention measures and early detection and treatment might substantially reduce cancer mortality rate. There are two components for efficient detection: 1. Early diagnosis – cancer that is diagnosed at an early stage, when it is not too large and has not spread, is more likely to better respond to effective treatment and can result in a greater improvement in survival rates, decrease in mortality, and less expensive treatment; 2. Screening – aims to detect cancer before the symptoms appear. The definition says that it is the presumptive identification of unrecognized disease or defects through tests, examinations, or other procedures that can be applied rapidly.[60]

However, implementation of the above preventive measures mentioned above in most cases cannot be accomplished due to the failure of systemic approaches.

Different Aspects of the Use of Nanocarriers - Prevention, Diagnostic and Therapeutic Application

Recently, increasing evidence demonstrates that nanoparticle-based targeting strategy is effective and promising at

Table 4 The Examples of Nanotechnology-Based Applications with Proved Antimicrobial Properties

Nanoparticles	Microbe	Mode of Action	Ref.
Metal and metal oxide NPs: 1. Ag NPs 2. BiOBr NPs with Fe^{3+}	*Staphylococcus aureus MRSA*	Disruption of the bacterial cell wall. Stops cell division by interaction with both DNA and RNA. Disruption of signal transduction, and ROS generation.	[40–43]
Carbon-based NPs: 1. GO 2. GO with curcumin 3. CNFs 4. NCQDs		Penetration of cell membranes. A significant enhancement of the anti-*MRSA*. Activity due to the illumination of LED lights. The positively charged NCQDs interacted with negatively charged bacteria and then anchored specifically to particular sites on the surface of *MRSA*.	[40,41,44,45]
Liposomes: 1. liposomal vancomycin 2. exosomal linezolid		Liposomal vancomycin (in comparison to free form) adequately accumulated vancomycin levels in the macrophages and exhibited a remarkable bactericidal effect against *MRSA* intracellularly. The in vivo assessment of exosomal linezolid (vs free linezolid) showed superior activity against intracellular *MRSA*.	[40,46–48]
Polymeric NPs: 1. diblock guanidinium polymer 2. encapsulation of vancomycin in the amphiphilic self-assembled supramolecular vesicles 3.C1-PNPs with gentamicin or ciprofloxacin 4.PEG-PLGA with Eudragit E100 and chitosan		The system is able to target macrophages, release an antibiotic inside the cell, and consequently increase the effectiveness against intracellular *MRSA*. Restore *MRSA* sensitivity to antibiotics as C1-PNPs increased the cellular uptake of gentamicin by *MRSA* and inhibited the *MRSA* efflux mechanism for ciprofloxacin. Target macrophages in an efficient way and also to improve the delivery of vancomycin to *MRSA* inside the cells, and conclusively improved the antibacterial activity of vancomycin on intracellular *MRSA*.	[40,49,50]
Silica NPs: 1.Gentamicin-loaded mesoporous silica NPs		The system is able to release loaded gentamicin upon the bacteria's presence followed with the bacterial toxins-caused degradation of the shell and thus could be used to treat the intracellular *MRSA*.	[40,51]
Polydopamine-based NPs: 1.PDA-PEG-Van NPs		High targeted antimicrobial activity against *MRSA* when exposed to NIR low-power radiation.	[52]
Metal and metal oxide NPs: 1. $Mg(OH)_2$ NPs	*Streptococcus bovis*	The mechanism of antimicrobial action of oxide NPs involves the production of active oxygen species which are known to induce bacterial cell death. The effect of MgO is stronger against Gram(+) bacteria than Gram(-) bacteria, most likely because of differences in bacterial membrane structure.	[53]
Metal and metal oxide NPs: 1. AG NPs 2. Tv-Ag NPs	*Helicobacter pylori*	Generation of ROS (oxidative damage) and exhibits antimicrobial and antibiofilm activity.	[54,55]
Noble metal NPs: 1.GNR@LDH-PEG NPs 2.GNSs 3.Tri-Ag NPs 4.dvPtNPs	*Escherichia coli*	NIR irradiation - thermally kill at least 99.99% of *E.coli*. Concomitant release of ROS and chemotherapeutic Pt^{2+}, resulting in tri-model (photothermic/photodynamic/chemotherapeutic) antibacterial activity against *E.coli*.	[56,57]
Metal sulfide/oxide NPs: 1.CS@MoS_2 2. MoS_2/PDA-RGD 3. CuS NPs 4. MnO_2 NPs		Photothermic and photodynamic effects of chitosan-assisted MoS_2 (CS@MoS_2), resulting in the inhibition of 99.84% of *E.coli*. MnO_2 can have interaction with GSH in bacteria and convert into Mn^{2+}, not only destroying the oxide balance of bacteria, as well as avoiding long-term body retention.	[56]

(*Continued*)

Table 4 (Continued).

Nanoparticles	Microbe	Mode of Action	Ref.
Polydopamine-based NPs: 1.PDA-coated polystyrene/silver NPs 2.PDA-modified magainin NPs		Specific interaction with bacteria and showed that under NIR radiation induced a temperature rise to 45 °C could cause a marked bacterial death.	[52]
Carbon-based NPs: 1. rGO-Au NPs	*Klebsiella pneumoniae*	Captured the bacterial cell wall membrane, and Au NPs destroyed the outer coating of the bacterial cell wall.	[41]
Metal and metal oxide NPs: 1. Ag NPs 2. GSH-Ag NPs	*Fusobacterium*	An exhibition of improved antimicrobial activity due to their enhanced surface-to-volume ratio. Trigger inflammatory response in human gingival fibroblasts by the increase of cytokine production.	[58,59]

Abbreviations: Ag NPs, silver nanoparticles; BiOBr NPs, polyethylenimine grafted bismuth oxybromide nanoplates with Fe^{3+}; C1-PNPs, pyridinium amphophile-loaded PLGA (poly (lactic-co-glycolic acid) nanoparticles; CNFs, carbon nanofibers; CS@MoS_2, chitosan-assisted MoS_2; CuS NPs, copper sulfide nanoparticles; dvPtNPs, dualvalent platinum nanoparticles; GNR@LDH-PEG, poly(ethylene glycol) (PEG) modified core-shell GNR@ (gold nanorod) layered double hydroxide nanoparticles; GNSs, gold nanospheres; GO, graphene oxide; GSH, glutathione; GSH-Ag NPs, glutathione-stabilized silver nanoparticles; MDR, multidrug-resistant; MIC, minimum inhibitory concentration; $Mg(OH)_2$ NPs, magnesium hydroxide nanoparticles; MnO_2 NPs, manganese oxide nanoparticles; MoS_2/PDA-RGD, molybdenum disulfide/polydopamine-arginine-glycine-aspartic acid; *MRSA*, methicillin-resistant *Staphylococcus aureus*; NCQDs, nitrogen-doped carbon quantum dots; NIR, Near-infrared irradiation; NPs, nanoparticles; PCE, tetrachloroethene; PDA-PEG-Van NPs, polydopamine-based nanoparticles modified with PEG and vancomycin; rGO-Au NPs, reduced-graphene-oxide functionalized with gold nanoparticles; ROS, Reactive Oxygen Species; Tri-Ag NPs, citrate-coated triangular nanoparticles; Tv-Ag NPs, toxicodendron vernicifluum silver nanoparticles.

a diagnostic and therapeutic level and might include many kinds of cancers, such as colorectal cancer, breast cancer, ovarian cancer, or lung cancer.[61,62] Nanotechnology can be used in the prevention of disease, diagnosis, and treatment, especially by enabling early disease detection and diagnosis, as well as a precise and effective therapy, which is vital for developing personalized treatment strategy. In effect, implementing the aforementioned new concept of personalized medicine potentially offers an efficient cure for virtually any type of malignancy. Various applications of nanotechnology concerning prevention, diagnosis and treatment fields of use are shown in Figure 3.

Many types of nanodevices could be clinically applicable, in different kinds of detection, such as imaging contrast agents, immunoassays, or targeted drug delivery systems. In Table 5, commonly used nanodevices and their primary areas of application are presented.

Treatments Using Drug Delivery Systems

An accurate cancer diagnosis is essential for adequate and effective treatment because each type of cancer requires a specific treatment regimen that encompasses one or more actions, such as surgery, radiotherapy, and chemotherapy. Determining treatment goals and palliative care is an essential first step, and health services should be integrated and patient-oriented. The fundamental aim is to cure cancer or to prolong life. Improving the patient's quality of life is not insignificant, and it can be achieved by supportive or palliative care with minimization of side effects of drugs as well as via psychosocial help.[73]

Nanocarriers used in drug delivery systems are typically about a size below 500 nm. They are made of organic (lipid, liposome, dendrimer, polymeric) or inorganic (carbon nanotubes, iron oxides, metallic) materials, as well as their hybrids of varying sizes, shape, and surface characteristics.[74] Examples of the most widespread anticancer drugs as part of drug delivery systems, specifying the nanocarriers and type of cancer, are presented in Figure 4.

To achieve targeted drug delivery with maximum pharmacokinetic activity at pathology sites, constant progress in drug delivery systems using nanotechnology strategies has been noted. The use of drug carriers offers several benefits in terms of the chemical and biological properties of the drug. From a chemical point of view, the application of nanocarriers exerts an impact on drug solubility and penetration ability. Moreover, surface characteristics, immobilization of homing molecules, as well as the sensitivity of carriers to different stimuli determine specific-site delivery, modulate drug release, exert the impact on biodistribution and retention process, as well as influence the immunomodulatory properties of carriers. The above-mentioned features show that a strong association between physicochemical and biological properties exists.

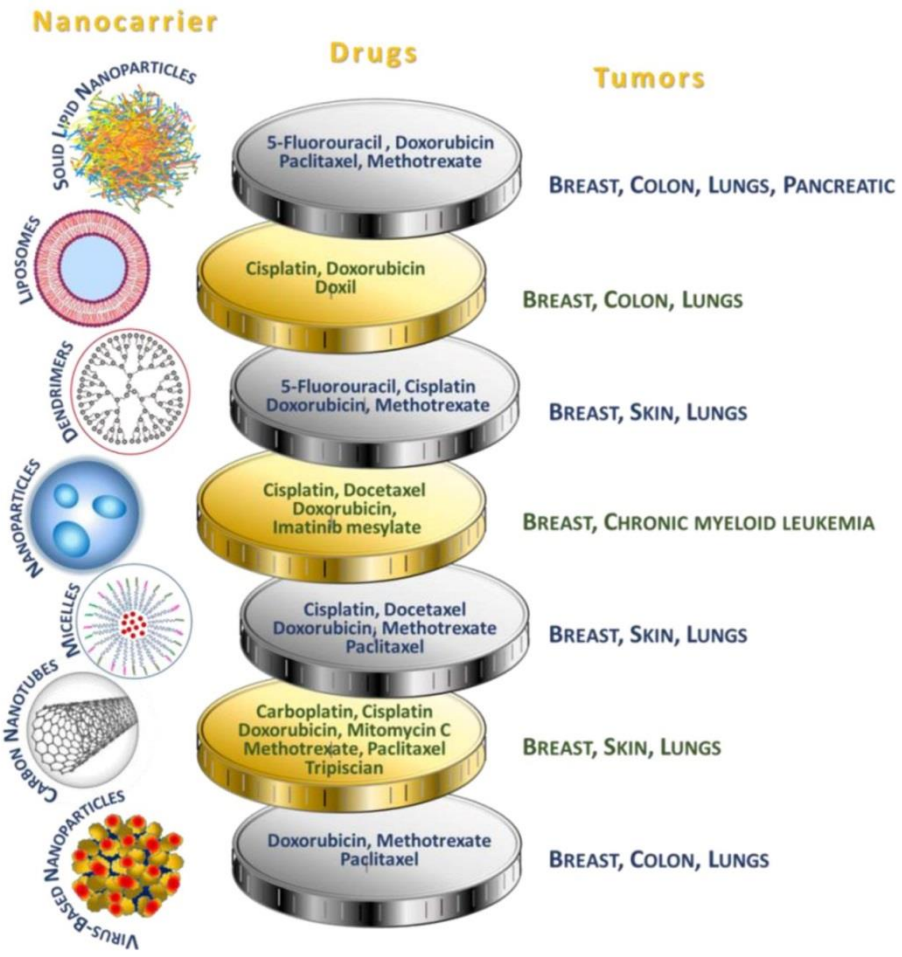

Figure 4 Use of nanocarriers.[73]

the hydrophobic region of micelles formed from diblock hydrophobic-hydrophilic polymers. As a result of drug encapsulation, a hydrophilic nanocarrier is created, which due to the typically low critical micelle concentration, remains stable even after dilution by body fluids. Drug-containing polymeric micelles, such as Genexol PM® containing Paclitaxel, already exist on the market.[76]

Another example of delivery of poorly soluble drugs is the liposomal formulation, where lipophilic drugs can be dissolved in the lipid segment of the phospholipid bilayer membrane. Liposomal carriers are very flexible when it comes to their structure and functionality. Lipid formulations of anticancer drugs have been successfully marketed, such as Endo®-TAG-1 which is a product containing Paclitaxel that uses positively charged phospholipid vesicles for pancreatic cancer treatment.[77,78]

Finally, hydrophilic dendrimeric polymers are recognized as suitable carriers because drugs can be

encapsulated in their interior. The presence of empty cavities can be controlled by affecting the polymer conformation by changing the pH, the type of solvent, as well as the design of the polymer structure itself. At the same time, the encapsulation mechanisms can utilize electrostatic, hydrophobic, acid-base interactions, or hydrogen bonds between the drug and the polymer. Although there is no dendrimer-based product on the cancer drug market, research shows that some known dendrimeric vehicles are good candidates. For example, it was reported that polyamidoamine branched polymers with hydrophobic Paclitaxel, in addition to better drug solubility, showed 10-fold higher anticancer activity compared to free drug, which is attributed to better uptake by tumor cells.[79]

Interestingly, in the latest literature, there are such bioinspired solutions for drug delivery as the use of amphiphilic proteins to stabilize the hydrophobic drug and induce biosilicification on its surface, which leads to the formation of drug-core silica-shell nanoparticles.[80]

Other interesting examples are hydrogels, biocompatible crosslinked hydrophilic polymer networks already well known for being a good hydrophilic drug delivery system, which can be modified to encapsulate hydrophobic drugs, for example, by having hydrophilic moieties or molecules having empty cavities in their structure, or even containing polymeric micelles or nanoparticles with the encapsulated drug.[81]

Targeted Drugs Delivery – the Passive and Active Crossing of Biological Barriers

A key element for the effectiveness of the drug is to successfully access diseased sites. This can be improved or enabled by the use of nanosized drug delivery systems, which themselves are capable of crossing biological barriers or allow the encapsulated drug to traverse them to achieve maximum effect at the target. Depending on the method of administration (intravenous, oral, or inhalation), the nanocarrier must cross various barriers on the way to the tissues or organs and subsequently to the cells or organelles, which takes place via two modes of transport, "passive" and "active". Targeted drug delivery systems (TDDSs) have many advantages, including (1) reducing the exposure of healthy cells to cytotoxic compounds, (2) overcoming the increasingly common drug resistance of tumors, and (3) reduction of side effects of therapy, which directly translates into profits from a pharmacoeconomic point of view.[82]

"Passive", non-specific targeting is associated with reduced nanoparticle sizes and surface properties, such as hydrophobicity, surface charge, or non-specific adhesion, which may result in reaching organs having porous endothelial capillaries (liver, spleen), helping to cross specialized epithelial, and penetrating the cell cytoplasm.[83] For example, in the case of cancer, the phenomenon of increased permeability and retention (EPR effect) can be observed, which is based on selective penetration into cancer cells compared to normal tissues due to the size of nanoparticles. This is caused by the leaky nature of the tumor-bearing blood vessels that have endothelial cell linings of 100 to 700 nm, which is 10- to 70-fold more than the normal endothelium. This, combined with the weak drainage system typical of solid tumors, leads to the accumulation of drug-loaded nanoparticles in the neoplasm.

Furthermore, due to the increased metabolism of tumor cells, their surroundings are characterized by acidic pH and slightly increased temperature, which can be used in the design of stimuli-responsive nanocarriers. Finally, tumors will release specific enzymes, such as metalloproteases, into their adjacent environment, which in addition to function as tumor markers, can also be recognized by functionalized drug delivery systems.[73]

Unfortunately, for some organs, the delivery of drugs passively using nanosystems is significantly impeded due to the poor permeability of biological barriers, such as the blood–brain barrier (BBB). In these cases, "active" transport methods can improve traversing through membranes.[84] "Active targeting" relies on the increased selectivity of the drug-loaded nanocarrier through its surface functionalization with a ligand showing an affinity for the pathological site. Such ligands, including antibodies, peptides, proteins, glycoproteins, growth factors, nutrient compounds, vitamins, or nucleic acids, are bound by receptors that are overexpressed on cancer cells. Then, receptor-mediated endocytosis ensures cellular uptake of nanocarriers providing higher drug concentration in the cytoplasm.[73] An interesting example of a ligand is folic acid, whose receptor (FR) is overexpressed in many types of cancer, such as breast, lung, ovarian, and colorectal tumors.[85] Among classical targets, there are transferrin receptors (TfR) or nicotinic acetylcholine receptors typical for the vasculature of brain tumors.[86]

Furthermore, targeting tumor endothelium on which there are numerous moieties, such as vascular endothelial growth factors (VEGFs) or vascular cell adhesion molecules (VCAMs) can be a complementary strategy to drug

delivery, as it involves the destruction of endothelial walls, and thus cutting off oxygen and nutrient access leading to cell death.[73]

Another advantage of nanocarrier functionalization is the conjugation of the carrier with a fluorescent marker that allows tracking of both the carrier and the drug in vitro and in vivo studies, which can be used in theranostics.[87]

Nowadays, most of the clinical trials using nanocarriers apply "passive" transport,[85] and the use of the EPR effect in the design of drug delivery systems has become standard. Some of these products are commercially available, such as Doxil®, a liposomal formulation of the cytotoxic Doxorubicin, or Caelyx®, a PEGylated liposomal formulation of this drug. Besides, many studies are documenting the in vivo antitumor activity of nanosystems using an "active" mechanism of cell penetration, and some of them are at the clinical trials, including a liposomal nanoplatform containing Doxorubicin with scFv antibody as a ligand targeting the human epidermal growth factor (HER2) receptor in advanced breast cancer, and a polymeric nanoplatform having Docetaxel with nucleic acid-based protein-ligand (ACUPA) targeting prostate-specific membrane antigen (PSMA) in solid tumors.[86]

Increasing Drug Stability and Controlled Release

Drug delivery systems (DDSs) have many advantages over using free medicines. Often, one type of carrier significantly improves a given therapy by improving several chemical properties of the formulation, thereby increasing the stability of the formulation and drug during storage, the stability of the formulation in vivo, and also allowing for prolonged release of the drug.

Maintaining the unchanged properties of the drug during storage and extending its suitability for use in the drug delivery systems can be very helpful. For example, it was reported that a carrier made of cyclodextrin could result in increased thermal stability and reduced drug volatility.[88] Another case was described by Hsiao and coworkers, who showed that chlorophyll, a valuable bioactive compound known for its sensitivity to oxygen, high temperature, and light, has been encapsulated in polycaprolactone, gaining greater stability and therefore being more convenient for storage.[86]

The drug delivery system can lead to increased drug stability in vivo and protect it from degradation before and after it gets into systemic circulation by decreasing metabolic clearance in blood and gastrointestinal tract (GIT) or renal reticuloendothelial system (RES) clearance. However, it is very important to maintain constant nanoparticle parameters, such as size, morphology, size distribution, porosity, or crystallinity, because their disturbance can lead to altered pharmacological properties of the drug-loaded nanosystem. Some active moieties, such as DNA or siRNA, possess disadvantaged physicochemical properties (molecular weight, charge, susceptibility to degradation by enzymes) and have to be applied clinically together with appropriate nanocarriers.[88,89] Specifically, in the case of immobilization of enzymes on nanocarriers, in addition to increased stability, they are attributed to such benefits as reduced protein degradation, resistance to mass transfer, high mechanical strength, and minimum diffusional problems.[90] One should also mention the "stealth" technology used for liposomes, which consists of attaching a synthetic polymer poly(ethylene glycol) (PEG) to the liposome structure. This modification extends the presence of intact pegylated nanocarriers in the blood through reduced uptake by the mononuclear phagocyte system (MPS).[91]

The formulation must be stable to external factors mimicking conditions in the body, and therefore without the evaluation nanomaterials cannot be used clinically.[83] For instance, Villamizar-Sarmiento et al carried out a comprehensive study and confirmed that the prepared nanomedicines based on poly(styrene sulfonate) polymer had unchanged hydrodynamic sizes and zeta potential for over a dozen days at a varying salt concentration (NaCl), pH, and temperature, and was durable despite freeze-drying and redissolving in water.[92] Similarly, Kanwar et al studied structural changes of nanostructured lipid carriers (NLCs) under stress conditions, such as changing electrolyte concentration, pH, and stabilizing polymer addition. Interestingly, NLCs are resistant to changes in the environment, which is important for their pharmaceutical applications.[93]

The immobilization of a drug, both hydrophobic and hydrophilic, helps to ensure its controlled slow-release and avoid burst effect, which would not have been possible without the carrier.[94] As a result of slow controlled drug release, the active substance has a prolonged circulation in the body and is released at pathological target sites. In one of the strategies, due to the specific chemical properties of the designed nanocarrier, its durability can be controlled in vivo by local stimuli, such as abnormal pH,[95]

temperature,[96] or ionic strength[97] (so-called stimuli-responsive materials). For instance, Guo et al reported the synthesis of carriers consisting of cationic liposomes coated with carboxymethyl chitosan, stable under physiological conditions, but in an acidic environment specific to the tumor (pH=6.5) quickly transformed into a cationic form, which aided tumor-specific cellular uptake. Moreover, in the presented studies synergistic use of two active molecules, the anti-cancer drug (doxorubicin) and oncogenic protein inhibitor (MDM2), was possible using the dual-drug delivery system.[98] Recently, Razavi et al described multi-stimuli-responsive block copolymers based on poly(*N*-(2-(dimethylamino)ethyl)-methacrylate) (PDMAEMA) and poly(methyl methacrylate) (PMMA) chains terminated with spiropyran, wherein the size of the nanoparticles, as well as the release of doxorubicin, was controlled through pH, light, and temperature.[99]

The Efficiency of Encapsulation/ Immobilization of Drugs in Carriers

From a pharmacological point of view, it is important to ensure efficient drug encapsulation to avoid such in vivo side effects of the use of nanocarriers in excess such as agglomeration resulting in excretion from the body by the immune system, high blood pressure, renal failure or systemic toxicity.[100] Unfortunately, the majority of currently known drug delivery systems are characterized by a low loading efficiency (less than 10%), which is associated with the use of a large amount of carrier.[101] To achieve good loading efficiency, the kind of materials used (characterized mainly by a large surface area) and their surface modification and the method of drug encapsulation/immobilization are important. In general, the mechanism of drug loading through non-covalent interactions most often results in low loaded drug carriers, and covalent or coordination bonds result in high drug loading efficiency. Such non-covalent bonds are electrostatic interactions, π-π stacking, hydrogen bonding, or hydrophobic/hydrophilic interactions of the drug with the surface of the carrier. For example, the most popular carrier, liposomal, depending on its morphology, is characterized by hydrophobic or hydrophilic drug–carrier interactions. In the case of polymer nanoparticles or dendrimers, they may form structures that allow the drug to become entrapped in a micellar or hollow structure, respectively, or to bind the drug via a chemical linker. Typically, enzymatically or chemically cleavable linkers are used, such as amide, ester, disulfide bonds, or phosphate esters. There are also examples of specific linkers sensitive to the stimulus or enzyme typical of the tumor environment. For example, disulfide bonds can be broken by glutathione, an enzyme that is overexpressed on cancer cells.[102,103]

Due to the type of nanocarriers' structure, the following types of high drug loading nanomedicines can be distinguished: 1. Inert porous material as a carrier (silica, carbon, or protein nanoparticles); 2. Polymer-drug conjugates (PDCs); 3. Coordination polymer nanoparticles (metal-organic frameworks); 4. Carrier-free nanomedicines (drug nanocrystals, amphiphilic drug–drug conjugates).[103] The PDCs systems used are solid dispersion of the drug in a hydrophilic polymer, and nanoconjugates of an amphiphilic or hydrophilic polymer with the drug. Recently, various PDCs carrier improvement strategies have been introduced to enhance loading efficiencies, such as the use of: 1. Multi-arm polymer conjugated with drug;[104] 2. The hydrophobic[105] as well as the hydrophilic[106] drugs as part of the core-shell carrier structure; 3. Two drugs with opposite hydrophilicity linked via a hydrophilic carrier (spacer);[107] 4. Encapsulation in core-crosslinked polymer.[108,109]

Another class of nanomaterials that overcomes the problem of low drug loading is nanocages (protein, gold, carbon, silica, or DNA NCs), which have a hollow structure and can contain up to thousands of drug particles inside.[97] A different way to increase the effectiveness of drug loading is surface modification. For example, porous iron oxide nanoparticles (IONPs) coated with materials, such as silica, surfactants, carbon, and polymers are used as drug carriers. Moreover, the introduction of functional groups on the surface allows its further modification, for example, with proteins, which further increases the affinity for drugs.[103] Another example describes calcium phosphosilicate nanoparticles (CPSNPs) as phospho-drug nanocarriers (5-Fluorouracil) where due to metal-ligand complexes between the phosphate group and calcium, efficient drug encapsulation is possible.[99]

It turns out that the effectiveness of the encapsulation procedure depends on many factors, and in the literature, comprehensive analyses can be found regarding specific carriers in combination with various medicines and encapsulation methods. For example, the fact that the route of immobilization should be selected depending on the type of medicine was described by Krukiewicz et al where two different loading methods have been tested with two various active substances. For quercetin, the highest loading

was achieved by immobilization on a polypyrrole matrix during the electropolymerization process, while in the case of a second drug tested, ciprofloxacin, incorporation during post-modification (polymer oxidation) was more efficient.[110] Furthermore, Perotto et al reported that in addition to such medicine characteristics as hydrophilicity and molecular weight, the charge of the drug might have the most significant impact on its encapsulation, as in the case of positively charged methylene blue-achieving up to 88% encapsulation efficiency in keratin nanoparticles.[111] Besides, the study of curcumin encapsulation into poly ε-caprolactone NPs was carried out by Nagy and coworkers using Box–Behnken experimental design, where the variables in the encapsulation procedure were the initial amount of the drug, the volume ratio of the organic and aqueous phases, as well as the composition of the organic phase. It was found that the volume of the organic phase containing a drug used for nanoprecipitation of the polymer was crucial for efficient drug loading.[112]

In the latest literature, one can also find reports about drugs encapsulated in high loading carriers by environmentally friendly methods. That is, due to aromatic–aromatic interactions and the formation of ionic pairs, hydrophilic and aromatic low molecular weight drugs (HALMD) were encapsulated in a poly(styrene sulfonate) (PSS) with the yield of about 50%.[113]

Application, Mechanism of Action, and Drug-Resistance of Selected Chemotherapeutics

Doxorubicin (DOX) is commonly used in various types of malignancies, such as sarcoma, leukemia, lymphomas, breast, lung, and ovarian cancer. There are two different mechanisms of action: intercalation of doxorubicin into DNA and inhibition of topoisomerase II leading to changes in chromatin structure; generation of free radicals and oxidative damage to biomolecules. Repeated doxorubicin administration leads to drug-resistant cancer cells; it also increases drug cytotoxicity. The interaction between signaling pathways can promote drug resistance through the induction of proliferation, cell cycle progression, and prevention of apoptosis. Doxorubicin-induced drug resistance and tumor growth can occur through the adaptive role of the MAPK/ERK pathway in the effort to protect tumor cells. The mechanism of drug resistance of the Anatomical Therapeutic Chemical Classification System (ATC) is related to the expression of multidrug-resistant 1 (MDR1) transporters. MDR1 transporters pump Dox molecules out of cells, reducing intracellular concentration of drug and inhibiting chemotherapeutic efficacy.[114,115]

5-Fluorouracil (5-FU) could be applied to treat solid tumors of the gastrointestinal tract, breast, head, and neck, as well as the pancreas. Mechanism of action involves blocking DNA synthesis and replication through inhibition of thymidylate synthase and incorporation of 5-FU metabolites into RNA and DNA. 5-FU resistance abrogated the anticancer effect amplified by the Chk1 inhibition, even in p53-deficient cancer cells. Chk1 inhibition might be effective in sensitizing 5-FU resistant cancer cells to 5-FU because Chk1 activation is reported to be related to the resistance to chemotherapy. It has also been observed that the synergistic cytotoxic potential for Chk1 inhibition during 5-FU treatment in p53-deficient colon cancer cells with or without 5-FU resistance.[116,117]

Paclitaxel (PTX) is used against many forms of cancer, for example, ovarian, breast, lung, Kaposi sarcoma, cervical, and pancreatic cancer. Mechanism of action relates to targeting microtubules – it disrupts the major function of microtubules, which is the production of the mitotic spindle during cell division, as well as maintenance of the cell structure, motility, and cytoplasmic movement within the cell. A weakened mitotic checkpoint confers only short-term resistance to mitotic arrest but also the activation of a mitotic checkpoint followed by mitotic slippage resulting in optimal cell killing. There are some identified markers of resistance or sensitivity to paclitaxel, such as proteasome subunits, cyclin-G1 (CCNG1), and solute carrier genes. The cytotoxicity of nanoparticles using tamarind seed polysaccharide and paclitaxel by epichlorohydrin crosslinking (PST-PTX) in cancer cell lines and resistant cancer cell lines were determined by MTT assay. Quantitative analysis of cell death was determined by Annexin V dead cell assay, Caspase 3/7 assay, and expression of pro-apoptotic protein Bax. Overexpression of the ABCB1 gene confers resistance to nab-paclitaxel in urothelial cancer cells.[118,119]

Each of these drugs has a different field of application, mechanism of action, and also various explanations of drug-resistance. Cells become resistant to different drugs through various mechanisms of modification of drug targets, alteration in drug metabolism, and genetic changes of cells to target pathways.[120] However, it is worth noting that despite these differences, resistance to drugs continues to be a principal problem in oncology, affecting most cancer patients.

Improving Activity and Help to Overcome the Drug-Resistance

Currently, major treatments for cancer management include cytotoxic chemotherapy, surgery, targeted therapy, radiation therapy, endocrine therapy, and also immunotherapy. Despite the efforts and achievements made in treating cancers during the last few decades, resistance to classical chemotherapeutic agents and novel targeted drugs remains a major problem in cancer therapies.[121] Drug resistance, also the one existing before treatment (intrinsic) or generated after therapy (acquired), is responsible for most relapses of cancer, which are the major causes of death of the disease. Heterogeneity among patients and tumors and the comprehensiveness of cancer to circumvent therapies make drug resistance even more difficult to deal with. A better understanding of the mechanisms of drug resistance is required to provide guidance to future cancer treatment and achieve better results.[121] The complexity of drug resistance development suggests that combinational and personalized treatment might provide better approaches and improved efficacy for fighting drug resistance in cancer.[122]

Cancer presents difficult challenges that would benefit from uniting experts from a broad cross-section of related and unrelated fields. Combining extant approaches with novel ones could help in raising this challenging health problem, enabling the development of therapeutics to stop disease progression and prolong patient lives.[122] Regardless of the research approaches, based on the results from clinical trials and research publications on the application of nanoparticles as drug delivery systems in the treatment of cancer, the main benefits are the enhancement of vascular and gastrointestinal permeability and selectivity of drugs/compounds to tumor cells. Abdifetah et al,[123] in their summary of the review, note the fact that due to the application of nanoparticles, the improved permeability and selectivity resulted in the overall improvement of cellular drug uptake, the inhibition of drug hepatic first-pass metabolism and P-gp efflux, the increase in drug solubility and stability, and the decrease in the rate of the drug excretion. As a consequence, a reduced dosage can be achieved without compromising the efficacy, which minimizes potential drug toxicity. Still, regardless of the therapeutic and research progress made, some of the challenges in cancer therapy, such as multidrug resistance (MDR), are being further investigated to better understand the molecular mechanisms and optimize the therapies concerning efficacy and safety. According to El-Readi et al,[124] due to the tumorous tissue specifics such as their abnormal blood vessels and pathologic processes that hinder effective cancer chemotherapy, the design and application of new methodologies for drug delivery like NPs are vital. MDR is known to be a result of synergistic processes taking place directly in cancer tissues and tumorous cells. In Figure 5, different mechanisms synergistically, causing multidrug resistance (MDR) are summarized.

The influence on membrane transport is one of the most important mechanisms in the development of resistance against anticancer drugs. The reduction of drug concentration can be achieved by reduced drug uptake or increased extrusion of the molecules. The overexpression of P-glycoprotein is responsible for efflux. The use of nanoparticles loaded with docetaxel (PLGA-PEG) has proven to be effective in overcoming the MDR as referenced in the article.[126] The authors also listed other advantages of the application of NPs in the therapy over the standard dosage forms; for example, nanosized drug carriers minimize the elimination of the molecules substantially through the liver or kidney. Other properties like improved permeability and accumulation of nanoparticles loaded with drugs are passively targeting tumor tissues resulting in lower systemic toxicity.

Another successful application of targeted anticancer nanocarriers using biocarriers is presented in the article by Radu et al.[127] Poly(3-hydroxybutyrate-*co*-3-hydroxyvalerate) carriers were obtained via the emulsification-diffusion method, loaded with 5-fluorouracil and therapeutic potential on human adenocarcinoma cells was investigated. As a result, it was observed that the drug-loaded carrier could significantly decrease cell viability, showing the high potential of destroying human adenocarcinoma cells. Overall, significant progress has been made in the field of nanocarriers in cancer treatment resulting in improved pharmacokinetic properties, better antitumor efficacy, and lower risk associated with the development of undesirable drug effects. Physicochemical properties of the therapeutic nanocarriers and pathophysiological tumor characteristics still need to be investigated to get deeper insights into the mechanisms allowing effective and safe cancer treatments. Arranja et al reported a list of clinically used nanomedicines containing mainly liposomes, polymer-drug conjugates, and polymeric micelles.[128] In contrast to traditional chemotherapy, nanomedicines are characterized by prolonged circulation half-lives, increased bioavailability, and better tumor disposition;

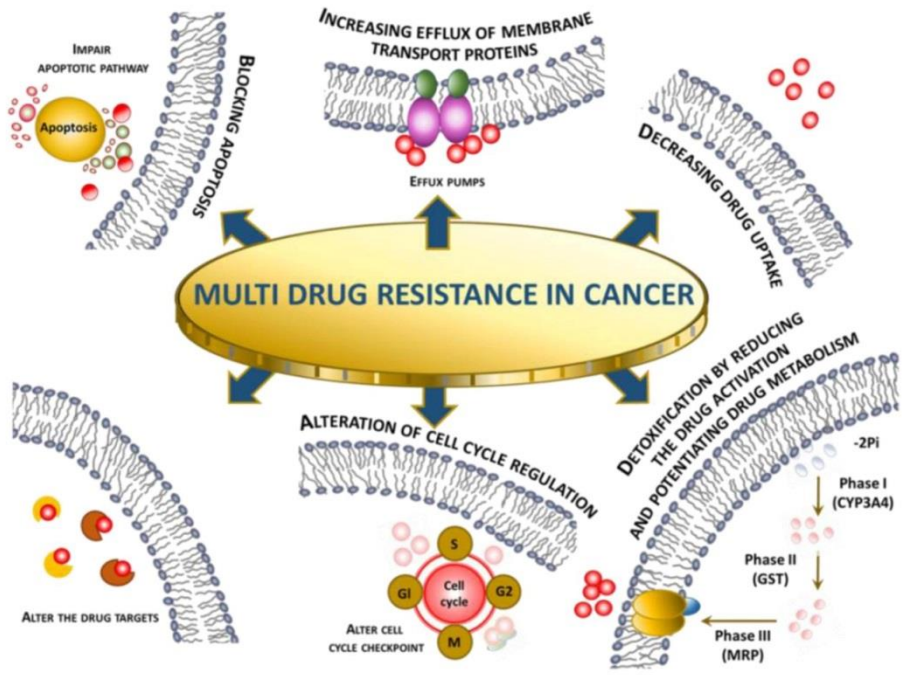

Figure 5 Multidrug resistance in cancer mechanism overview.[125]

however, they rely mainly on the EPR effect. To increase our understanding of actively targeted nanodrugs, the authors suggest and discuss the application of strategies from theranostics. The main aim of this approach is to integrate molecular imaging properties into therapeutic agent formulations to monitor tumor accumulation and therapeutic efficacy of nanomedicines at the same application time. More controlled targeted drug delivery should further optimize therapeutic effects minimizing unwanted cytotoxicity in the off-target tissues.

Cancer multidrug resistance (MDR) to chemotherapy is a crucial barrier in the effective treatment of malignancies, which may lead to therapeutic failure of the treatment regimen. Nanotechnology ensures a novel and unconventional approach to circumvent MDR. In Table 6, recent literature examples of application nanocarriers to overcome MDR are presented. Mechanisms and advantages of various types of nanocarriers were discussed below as well as potential approaches to overcome these limitations.

Establishing a practical nanotechnology-based drug delivery systems may help in the future to improve the bioavailability and therapeutic efficacy of antitumor drugs while providing better accumulation at the target site compared with conventional antitumor drug delivery systems.

Pharmacoeconomic Aspect of Drug Carriers

The efficacy of selected drugs due to their equivalents in nanocarriers could have an impact on reducing or minimizing costs in pharmacoeconomic analysis, especially in shortening the time of hospitalization or a smaller number of tests carried out. We could also avoid some intangible costs, such as pain, suffering, or anxiety – if the patient stays shorter in the ward and could be faster at home. What is more, we can reduce the number of inpatient days, resulting in decreased risk of infections and

Table 6 Mechanism of Overcoming Drug Resistance and Benefits of Nanocarrier Use

Drug	Type of Cancer	Type of Nanocarrier	Mechanism of Overcoming Drug Resistance and Benefits of Nanocarrier Use	Ref.
DOX	Ovarian cancer	Iron oxide-titanium dioxide core-shell nanocomposites	Downregulation of TfR1 expression	[120]
	ATC	Dopamine-melanin NPs	Increased cellular uptake	[129]
5-FU	CRC	Mesoporous silica NPs grafted with EGF	Cell death through S phase arrest Downregulation of DPYD expression	[117]
	GC	Gelatinase-stimuli di-block copolymers poly(ethylene glycol)-*b*-poly(ε-caprolactone) (PEG-*b*-PCL)	Upregulation of TFAP2E and downregulation DKK4	[130]
		Chitosan NPs	Downregulation of HIF-1α expression	[116]
PTX	Lung cancer	Galactoxyloglucan	Downregulation of the expression of multi-drug resistant proteins P-gp and BCRP	[131]
	MDCK-MDR1	Two diblock copolymers, MePEG114-*b*-PCL200 and MePEG17-*b*-PCL5 (PCL200/PCL5) + ultrasound	Increased accumulation of drug	[132]

Abbreviations: 5-FU, 5-Fluorouracil; ATC, anaplastic thyroid cancer; BCRP, breast cancer resistance protein; CRC, colorectal cancer; DKK4, Dickkopf WNT Signaling Pathway Inhibitor 4; DOX, Doxorubicin; DPYD, Dihydropyrimidine Dehydrogenase; EGF, Epidermal Growth Factor; GC, Gastric Cancer; HIF-1α, Hypoxia-inducible factor 1-alpha; MDCK-MDR1, Madin Darby canine kidney (MDCK) cells with the MDR1 gene; NPs, Nanoparticles; P-gp, Permeability glycoprotein; TFAP2E, Transcription Factor AP-2 Epsilon; TfR1, Transferrin Receptor 1.

medication side effects, improve quality of treatment, and increase hospital profit through more efficient bed management.[133]

As a result of the use of drug carriers we can observe the following benefits: 1. The economic benefits result from the savings associated with a more cost-effective medical procedure; 2. Clinical benefits are defined as the direct positive effects of the applied therapy, measured by primary or secondary endpoints. The size of clinical benefits is a measure of the clinical effectiveness of the examined medical procedures; 3. Unmeasurable benefits concern the reduction of pain, anxiety, and improvement of life comfort and its duration.

Comparing the use of traditional therapy with alternative therapy, such as nanocarrier-based-therapy, we can evaluate examples of systemic treatment parameters in oncology such as Evaluation of response to treatment (%); Percentage of corresponding patients (%); Percentage of total remissions (%); Time to relapse (months, years); Percentage of reduction in risk of recurrence (%); Percentage of 5-year survival rate (%); Percentage of responses to treatment (%); Percentage of total pathological remissions (%); Total survival time (months); Median survival (months); Indicators of quality of life and reduction of symptoms, such as VAS procedure.[134]

Clinical studies have demonstrated the effects of using PEGylated-liposomal doxorubicin in adjuvant chemotherapy for advanced and metastatic breast cancer (Table 7). Reflected in Table 7, results review the clinical application of PLD in the adjuvant chemotherapy of breast cancer and illustrate the therapeutic effects of pegylated liposomal doxorubicin in various treatment regimens. These clinical studies, which presented therapeutic strategies for applying listed drugs to such adjuvant chemotherapy, show a significant improvement in the treatment results in terms of increased survival time as well as progression-free survival time. Both of these indicators are crucial in the effective treatment of oncological patients.

Over the past decade, the application of nanomaterials for the treatment of cancer features high sensitivity, specificity, and efficacy. Nanomaterials could be applied to employ specific ligands to target cancer cells predictably and deliver encapsulated load capacity effectively. Besides, nanomaterials can also be created for enhanced drug loading, greater half-life in the body, sustained release, and selective distribution by transforming their size, composition, morphology, and surface area. For instance, carbon-based materials, polymeric nanomaterials, metallic nanoparticles, dendrimers, and liposomes have been developed as smart drug delivery systems for cancer treatment, showing improved pharmacokinetic and

Table 7 The Effects of Using Pegylated Liposomal Doxorubicin (PLD) for Adjuvant Chemotherapy of Advanced and Metastatic Breast Cancer

Treatment Regimen	Phase of Clinical Study	Number of Enrolled Patients	Results			Ref.
			The Total Effective Rate (%)	Median Progression-Free Survival (Months)	Total Median Survival (Months)	
Evaluation of the effect and safety of salvage chemotherapy for treating metastatic breast cancer with PLD (40 mg/m^2)+cyclophosphamide (500 mg/m^2) and 5-fluorouracil (500 mg/m^2) in the presence of paclitaxel.	II	45	41.9	8.2	Up to 36.6	[135]
Adjuvant chemotherapy in patients with advanced HER-2 positive breast cancer by PLD (administered at 40 mg/m^2 every four weeks) in combination with lapatinib and trastuzumab.	II	-	4	5.8	23.3	[136]
Comparison of combined PLD (administered at 30 mg/m^2 every three weeks) and docetaxel with the separate use of docetaxel.	III	-	25-36	7.0-9.8	-	[137]
Examination of the therapeutic efficacy of PLD (administered at 20 mg/m^2 every two weeks) in elderly patients with advanced breast cancer and all patients enrolled were older than 70 years.	-	-	33.3	10.3	-	[138]
Evaluation of the combined regimen of PLD (administered at 40 mg/m^2 every four weeks) and navelbine (administered at 25 mg/m^2 every four weeks) and its therapeutic efficacy in first-line chemotherapy in elderly patients with metastatic breast cancer.	-	34	50	-	3 of 34	[139]

pharmacodynamic parameters over standard formulations because of their nanosize and individual physicochemical properties.

The data presented in Table 3, suggest that nanotechnology will provide new opportunities for cancer management. Moreover, a range of nanoparticles demonstrate significant efficacy for anticancer therapies, and their application can also be discussed in the pharmacoeconomic context. Considering that all the presented benefits from the use of nanomaterials make nanotechnology much cheaper than conventional treatment, it can also be reflected in the expected pharmacoeconomic efficacy. This could result in the reduction or total avoidance of costs in the management of cancer patients, particularly by reducing costs of interventions, shortening the time of hospitalization or avoided expenditure on illness which results in fewer medical procedures carried out, leads to the reduction of personnel costs and allows patients to return to professional life.

Clinical Application of Drug Carriers

The website clinicaltrials.gov was searched on 09.12.2020. The search was conducted using the keywords: cancer and nanoparticle. The start and end dates of the study were determined from 01.01.2015 to 09.12.2020. The status of the study was also defined – only studies with "completed" status were taken into consideration. As a result of this search, 13 studies meeting the above criteria were found. The search strategy is presented in Table 8.

To summarize, in Table 9, all studies are interventional (clinical trials), which are presented on the clinicaltrials.gov website. Each study involves a different number of patients, ranging from 2 to 146 participants. Different types of cancer were investigated, and the degree of severity is also taken into account, whether or not it is metastatic cancer. Each study describes arms – experimental or placebo, as well as treatment/other intervention. The selected endpoints – primary, secondary, or other – are included in the studies as per the protocols.

Table 8 Terms and Synonyms Searched in Clinical Trials Database

Terms/Synonyms	Search Results*	Entire Database**
Nanoparticle	13 studies	452 studies
Cancer	13 studies	78,405 studies
Neoplasm	13 studies	70,026 studies
Tumor	6 studies	17,331 studies
Malignancy	-	3274 studies
Neoplasia	-	651 studies
Neoplastic Disease	-	22 studies
Neoplastic syndrome	-	618 studies
Oncology	-	1348 studies

Notes: - No search results. *Number of studies in the results matching the search term or synonym. **Number of studies in the entire database matching the search term or synonym.

Unfortunately, so far, no results have been published for any of the thirteen studies, so we cannot draw any conclusions, but we can state that the use of nanoparticles in medicine, in the treatment of cancer, is becoming increasingly popular.

Business Criteria for the Development of Drug Carriers

During the manufacturing of drug forms, different methods should be considered. The selection of manufacturing methods often depends on the final product's requirements in terms of clinical efficacy, including size distribution, chemical composition, and drug release characteristics together, which dictates the pharmacokinetic demonstration of adsorption, distribution, metabolism, and elimination (ADME).[181]

Reducing Cost/Reagents/Green Synthesis

It is estimated that the development of a new nanodrugs takes only about 3–4 years and $20- $50 million. In comparison, discovering new active molecules takes more than 10 years, costing an average of about $500 millions.[182]

In order to perform the procedure for obtaining a drug delivery system designed in a laboratory on an industrial scale, careful optimization of the synthesis must be carried out in order to reduce the production costs. For example, Ding et al carried out a tedious optimization of polymer synthesis for protein therapy by changing time, solvent, and equivalents of reagents. As a result, the cost of a polymer prepared on a few hundred grams scale, following the principles of green chemistry, was reduced by almost 90%.[183]

Furthermore, in industrial-scale production, the time of synthesis directly translates into cost; thus, it is important to choose the most time-efficient[184] and inexpensive production method.[185] Finally, affordable, non-toxic, and common solvents, such as water, are most desirable.[111] Interestingly, to reduce the time and cost of formulation development, computational methods are used to predict in vitro/in vivo properties of carriers, such as stability, solubility, and potential toxicity.[186]

It is also worth noting that multifunctional carriers with targeting and imaging properties as well as multistep synthesis and greater regulatory hurdles thereof are worth the cost due to their numerous advantages, such as reducing side effects, dosing frequency, use in theranostics, and even reducing the toxicity of the drug, as proved by Cheng et al.[187] Despite higher production costs, recent analyses show that the use of targeted drug delivery systems for cancer patients leads to long-term reduced healthcare utilization and expense.[188]

Transfer of Drug Carriers Synthesis Methods from Lab to Industry - Challenges

Despite increased interest in nanodrugs in recent years, the transfer of methods to the market is still a challenge due to the difficult industrial transfer.[189,190] In general, procedures of nanocarriers synthesis are sensitive to reaction conditions and the characteristics of nanomaterials (size, charge, shape, morphology, and dispersity) can be easily disturbed due to scaling-up and thus formulation and effectiveness of nanodrugs may change.[191] Furthermore, these parameters are very important for the in vivo stability and toxicity of nanocarriers.[192]

In an industrial plant, the particle size can be affected by the available chemical reactor volume, stirring velocity, and time, as well as the energy used during the synthesis. These fluctuations in features may further lead to decreased efficiency of drug loading.[193,194] One of the examples of difficulties associated with large-scale production can be Doxil, the first nanodrug authorized in 1995, whose sales were suspended in 2011–2014 due to production and sterility problems.[195] Furthermore, it was described in the literature how scaling-up generated new minor impurity, which was found to be cytotoxic and

Table 9 Characteristics of Searched Clinical Studies

I. An Early Phase Study of Abraxane Combined with Phenelzine Sulfate in Patients with Metastatic or Advanced Breast Cancer[168] (8 participants) Metastatic Breast Cancer		
Selected Study Details	**Arms and Interventions**	**Brief description and Outcome Measures**
Official Study Title: A Phase Ib Safety and Pharmacokinetics (PK) / Pharmacodynamics (PD) Study to Determine the Dosage of Abraxane in Combination with Phenelzine Sulfate in Metastatic or Inoperable Locally Advanced Breast Cancer **Study Phase:** Phase 1b **Study Objectives:** To determine the safety and efficacy after administration of Abraxane and phenelzine sulfate (Nardil) for metastatic or locally advanced breast cancer. **Study design:** Interventional, open label, non-randomised, cumulative cohort group design (5 groups) with a defined target toxicity fraction of 30% and a corresponding margin of 10%. The toxicity fraction is defined for clinical application and study as the number of study subjects receiving particular dose who experience a Dose-Limiting Toxicity. **Study Status:** completed (October 30, 2019)	**Arms:** 1. Nanoparticle albumin-bound paclitaxel. Abraxane *i.v.* (100mg/m^2) 2. Phenelzine Sulfate (Nardil) *p.o.* initial dose of 15mg/d to a max dose of 90mg/d	**Brief Summary** Participants diagnosed with metastatic breast cancer or inoperable locally advanced breast cancer of age 18 years or above will receive combination of intravenously administered Abraxane and phenelzine sulfate orally. Both of the medicinal products have been applied in clinical practice for years however the combination of both has not been given for cancer therapy. The aim of the study is to investigate the effect of those two drugs together. Safety and efficacy will be assessed weekly for the period of 3 administration cycles. Abraxane administration is given weekly for the first 3 weeks of a 4-week period for 3 consecutive cycles. Phenelzine sulfate will be given daily for 3 cycles. Study is divided in five cohort groups each will be given a progressively increasing dose of phenelzine sulfate. **Primary Outcome Measures** Dose-Limiting Toxicity events assessed during the initial fifty-six (56) days **Secondary Outcome Measures** 1. Abraxane Cmax 2. Abraxane Tmax 3. Abraxane Half-life 4. Abraxane AUC 5. Nardil Cmax 6. Nardil Tmax 7. Nardil Half-life 8. Nardil AUC 9. Circulating Tumour Cell burden: a. PDL1 expressing Circulating Tumour Cell b. HER2 expressing Circulating Tumour Cell c. FFPE Tumour cells d. FFPE Stoma cells e. FFPE Cancer Stem Cells

II. Nanoparticle Albumin-Bound Rapamycin in Treating Patients With Advanced Cancer With mTOR Mutations[169]

(2 participants)

Advanced Malignant Neoplasm; Cervical Squamous Cell Carcinoma; Endometrial Carcinoma; Malignant Uterine Neoplasm; Recurrent Carcinoma of Bladder, Breast, Cervical, Head, and Neck, Ovarian, Prostate; Recurrent Malignant Neoplasm; Recurrent Renal Cell Carcinoma; Solid Neoplasm Stage III; Bladder Cancer Stage: III, IVA, IVB; Prostate Cancer Stage III & Stage IV; Renal Cell Cancer Stage IIIA & Stage IVA; Breast Cancer Stage: IIIA, IIIB, IIIC, IV; Cervical Cancer Stage: IIIA, IIIB, IVB; Ovarian Cancer Stage: IIIB, IIIC, IV

Selected Study Details	Arms and Interventions	Brief description and Outcome Measures
Official Study Title: A Pilot Study of a Rapid Access Platform for Investigational Drugs (RAPID) in Advanced Cancers **Study Phase:** Phase 1 **Study Objectives:** PRIMARY OBJECTIVES: I. To investigate efficacy. II. To determine the confirmed response rate of nab-rapamycin SECONDARY OBJECTIVES: I. To estimate other clinical outcomes II. To assess the adverse event profile III. To assess the clinical benefit IV. To assess progression-free survival and overall survival of these patients. TERTIARY OBJECTIVES: I. To assess quality of life and to correlate HRQOL/ symptoms with genomic markers. II. To assess the rate of individual mTOR pathway aberrations **Study design:** Interventional, Open label, Single group assignment **Study dates:** Study start date: January 2016; Study end date: April 24, 2018 **Study Status:** completed	**Arms:** 1. Nanoparticle albumin-bound rapamycin **Interventions** 1. Laboratory Biomarker Analysis 2. Quality-of-Life Assessment	**Brief Summary** This pilot study investigates clinical response to rapamycin administered to patients with advanced cancer and having abnormal genetic test results in a protein called mechanistic target of rapamycin (mTOR). Patients are given nanoparticle albumin-bound rapamycin, which may inhibit the growth of cancer cells by influencing the mTOR enzyme, required for cell growth. **Primary Outcome Measures** 1. Proportion of confirmed responses (clinical benefit) **Secondary Outcome Measures** 1. Incidence of adverse events 2. Relationship of adverse events to study treatment 3. Survival time 4. Time to disease progression **Other Outcome Measures** 1. Quality of life (EORTC QLQ-C30 questionnaire) 2. Rate of mTOR pathway aberrations

(*Continued*)

Table 9 (Continued).

III. A Study of CriPec® Docetaxel Given to Patients With Solid Tumours[170] (33 participants) Metastatic Cancer, Solid Tumors		
Selected Study Details	**Arms and Interventions**	**Brief description and Outcome Measures**
Official Study Title: A Phase I Open-Label, Safety, Pharmacokinetic and Preliminary Efficacy Study of CriPec® Docetaxel in Patients With Solid Tumours **Study Phase:** Phase 1 **Study Objectives:** To assess safety and tolerability **Study design:** Open Label, Single Group Assignment **Study dates:** Study start date: August 2015 Study end date: July 2018 **Study Status:** completed	**Arms:** 1. CriPec® docetaxel Docetaxel containing nanoparticle 3 weekly IV dose	**Brief Summary** The aim of this study is to determine safety by finding the highest safe dose of CriPec® docetaxel that can be administered to patients with solid tumours **Primary Outcome Measures** 1. Incidence of adverse events (grade 3 or 4) as a measure of safety and tolerability. 2. Incidence of abnormal clinical laboratory values as a measure of safety and tolerability. 3. Incidence abnormal of electrocardiogram findings as a measure of safety and tolerability. 4. Incidence of adverse events at the Maximum Tolerated Dose (grade 3 or 4) 5. Incidence of abnormal lab values at the Maximum Tolerated Dose 6. Incidence of ECG abnormalities at the Maximum Tolerated Dose 7. Pharmacokinetics: (Tmax), (Cmax), volume of distribution (Vd), half life (t1/2), total body clearance (CL) and area under the concentration-time curve (AUC) **Secondary Outcome Measures** 1. Early signs of anti-tumor efficacy (overall response rate) 2. Early signs of anti-tumor efficacy (duration of response)

IV. PIPAC Nab-pac for Stomach, Pancreas, Breast and Ovarian Cancer[171] (20 participants) Ovarian Cancer Stage: IIIB, IIIC, IV; Breast Cancer Stage: IIIB, IIIc, IV; Stomach Cancer; Stage III & Stage IV with Metastases; Pancreas Cancer, Stage IIIB Stage IV		
Selected Study Details	**Arms and Interventions**	**Brief description and Outcome Measures**
Official Study Title: Intraperitoneal Aerosolization of Albumin-stabilized Paclitaxel Nanoparticles for Stomach, Pancreas, Breast and Ovarian Cancer **Study Phase**: Phase 1 **Study Objectives:** To assess maximal tolerated dose via dose escalation combined with pharmacokinetic/pharmacodynamic modelling which incorporates, in addition to plasma, tumor tissue, and peritoneal drug concentrations, biomarkers of toxicity and efficacy **Study design:** Interventional, Randomized, Single Group Assignment, Double blinded, Multicenter **Study dates**: Study start date: September 16, 2017; Study end date: May 6, 2020 **Study Status**: completed	**Arms:** 1. PIPAC with Abraxane (35 mg/m²) 2. PIPAC with Abraxane (70 mg/m²) 3. PIPAC with Abraxane (90 mg/m²) 4. PIPAC with Abraxane (112,5 mg/m²) 5. PIPAC with Abraxane (140 mg/m²)	**Brief Summary** The study is designed to assess the maximal tolerated dose of albumin bound nanoparticle paclitaxel (nab-pac) administered with repeated pressurized intraperitoneal aerosol chemotherapy **Primary Outcome Measures** 1. Maximally tolerated dose of Abraxane (dose limiting toxicities) **Secondary Outcome Measures** 1. Surgical morbidity will be measured (Dindo-Clavien classification) 2. Maximum plasma concentration of Abraxane 3. Area Under the Curve (AUC) 4. Pharmacodynamics of Abraxane will be analyzed using tumor markers (CA15.3 for breast cancer, CEA for stomach cancer, CA19.9 for pancreatic cancer, CA125 in case of ovarian cancer). 5. Pharmacodynamics (PD) of Abraxane will be analyzed by tumor biopsies 6. Quality of Life (EORTC QLQ-C30 questionnaire) 7. Quality of Life (FACT-G questionnaire) 8. Neutropenia (neutrophil count) 9. Decreased platelet count

(*Continued*)

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Table 9 (Continued).

V. Ceritinib and Combination Chemotherapy in Treating Patients With Advanced Solid Tumors or Locally Advanced or Metastatic Pancreatic Cancer[172] (38 participants) Advanced Malignant Solid Neoplasm; ALK Positive Metastatic Pancreatic Adenocarcinoma Pancreatic Cancer Stage III & Stage IV		
Selected Study Details	**Arms and Interventions**	**Brief description and Outcome Measures**
Official Study Title: A Phase I Study of Ceritinib (LDK378), a Novel ALK Inhibitor, in Combination With Gemcitabine-Based Chemotherapy in Patients With Advanced Solid Tumors **Study Phase:** Phase 1 **Study Objectives:** PRIMARY OBJECTIVES: I. Determine the maximum tolerated dose and recommended Phase II dose of ceritinib in combination with gemcitabine (gemcitabine hydrochloride) alone, gemcitabine/nab-paclitaxel and gemcitabine/cisplatin in patients with advanced solid malignancies. SECONDARY OBJECTIVES: I. Assess the safety profile II. Determine the pharmacokinetic characteristics III. Determine the preliminary efficacy of the study drug combinations. TERTIARY OBJECTIVES: I. Investigate potential biomarkers of efficacy **Study design:** Interventional, Non-randomized, Open label, Parallel assignment **Study dates:** Study start date: January 8, 2015; Study end date: February 12, 2019 **Study Status:** completed	**Arms:** 1. Ceritinib maximum tolerated dose (MTD) then gemcitabine alone **Interventions 1:** Drug: Ceritinib Drug: Gemcitabine Hydrochloride Other: Laboratory Biomarker Analysis Other: Pharmacological Study 2. Ceritinib maximum tolerated dose then with gemcitabine and paclitaxel albumin-stabilized nanoparticle **Interventions 2:** Drug: Ceritinib Drug: Gemcitabine Hydrochloride Drug: Paclitaxel Albumin-Stabilized Nanoparticle Formulation Other: Laboratory Biomarker Analysis Other: Pharmacological Study 3. Ceritinib maximum tolerated dose then with gemcitabine and cisplatin **Interventions 3:** Drug: Ceritinib Drug: Cisplatin Drug: Gemcitabine Hydrochloride Other: Laboratory Biomarker Analysis Other: Pharmacological Study	**Brief Summary** Study to determine safety - the maximum tolerated dose and recommended Phase II dose for chemotherapy with Ceritinib (LDK378) in patients with advanced solid tumors **Primary Outcome Measures** 1. Maximum tolerated dose and recommended Phase II dose of ceritinib in combination with gemcitabine hydrochloride alone 2. Maximum tolerated dose and recommended Phase II dose of ceritinib in combination with gemcitabine hydrochloride and cisplatin 3. Maximum tolerated dose and recommended Phase II dose of ceritinib in combination with gemcitabine hydrochloride and paclitaxel albumin-stabilized nanoparticle formulation **Secondary Outcome Measures** 1. Incidence of adverse events of Ceritinib in combination treatment with gemcitabine hydrochloride chemotherapy Safety profile based on event type, frequency, severity, time relationship, seriousness and relationship to study treatment. 2. Pharmacokinetics of Ceritinib and Gemcitabine hydrochloride combined: A population based pharmacokinetic model to estimate individual AUC or clearance of Ceritinib 3. Pharmacokinetics of Ceritinib, Gemcitabine hydrochloride, and paclitaxel albumin-stabilized nanoparticle formulation; A population based pharmacokinetic model will be developed to estimate individual AUCs or CL of Ceritinib in combination with Gemcitabine hydrochloride and nab-Paclitaxel. 4. Pharmacokinetic characteristics of paclitaxel albumin-stabilized nanoparticle formulation, and cisplatin; A population based pharmacokinetic model will be developed to estimate individual AUCs or CL of Ceritinib in combination with Gemcitabine hydrochloride and Cisplatin. 5. Progression free survival 6. Response rate as estimated by the RECIST 1.1 **Other Outcome Measures:** 1. Tumor biomarkers and levels of serum

VI. Phase 1 Trial of PAN-301-1 (SNS-301) in Cancer Patients[173] (12 participants) Prostate Cancer		
Selected Study Details	**Arms and Interventions**	**Brief description and Outcome Measures**
Official Study Title: Phase 1, Open Label Trial to Evaluate the Safety and Immunogenicity of PAN-301-1 in Cancer Patients **Study Phase:** Phase 1 **Study Objectives:** **Study design:** Interventional, Sequential Assignment, Open label **Study dates:** Study start date: December 2016; Study end date: December 2018 **Study Status:** completed	**Arms:** 1. PAN-301-1 (SNS-301) Vaccine **Intervention:** Biological: PAN-301-1	**Brief Summary** Research phase 1 study of PAN-301-1 (SNS-301), a HAAH directed nanoparticle vaccine, given intradermally in cohorts of patients with biochemically relapsed prostate cancer, aiming to assess safety parameters. **Primary Outcome Measures** Safety assessment to determine maximum tolerated dose by monitoring the development of adverse events and dose-limiting toxicity **Secondary Outcome Measures** Safety assessment by monitoring administration site reactions, abnormal lab values and adverse events

(*Continued*)

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Table 9 (Continued).

VII. Gemcitabine Hydrochloride, Cisplatin, and Nab-Paclitaxel in Treating Patients With Advanced or Metastatic Biliary Cancer[174] (62 participants) Stage III Intrahepatic Cholangiocarcinoma AJCC v7 Stage IIIA Gallbladder Cancer AJCC v7 Stage IIIB Gallbladder Cancer AJCC v7 Stage IVA Gallbladder Cancer AJCC v7 Stage IVA Intrahepatic Cholangiocarcinoma AJCC v7 Stage IVB Gallbladder Cancer AJCC v7 Stage IVB Intrahepatic Cholangiocarcinoma AJCC v7 Unresectable Extrahepatic Bile Duct Carcinoma Unresectable Gallbladder Carcinoma		
Selected Study Details	**Arms and Interventions**	**Brief description and Outcome Measures**
Official Study Title: A Phase II Study of Gemcitabine, Cisplatin, and Abraxane in Advanced Biliary Cancers **Study Phase**: Phase 2 **Study Objectives:** PRIMARY OBJECTIVES: I. Determine the progression-free survival of gemcitabine hydrochloride (gemcitabine), cisplatin, and nab-paclitaxel in advanced, untreated biliary cancers. SECONDARY OBJECTIVES: I. Determine the response rate and disease control rate II. Determine overall survival of gemcitabine, cisplatin, and nab-paclitaxel in advanced biliary cancers. III. Evaluate the toxicity of gemcitabine, cisplatin, and nab-paclitaxel in advanced biliary cancers. **Study design:** Interventional, Single Group Assignment, Open - label **Study dates:** Study start date: April 2, 2015; Study end date: August 13, 2020 **Study Status:** completed	**Arms:** 1. Treatment (nab-paclitaxel, cisplatin, gemcitabine) **Intervention/treatment** Drug: Cisplatin Given IV Drug: Gemcitabine Hydrochloride *i.v.* Drug: Nab-paclitaxel *i.v.* Other: Laboratory Biomarker Analysis, Correlative studies	**Brief Summary** This study investigates the efficacy of the intervention drugs administered in patients with biliary cancers. **Primary Outcome Measures** Progression free survival **Secondary Outcome Measures** Incidence of adverse events

VIII. Radiosensitization of Multiple Brain Metastases Using AGuIX Gadolinium Based Nanoparticles[175] (15 participants) Brain Metastases		
Selected Study Details	**Arms and Interventions**	**Brief description and Outcome Measures**
Official Study Title: Phase I Clinical Study of Radiosensitization of Brain Metastases By Gadolinium Nanoparticles **Study Phase:** Phase 1 **Study Objectives:** To study safety and preliminary efficacy **Study design:** Interventional, open label, Single Group Assignment **Study dates:** Study start date: March 2016; Study end date: February 2019 **Study Status:** completed	**Arms:** 1. AGuIX and radiotherapy With the following escalation cohorts: 15 mg/kg, 30 mg/kg, 50 mg/kg, 75 mg/kg and 100 mg/kg **Intervention/treatment** Drug: AGuIX	**Brief Summary** Study investigates if AGuIX particles may increase the effectiveness of radiation therapy by sensitizing tumor cells to radiation. This trial studies the side effects and optimal dose of AGuIX when injected together with whole brain radiation therapy. The preliminary effectiveness of the combination of AGuIX and radiation therapy will be also assessed. **Primary Outcome Measures** 1. Maximum tolerated dose of AGuIX given with the whole brain radiation therapy **Secondary Outcome Measures** 1. Pharmacokinetic parameter of AGuIX particles - Cmax 2. Pharmacokinetic parameter of AGuIX particles - AUC 3. Pharmacokinetic parameter of AGuIX particles -T1/2 4. MRI to evaluate distribution of AGuIX particles in brain metastases and surrounding healthy tissue 5. MRI to assess intracranial progression-free survival 6. Overall survival

(*Continued*)

Table 9 (Continued).

IX. Study to Evaluate CORT125134 in Combination With Nab-paclitaxel in Patients With Solid Tumors[176] (146 participants) Solid Tumors		
Selected Study Details	**Arms and Interventions**	**Brief description and Outcome Measures**
Official Study Title: Phase 1/2 Study of CORT125134 in Combination With Nab-paclitaxel in Patients With Solid Tumors **Study Phase:** Phase 1/2 **Study Objectives:** To assess the safety and to determine the preliminary efficacy **Study design:** Interventional, non-randomized, open label, Single Group Assignment, multicenter **Study dates:** Study start date: May 2016; Study end date: May 2020 **Study Status:** completed	**Arm** CORT125134 with nab-paclitaxel **Intervention/treatment** CORT125134 with nab-paclitaxel *p.o.* Nab-paclitaxel *i.v.*	**Brief Summary** The purpose of this study is to assess the safety and preliminary efficacy of the combination of drugs: CORT125134 and nab-paclitaxel administered in patients with solid tumors. **Primary Outcome Measures** 1. Maximum Tolerated Dose of CORT125134 **Secondary Outcome Measures** 1. Incidence of Treatment-Related Adverse Events **Other Outcome Measures** 1. Objective response rate, progression free survival, overall survival 2. Objective response rate, progression free survival, and overall survival in patients with GR-positive or GR negative solid tumors. 3. Pharmacokinetics: exposure-response 4. Pharmacodynamics: modulation of GR function, hormonal changes and FKBP5

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X. Study of Topical SOR007 Ointment for Cutaneous Metastases[177] (23 participants) Cutaneous Metastasis		
Selected Study Details	**Arms and Interventions**	**Brief description and Outcome Measures**
Official Study Title: Phase 1/2 Dose-Rising, Safety, Tolerability and Efficacy Study of Topical SOR007 for Cutaneous Metastases **Study Phase:** Phase 1/2 **Study Objectives:** To evaluate the safety, tolerability and preliminary efficacy of SOR007 (in different concentrations) **Study design:** Interventional, Non-Randomized, Sequential Assignment, open-label, dose-rising **Study dates:** Study start date: January 31, 2018; Study end date: April 29, 2020 **Study Status:** completed	**Arms:** 1. SOR007 0.15% (Uncoated Nanoparticle Paclitaxel) Ointment 2. SOR007 1.0% (Uncoated Nanoparticle Paclitaxel) Ointment 3. SOR007 2.0% (Uncoated Nanoparticle Paclitaxel) Ointment **Intervention** Drug: Uncoated Nanoparticle Paclitaxel) Ointment	**Brief Summary** To study a topical nanoparticle paclitaxel ointment (SOR007) for the treatment of cutaneous metastases from non-melanoma cancer in adults. **Primary Outcome Measures** 1. Incidence of treatment emergent adverse events **Secondary Outcome Measures** 1. Difference in total area of eligible lesion(s) in the treatment area 2. Objective clinical response 3. Reduction in pain at the treatment area 4. Pharmacokinetic parameter - AUC 5. Pharmacokinetic parameter - Cmax 6. Pharmacokinetic parameter - Tmax

(*Continued*)

Table 9 (Continued).

XI. PET Study With [89Zr]-Df-CriPec® Docetaxel[178] (7 participants) Solid Tumor		
Selected Study Details	**Arms and Interventions**	**Brief Description and Outcome Measures**
Official Study Title: A Clinical Phase I, Open-label, PET Study With [89Zr]-Df-CriPec® Docetaxel in Patients With Solid Tumours to Assess Biodistribution and Tumour Accumulation of [89Zr]-Df-CriPec® Docetaxel **Study Phase:** Phase 1 **Study Objectives:** To assess biodistribution and tumour accumulation of the drug administered. **Study design:** Open Label, Single Group Assignment **Study dates:** Study start date: April 1, 2018; Study end date: May 8, 2020 **Study Status:** completed	**Arms:** 1. [89Zr]-Df-CriPec® docetaxel	Study with administration of [89Zr]-Df-CriPec® docetaxel in patients with solid tumours to assess biodistribution and tumour accumulation of the drug given. **Primary Outcome Measures** Detection (visual) of [89Zr]-Df-CriPec® docetaxel in tumour lesions Visual detection (absent/present) of tumor uptake Detection (quantitative) of [89Zr]-Df-CriPec® docetaxel in tumour lesions **Secondary Outcome Measures** Dosimetry of [89Zr]-Df-CriPec docetaxel based on activity concentration and biodistribution Optimal time point for PET imaging after [89Zr]-Df-CriPec® docetaxel administration Linearity between [89Zr]-Df-CriPec® docetaxel and total docetaxel Biodistribution of low dose dose [89Zr]-Df-CriPec® docetaxel before and after administration of therapeutic dose of CriPec® docetaxel.

XII. A Sunscreen Based on Bioadhesive Nanoparticles[179] (13 participants) Melanoma; UV Ray Skin Damage		
Selected Study Details	**Arms and Interventions**	**Brief description and Outcome Measures**
Official Study Title: Assessing the Safety and Efficacy of Multifunctional Skin-adhesive Nanoparticles for UV Protection in Humans **Study Phase:** Phase 1 **Study Objectives:** To evaluate the duration of protection and efficacy of a bioadhesive nanoparticle sunscreen **Study design:** Randomized, double blinded, parallel assignment **Study dates:** Study start date: July 17, 2017 Study end date: August 18, 2017 **Study Status:** completed	**Arms:** 1. UV filtering agent and bioadhesive nanoparticles (BNPs) 2. Standard Sunscreen consisting of padimate O (7%) and oxybenzone (3%). 3. Sham Comparator: A placebo strips with no UV filtering 4. Control, no agent applied.	**Brief Summary:** Study assesses the safety, sun protection factor (SPF) characteristics, and the duration of protection. **Primary Outcome Measures** Skin Reaction assessed by examination (evidence of irritation, inflammation, follicular occlusion).

(*Continued*)

Table 9 (Continued).

XIII. NU-0129 in Treating Patients With Recurrent Glioblastoma or Gliosarcoma Undergoing Surgery[180] (8 participants) Gliosarcoma; Recurrent Glioblastoma		
Selected Study Details	**Arms and Interventions**	**Brief description and Outcome Measures**
Official Study Title: A Phase 0 First-In-Human Study Using NU-0129: A Spherical Nucleic Acid (SNA) Gold Nanoparticle Targeting BCL2L12 in Recurrent Glioblastoma Multiforme or Gliosarcoma Patients **Study Phase**: Phase 0 **Study Objectives:** PRIMARY OBJECTIVES: I. To assess the safety of *i.v.* administration of NU-0129 SECONDARY OBJECTIVES: I. To assess serum drug concentration II. To verify intratumoral penetration of NU-0129. III. To verify the feasibility of administering NU-0129 as a standard treatment for recurrent glioblastoma multiforme or gliosarcoma. TERTIARY OBJECTIVES: I. To analyze Bcl2L12 expression levels II. Progression free survival and overall survival at 6 months; overall response rate. **Study design**: Interventional, Open Label, Single Group Assignment **Study dates:** Study start date: May 25, 2017; Study end date: August 19, 2020 **Study Status:** completed	**Arms:** 1. Experimental treatment (NU-0129) **Intervention:** 1. Laboratory Biomarker Analysis 2. Pharmacological Study	**Brief Summary:** The aim of this study is to evaluate the safety of the administered drug, NU-0129, (via application of nucleic acids arranged on the surface of a small spherical gold nanoparticle) in patients with recurrent glioblastoma multiforme or gliosarcoma. The researchers expect that targeting the Bcl2L12 gene with NU-0129 will stop cancer cells from growing. **Primary Outcome Measures** Incidence of Adverse Events **Secondary Outcome Measures** 1. Drug concentration in blood 2. Biodistribution of NU-0129 in tumor tissue (concentration of particles in various parts of the tumors). 3. Feasibility of administering NU-0129 as a standard treatment

changed the colloidal and structural properties of nanoparticles.[196]

Another challenge regarding the industrial transfer of nanodrugs is an insufficient number of guidelines for the characterization of nanoparticles concerning their safety and non-toxicity and lack of strict legal regulations.[188,197] Given the listed challenges, to obtain the desired features during the synthesis of drug formulations, the Food and Drug Administration (FDA) introduced in the 2000s a method of quality by design, which provides product quality controls at every stage of the process (by using pH or ionic strength sensors). In this way, the key parameters of the drug carrier synthesis must be obtained via standardized procedures and scalable chemical equipment. Since the synthesis conditions in the industrial plant are different from in the laboratory, each stage of the synthesis must be transferred according to Chemistry, Manufacturing and Controls (CMCs) and follow good manufacturing practice (GMP) requirements.[191,198] However, as it is not easy to control the process taking into account so many parameters that nanoparticles desire, a reproducibility problem arises.[195] As a consequence, each batch of produced material must be thoroughly tested to ensure its characteristics, safety, and non-toxicity.[190] Some researchers suggest that routine testing of large-scale formulations in animal models would be desirable.[191]

One may notice a deficit of simple industrial procedures for the synthesis of nanomaterials regarding the limited possibilities of the industrial plant.[199] Methods for the synthesis of nanomaterials described in the literature have many limitations, such as the difficult removal of a toxic organic solvent (in solvent emulsification-diffusion technique applied to lipids) or challenging maintaining sterility of the product.[185,191,200] Furthermore, some methods to produce nanoformulations, such as freeze-drying and spray-drying used to fabricate nanoencapsulates in powder form, are expensive and may affect particle size.[201] Large-scale preparation of nanocarriers that will be biodegradable in vivo is another challenge.[201,202] Therefore, top-down processes (consisting of mechanical fragmentation of the product) are still more common than the bottom-up approach (generating nanoparticles starting from molecules or atoms).[203] However, some production methods seem to be more useful than others for large-scale applications, such as supercritical reverse-phase evaporation or microfluidic mixers.[191,192]

Furthermore, as usually creating a new drug delivery system is a reformulation of a previously known drug, pharmaceutical companies often do not consider this process worth the time and costs compared to profits and prefer investing in the search for new drugs by simply screening libraries of small compounds.[188,199]

Green Synthesis in Drug Carriers Manufacturing

"Green nanomedicine" is a new field of drug delivery systems based on nanomaterials, which provides tools for more economical nanocarriers synthesis. However, currently, only a few literature examples of research can be found in which at least a few of the dozen "green chemistry" postulates have been met. Among syntheses of such drug carriers as nanometallic compounds, polymer nanocomposites, and quantum dots one can find examples of the use of safer reagents, solvents or auxiliaries, the design of safer, atom economical syntheses, application of renewable energy sources, or the synthesis of biodegradable carriers. Among the described nanosystems, protein and lipid compounds are the safest of known drug carriers.[204,205]

A very important aspect is the choice of the synthesis method among those available.[206] A separate group of non-toxic reactions in nanomedicine are methods that use plant extracts as reagents. For example, Palai et al described the synthesis of a decorated graphene nanocomposite, where the aqueous neem leaf extract was used to reduce graphene oxide, while the synthetic procedure was modified to reduce the number of toxic gases and impurities generated.[207]

One of the latest examples of the use of eco-friendly reagents was delivered by Uthappa et al, who described the green synthesis of natural diatoms modified with polydopamine as a drug delivery system, in which additionally the synthesis time was reduced and no toxic reagents and solvents were used.[87] Furthermore, Hasan et al described the eco-friendly synthesis of silver nanoparticles in which the reduction process by chemical compounds has been replaced by a reduction by a biopolymer (dextrin).[208] An alternative to green solvents may be the use of ionic liquids.[209]

Despite the existence of more adaptive techniques, such as reverse-phase evaporation or thin-film hydration, a green technique, energy-saving probe sonification method using only water as a solvent, was chosen for the production of niosomes by Khan et al.[113] Next, Ca^{2+} cross-linked Fe-guanosine monophosphate (Fe-GMP) hydrogel

for doxorubicin delivery was prepared by facile mixing of appropriate components at ambient conditions.[210] Finally, it is important to select those biocompatible from among the available polymers (poly(sodium 4-styrene sulfonate), PSS), and assure that the encapsulation of the drug takes place using a simple green method, for example, by mixing of two aqueous phases containing the polymer and the drug, respectively.[92] From the producer's point of view, the more "green" the process, the cheaper and safer for the final product due to the lack of toxic impurities.

Pharmacokinetic and Toxicological Studies of Nanoparticles as a Delivery System

Pharmacokinetics, often described as what an organism does to a drug, is a branch of pharmacology dealing with the study of the activity of compounds in the body over a period of time with a primary focus on processes by which medicinal products and drugs are absorbed, distributed, metabolized, and finally excreted (ADME). Pharmacokinetics depends on many factors that are related to the physicochemical properties of the complex substance as well as to patient-related conditions like gender, age, individual physiology, or genetics. Knowledge of pharmacokinetics is crucial for targeted and safe application of drugs to achieve the maximum therapeutic effect and the minimum risk associated with the occurrence of adverse effects.

An ideal drug should be highly specific concerning the pathologic processes and changes without any toxicity to healthy organs, tissues, or cells. The most desired properties of an active compound should directly lead to proper absorption and drug distribution, low metabolism, decent elimination, and low toxicity.

Pharmacokinetic key parameters used for defining and describing the ADME processes include bioavailability (by determining the area under the plasma concentration–time curve), elimination half-life ($t_{1/2}$), the volume of distribution (Vd), and clearance (CL).[123] These factors play a crucial role in the determination of the concentration of the drug in the body at a specific therapeutic target. Pharmacokinetics is applied to estimate the exposure and the most important parameters used to define the optimal dosage form and the dosing regimen in clinical practice to achieve maximum efficacy and lowest toxicity.[211]

Pharmacokinetic Aspect of the Application of Nanoparticles as Delivery Systems

Drugs encounter many barriers in living organisms from the time of administration in a specific dosage form until the therapeutic molecules reach the target. Advances in technology allow us to make structural changes that make significant improvements in drug properties and help overcome the limitations of reduced drug efficacy and potential safety issues. Advances in nanotechnology over the past decades did revolutionize drug delivery systems by improving their pharmacokinetic and pharmacodynamic properties, such as higher solubility, duration of exposure, and targeted delivery to the site of action.[212]

The tabulation below briefly summarizes the main differences in pharmacokinetic properties of small drug molecules and the desirable drug-loaded nanoparticles (Table 10).

There are many different types of nanoparticles used as carriers for therapeutic compounds, as shown in Figure 6., each of them having different properties.

As mentioned in previous sections, nanoparticles differ in their surface charge, particle size and shape, efficiency, loading capacity, and stability, leading to substantial variability in pharmacologic effects and the safety of different nanocarriers. Petschauer et al summarize in their review the main factors affecting the pharmacokinetic (PK) and pharmacodynamic (PD) properties of anticancer carrier-mediated agents in patients.[213] The discussion includes the following elements: Uptake by the mononuclear

Table 10 Pharmacokinetic Properties Comparison Between Small Molecule Drugs and Drug-Loaded Nanoparticles

PK Property	Small Molecule Medicine	Drug-Loaded Nanoparticle
Volume of distribution	High	Low
Bioavailability (AUC)	Low	High
Circulation half time	Short	Long
Tumor accumulation	Poor	Good
Clearance	Rapid	Slow

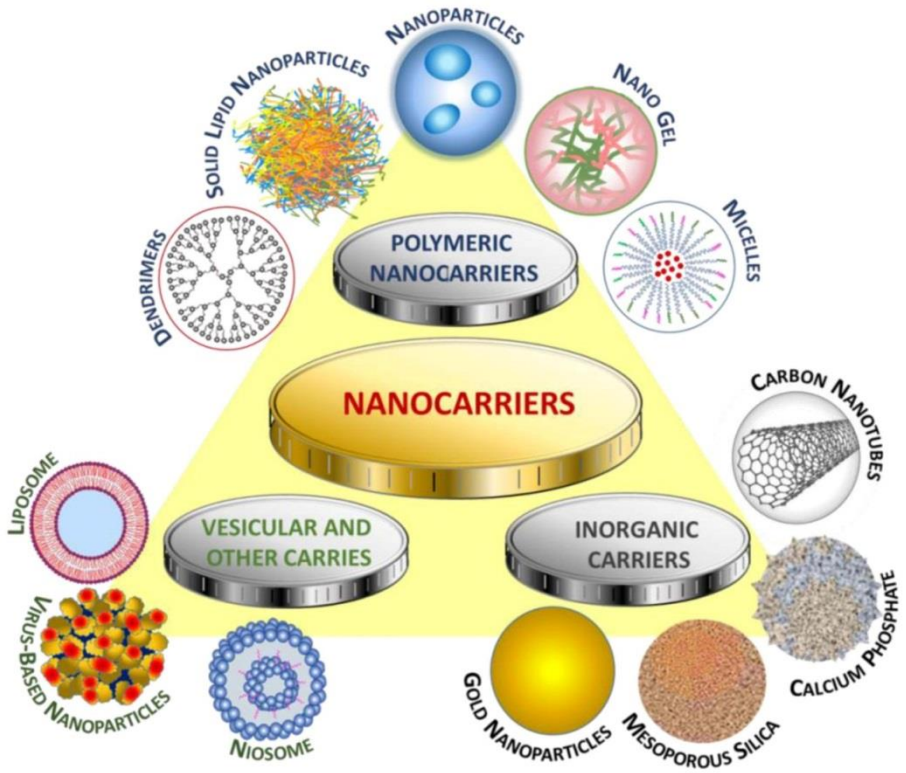

Figure 6 Classification of nanocarriers for drug delivery.

phagocyte system; Delivery of the compounds in tumors: nanoparticles (NPs) can get into tumors' tissue due to the leaky vasculature, which results in enhanced permeability and retention effect.; Particle size and shape: NPs between 100 and 200 nm have been observed to be most efficient in uptake by tumors; in turn, particles smaller than 50 nm showed short circulation time, and NPs greater than 300 nm prevented particles from taking advantage of the EPR effect, leading to lower tumor accumulation; Surface modification and charge (Conjugation of PEG to the surface of NPs increases circulation time and bioavailability – measured by Area Under The Curve – AUC; Uncharged particles have less mononuclear phagocyte system uptake, which results in longer circulation time); The concentration of NPs administered: a higher concentration level of particles per dose given increases the drug exposure in both plasma and tumor.

Besides, the authors stress the fact of the existence of a relationship between NP clearance and patient age, gender, disease conditions like liver or renal impairments, or concomitant medications. Another point to consider is the possibility to predict pharmacokinetic properties of PEGylated liposomal NPs based on the monocyte and dendritic cells function.

Advances in computational sciences over the past decade allow researchers to focus on mathematical and statistical approaches. Dogra et al describe a novel modeling approach aiming to predict whole-body nanoparticle pharmacokinetics and their tumor delivery.[214] The identified main factors governing NP kinetics in the tumor

interstitium were nanoparticle size, tumor vascular fraction, tumor vascular porosity, nanoparticle degradation rate, and tumor blood viscosity. Since the number of potential factors having an influence on the ADME processes in the living organism is huge by nature, mathematical modeling in this parameter space is proposed as an efficient alternative to traditional experiments.

The authors discuss the impact and particular values of parameters to optimize the delivery of NPs into tumor tissue. Garofalo et al present another methodology combining computer-aided drug design from the domain of computational chemistry and drug delivery techniques.[215] The multidisciplinary approach gives promising results in overcoming some of the main challenges, such as poor selectivity for the target or poor ADME properties. The authors discuss selected applications of the new approach, aiming to provide insights into a novel rational design of anticancer therapies. According to the authors, the computer-aided drug delivery system design should be combined with "wet" laboratory techniques that allow better prediction of drug delivery systems in vivo and helps in designing drug molecules that increase therapeutic targeting and reduce the optimal dosage.

Despite the fact that nanoparticles demonstrate excellent potential as drug delivery agents, the nano-protein interaction and the formation of a protein corona have been found to interfere with the nanoparticle delivery. In recently published studies, Zhang et al provided a brief summary of the latest developments on the nano-protein interactions between NPs and enzymes of the digestion and initiated an engaging discussion on the possibility of the use of the digestive enzyme corona for the targeted delivery in the colon.[216] The authors described physicochemical properties that are closely linked to the oral absorption of NPs, which include: size, zeta potential and surface molecules, which are greatly affected by the interaction of nano-enzymes and the formation of the enzyme corona. Moreover, it has been shown that the uptake of NPs by epithelial cells is significantly increased after the formation of the enzyme corona. The interaction of nano-enzymes is thus a major challenge for oral delivery of NPs and might exert an impact on pharmacological properties. On the other hand, a nano-enzyme interaction could also be applied to advanced oral delivery. As epithelial absorption of NPs is inhibited by the enzyme corona, a great number of NPs have a high chance of passing into the colon in the form of the NP-corona complex. After that, inside the colon, the enzyme corona and indeed NPs could be degraded and metabolized throughout the greatest microbiota in the organism, resulting in the release of loaded drugs straight into the colon area. The same problem has been previously discussed by Peng et al.[217] They synthesized the cationic NPs (CNPs) based on poly (3-hydroxybutyrate-*co*-3-hydroxyhexanoate) and examined the interaction of CNPs with digestive enzyme and its impact on cellular uptake. Author's results show for the first time the formation of the enzyme corona and its inhibitory effect on CNP uptake by epithelial cells. In another paper, Peng et al assessed the interaction between proteins and nanomaterials, which results that, in the in vivo performance of nanomaterials, are significantly different from these in vitro. It has been shown that the protein–nanomaterial interaction may induce remarkable changes in the properties of nanomaterials as well as their associated proteins.[218] These changes in properties will eventually lead to undesirable outcomes, which include: 1. Fast clearance of the bloodstream owing to opsonin adsorption; 2. Capillary blockage risk from the increased size after adsorption of serum proteins; 3. The loss of ability to target due to the original surface ligand being covered by the protein corona; 4. Possible toxicity due to the change in conformation of bound proteins. On the one hand, the above interactions are a major challenge for the safe and effective use of nanomaterials in clinical way, but, on the other hand, these interactions could be the possibility of decorating nanomaterial-based drug delivery systems. Consequently, in vivo transport and subsequent behavior of the protein-nanomaterial complex is much more controlled and indeed such a complex holds greater promise for being transferred to the practical products. In effect, it could be supposed that in the near future, these new smart products will be on the market for clinical use.

Toxicity of Drug Delivery Systems

Toxicity remains a challenge even when applying nanoparticles as drug carriers. Highly complex interactions between the molecules, cells, and the host environment are influenced by nanoparticles with many questions arising concerning their long-term safety.

Khan et al describe some of the potential NPs toxicities, which depend on various factors and types of particles used.[219] One of them, as pointed out by the authors, is the ability to organize around the protein concentration. This particular feature depends on particle size, curvature, shape and surface charge, functional groups, and free energy. Based on these properties, there is at least

a theoretical possibility for NPs to generate adverse and unexpected outcomes through protein unfolding, crosslinking, or causing loss of enzymatic activity.

It becomes evident that despite the promising results and improvement of pharmacokinetic properties of anticancer drug-loaded NPs, long-term research and further studies must be rolled out to better understand complex interactions at the molecular level in vivo.

The Problems of Nanotechnology in Practical Use. The Limitations and Concerns of Different Types of Nanoparticles for Drug Delivery Applications

In view of this paper, the use of nanotechnology in practice may face some challenges. The biggest concern is that the health and safety implications of the specific properties of nanoparticles have not yet been addressed by the regulatory authorities. The new European chemicals policy REACH does not consider side effects. Nanoparticles raise a number of safety and regulatory issues that governments are now beginning to address. A review of recent regulations and ongoing monitoring by authorities is necessary.[220] Moreover, some problems such as toxicity demonstrated by some nanoparticles cannot be overlooked when considering the application of nanomedicine in routine clinical practice. Recently, nanoparticles are mostly used together with natural products to reduce toxicity problems. The green chemistry pathway in the design of drug-containing nanoparticles is being extensively promoted due to the fact that it minimizes harmful components in the process of biosynthesis. Therefore, the use of "green" nanoparticles for delivering drugs can potentially reduce the side effects of drugs.[189]

The use of an optimal nanoparticle drug delivery system is mainly determined by the biophysical and biochemical properties of the targeted drugs that are selected for treatment and could help to improve the successful delivery of nanosystems and optimize the pharmacoeconomic impacts.[189]

Recently, various nanotechnology-based solutions for drug delivery in the field of medicine have attracted great interest. Despite the above, unfortunately there are still many concerns about the safety application of nanoparticles as drug delivery systems.[221]

Studies carried out on nanotechnology have proven that every type of nanoparticles has some limitations in practical use. The NPs' toxic effects are in general associated with the poor biocompatibility of the nanomaterials that were used to develop them. Carbon nanotubes (CNTs) are the type of NPs with more toxic potential observed. They have been found to be lung carcinogenic, but they are also toxic to CNS, blood and GIT. Heavy metals may accumulate in the liver and kidneys and can be toxic to the CNS and GIT. Silicates also have a significant potential to accumulate in the liver and lungs, leading to fibrosis. Direct toxicity of liposomes may be caused primarily by their size, charge or composition. For instance, cationic liposomes may interact with lipoproteins, serum proteins or even with the extracellular matrix, resulting in aggregation or release of agents that are loaded before they reach target cells, causing the systemic toxicity. At doses much higher than those administered (multiple injections of ≥100 mg/kg lipid), liposomes have been demonstrated to cause RES impairment, granulomas, hepatomegaly or even splenomegaly. Furthermore, the increase in lipid dose has been demonstrated to deplete plasma of different proteins. While the identification and importance of all deleted proteins remain unclear, it is possible that their loss will cause impairment in normal homeostasis. Metallic NPs could lead to peribronchitis, granulomas, interstitial fibrosis, collagen deposition, adenocarcinoma and pleural lesions. Nanoemulsions could be responsible for interference with the close linkage in GIT and direct cytotoxicity. Carbon NPs exhibit the oxidative stress, depletion of glutathione, an increase in the number of dermal cells, and also thickening of the skin and rash. Dendrimers and gold nanoshells demonstrate toxicity induced by macrophages, plasma protein depletion, aggregation of platelets and also their pathway of synthesis is complicated.[222,223]

In view of the above, the awareness of particle levels that may cause health effects is imperative for both workers and exposed patients.[222]

Challenges in Pharmacoeconomic Aspects of Nanocarriers as Drug Delivery Systems

Nanomedicine adopts the use of nanotechnology for highly specific medical interventions for the prevention, diagnosis and treatment of diseases, all of which are presented in this paper. The development of nanomedicines tends to improve the therapeutic efficacy, reduce the dose that is therapeutically effective, and decrease the risk of developing side effects.[224] Nanocarriers as DDS are

designed to reduce the cost of administering the drug, improve the compliance and help patients to recover as soon as possible. All of these aspects are reflected in pharmacoeconomics, a discipline that aims to provide reliable information on the cost of therapies and to choose the best one, considering its effectiveness at the lowest possible costs. In the above paper, nanotechnology solutions and standard therapies, their costs and effectiveness were discussed.[224]

The clinical development of nanomedicine encompass many aspects, there are some key issues to look out for: biological development (appropriate in vivo structural stability of the nanomedicine after application); process of manufacturing (production on a large scale according to GMP standards, which includes: reproducibility, techniques, infrastructure, experience, and costs of the whole process; tests used to control the quality for characterization which includes: charge, size, morphology, dispersion, encapsulation, modification of the surface, stability and purity); biocompatibility and safety concerns (development of much more targeted toxicoassays for nanomedicines; appropriate understanding of nanocarriers interactions with cells and tissues; reduced level of nanoparticles accumulation in targeted cells, tissues or organs); intellectual property (understand of the nanomedicine patent complexity); government regulations (development of clear nanomedicine regulatory guidelines); and total cost-effectiveness compared to standard treatment regimens (restricted understanding of the nanomedicine's biological interactions with the patient's biological environment, leading to an impossibility to apply a pharmacoeconomic approach).[225–229]

Such determinants could be substantial obstacles that limit the market emergence of nanomedicines, despite their therapeutic efficacy.

Conclusion

The availability of evidence resulting from the application of pharmacoeconomics can be useful in health policy decision-making. It can be applied by healthcare professionals such as policymakers, primary healthcare providers, health-care administrators, and health managers.

Pharmacoeconomics can certainly help in decision-making when evaluating the affordability and access to the right medication for the right patient at the right time, comparing alternative drugs from the same therapeutic class or drugs with a similar mechanism of action, and establishing accountability that the claims by a manufacturer regarding a drug are justified. Proper application of pharmacoeconomics will allow the pharmacy practitioners and administrators to make better and more informed decisions regarding the products and services.

Based on the published literature, the engagement of nanoforms at different stages, including prevention, diagnosis, and treatment, might provide significant benefits from an economic as well as treatment perspective. Those include but are not limited to the factors like faster diagnosis, increase in the viability of patients during antitumor therapy, overcoming the mechanisms of resistance in neoplastic cells, or enhancing therapeutic efficacy via synergistic or additive interactions.

The use of drug nanocarriers is a unique opportunity for an economically attractive improvement of known drugs because the development of novel nanoformulations is much cheaper and faster than the discovery of new drugs. Despite higher production costs, greater regulatory hurdles and difficult industrial transfer, they are worth the cost due to their numerous advantages.

Nanoencapsulation can increase the bioavailability of poorly soluble drugs, facilitate access to pathologically altered sites by improving the crossing of biological barriers and increased selectivity of drug-loaded nanocarriers, provide better storage and in vivo stability, and enable slow, controlled release of the drug in the human body. From a pharmaceutical and economic point of view, all of these benefits can reduce dose and associated toxicity, dosing frequency, side effects and costs, improve formulation, protect patents, and enhance patient compliance. In addition, the use of drug nanocarriers has found wide application in theranostics.

However, there are doubts about the use of drug carriers regarding the risks associated with the excess of nanocarriers used, such as high blood pressure or systemic toxicity. These side effects can be countered by selecting an appropriate carrier material as well as proper drug-carrier binding to ensure a low drug-to-carrier ratio in the formulation. Furthermore, changes in the stability and toxicity of carriers associated with industrial production can be avoided due to carefully optimized synthesis, including product control at every stage of its production, as well as the preparing guidelines for the synthesis of nanomaterials.

It is impossible to say which is better: discovering new drug nanocarriers or searching for new, more effective active substances. But surely to improve the well-known

drugs with serious side effects through the use of their nanoformulations is very desirable and cost-effective from the point of view of pharmacoeconomics.

Nowadays, nanotechnology has many advantages, which includes: great bioadhesive properties, high biocompatibility, low toxicity, high encapsulation efficiency and also great drug-loading capacity. Analyzing the above features of nanoformulations it can be concluded that nanoparticles hold a huge potential as drug delivery systems, imaging agents and also in phototherapy. Despite these advantages, there still remain many issues that need to be resolved before nanoparticles can be used in a safe and comprehensive clinical way. Some aspects need further studies, such as: to the generation of nanoparticles with desired sizes; control of the thickness of each layer of the nanoparticles and the impact of it on the therapeutic efficacy; development of more stable nanoparticle; optimization of the drug release profile from nanomaterials; presently, the release rates differ significantly and depend on how drugs are integrated into nanomaterials (mostly by: surface adsorption, conjugation or encapsulation); safety and clinical applications; their biodistribution and long-term toxicity profile. Finally, to understand the mechanisms for metabolism, accumulation and biodegradation of nanoparticles in in vivo studies; discover interactions of nanoparticles with other materials, substances, drugs, and living organisms.

Furthermore, current studies have been limited to the in vitro stage and do not show in-depth toxicology and pharmacokinetic parameters. However, as time passes, the science and publication aspects are broadening, and we are getting much more data that allow us to evaluate and predict the effects of the nanocarriers formulations.

Moreover, from a manufacturing point of view, optimization of the synthesis parameter, encapsulation efficacy, and improved stabilization of nanoproducts will also provide a better understanding of their mode of action and potentially predict the risks of eventual use. In effect, it might be a substantial achievement in reducing the direct and indirect costs of therapy.

Undoubtedly, implementation of new therapeutic options, such as nanotherapy, will be associated with to date still unknown risks; however, expansion and development of the currently performed studies will consequently eliminate the existing gap in our knowledge and understanding of relevant mechanisms when applying nanotechnology to drug development and related costs.

Given the above, more attention should be paid to the pharmacoeconomic aspects of the nanocarriers, to properly assess the risk and benefit balance of the very promising technology presented in this review.

Abbreviations

5-FU, 5-Fluorouracil; p53, cellular tumor antigen p53; ABCB1, ATP-binding cassette subfamily B member 1; ACUPA, Acid-based protein-ligand; ADME, Absorption, distribution, metabolism, and excretion; AEs, adverse events; AgNPs, silver nanoparticles; AGuIX NPs, Polysiloxane gadolinium-chelates based nanoparticles; ALK, Anaplastic lymphoma kinase; ALT, Alanine aminotransferase; ANG 1005, Angiopep-2 paclitaxel conjugate; AST, Aspartate transaminase; ATC, Anatomical Therapeutic Chemical Classification System; ATC, Anaplastic thyroid cancer; AUC, Area under the curve; BBB, blood–brain barrier; BCL2L12, BCL-2-related proline-rich protein; BCRP, Breast cancer resistance protein; BiOBr NPs, Polyethylenimine grafted bismuth oxybromide nanoplates with Fe^{3+}; BNPs, bioadhesive nanoparticles; BSA, Bovine serum albumin; C1-PNPs, pyridinium amphophile-loaded PLGA (poly (lactic-co-glycolic acid) nanoparticles; CA, Carcinoma antigen; CBA, Cost-benefit analysis; CCNG1, Cyclin-G1; CD NPs, Cyclodextrin nanoparticles; CEA, Cost-effectiveness analysis; Chk1, checkpoint kinase 1; CL, Clearance; CMA, Cost-minimization analysis; Cmax, the maximum concentration; CMCs, chemistry, manufacturing and controls; CNFs, carbon nanofibers; CNS, Central nervous system; CNTs, carbon nanotubes; CPSNPs, calcium phosphosilicate nanoparticles; CR, complete clinical response; CRC, colorectal cancer; CS@MoS_2, Chitosan-assisted MoS_2; CSC, cancer stem cells; CTC, circulating tumour cell; CTCAE, Common Terminology Criteria for Adverse Events; CUA, cost-utility analysis; CuS NPs, copper sulfide nanoparticles; DDSs, drug delivery systems; DKK4, Dickkopf WNT signaling pathway inhibitor 4; DMSO, Dimethyl sulfoxide; DNA, Deoxyribonucleic acid; DOX, Doxorubicin; DPYD, Dihydropyrimidine dehydrogenase; DLT, Dose-limiting toxicity; dvPtNPs, dualvalent platinum nanoparticles; ECG, Electrocardiogram; EGF, Epidermal growth factor; EORTC QLQ-C30, The European Organisation for Research and Treatment of Cancer Quality of Life 30-item questionnaire; EPR effect, enhanced permeability and retention effect; Eudragit S100, anionic copolymers based on methacrylic acid and methyl methacrylate; FACT-G, functional Assessment of Cancer Therapy – General questionnaire; FDA, Food and Drug

Administration; FFPE, Formalin-fixed paraffin embedded; FKBP5, FKBP Prolyl Isomerase 5; FR, Folate receptor; GBM, Glioblastoma multiforme; GC, Gastric cancer; GIT, Gastrointestinal tract; GMP, Good manufacturing practice; GNR@LDH-PEG NPs, Poly (ethylene glycol) (PEG) modified core-shell GNR@ (gold nanorod) layered double hydroxide nanoparticles; GNSs, Gold nanospheres; GO, Graphene oxide; GR, Glucocorticoid receptor; GS, Glioblastoma stem-like cell cultures; GSH, Glutathione; GSH-AgNPs, Glutathione-stabilized silver nanoparticles; Gy, Gray (SI unit); HA, Hyaluronic acid; HAAH, Human aspartyl (asparaginyl) β-hydroxylase; (HA)-PBCA-NPs, hyaluronic acid coated poly(butyl cyanoacrylate) nanoparticles; HALMD, hydrophilic and aromatic low molecular weight drugs; HER2, human epidermal growth factor 2; HIF-1α, Hypoxia-inducible factor 1-alpha; HO-GC, hydrotropic oligomer-conjugated glycol chitosan; HPG, Hyperbranched polyglycerol; HPG-C10-PEG, hyperbranched polyglycerol-C10- poly(ethylene glycol); ICER, Incremental cost-effectiveness ratio; ICPMS, inductively coupled plasma mass spectrometry; IONPs, Iron oxide nanoparticles; *i.v.*, intravenosa; MDCK-MDR1, Madin darby canine kidney (MDCK) cells with the MDR1 gene; MDM2, mouse double minute 2 homolog; MDR, Multidrug resistance; MDR1, Multidrug-resistant 1 transporters; $Mg(OH)_2$ NPs, magnesium hydroxide nanoparticles; MIC, Minimum inhibitory concentration; MnO_2 NPs, Manganese oxide nanoparticles; MNPs, magnetic nanoparticles; MoS_2/PDA-RGD, molybdenum disulfide/polydopamine-arginine-glycine-aspartic acid; mPEG-CHO-chitosan NPs, Methoxypoly (ethylene glycol) conjugated chitosan nanoparticles; MPS, Mononuclear phagocyte system; MRI, Magnetic resonance imaging; MRSA, Methicillin-resistant *Staphylococcus aureus*; MTD, The maximum tolerated dose; MTT, The (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) tetrazolium reduction assay; mTOR, mechanistic target of rapamycin; NaCl, sodium chloride; NCI's CTCAE, The National Cancer Institute Common Terminology Criteria for Adverse Events; NCs, Neocarzinostatin; NCQDs, Nitrogen-doped carbon quantum dots; NIH, National Institutes of Health; NIR, Near-infrared irradiation; NK 105, Paclitaxel-incorporating micellar nanoparticle formulation; NLCs, nanostructured lipid carriers; NPs, Nanoparticles; NRS-11, The Numeric Rating Scale; ORR, Overall response rate; OSA, Octyl-modified bovine serum albumin; OS, Overall survival; PBCA-NPs, Poly(butyl cyanoacrylate) nanoparticles; PCE, Tetrachloroethene; PCL, Poly(ε-caprolactone); PCMB-Dox NPs, PEGylated carborane-conjugated amphiphilic copolymer doxorubicin nanoparticles; PD, Pharmacodynamics; PD, Progressive disease; PDA-PEG-Van NPs, Polydopamine-based nanoparticles modified with PEG and vancomycin; PDCs, Polymer-drug conjugates; PDL1, programmed death-ligand 1; PDMAEMA, Poly(N-(2-(dimethylamino)ethyl)-methacrylate); PEG, Poly(ethylene glycol); PEG, Polyaspartate micellar nanoparticles; PEG-PCL, Poly (ethylene glycol)-poly(ε-caprolactone); PEG-PE, Poly (ethylene glycol)phosphatidyl ethanolamine; PEI-C18-HPG, polyethyleneimine (PEI)-C18-HPG; PET, Positron emission tomography; PFS, Progression free survival; PG, Pharmacogenomic; P-gp, permeability glycoprotein; pH, potential of hydrogen; PIPAC, Pressurized intraperitoneal aerosol chemotherapy; PK, Pharmacokinetics; PLA, Polylactide; PLD, PEGylated liposomal doxorubicin; PLGA, Poly(lactic-*co*-glycolic acid); PLGA-PEG, Poly(lactic acid-*co*-glycolic acid)-poly(ethylene glycol); PLMB-Dox NPs, Doxorubicin-loaded carborane-conjugated polymeric nanoparticles; PMMA, Poly(methyl methacrylate); *p.o.*, Per os; PR, Partial response; PSMA, Prostate-specific membrane antigen; PSS, Poly(sodium 4-styrene sulfonate); PST-PTX, Poly(styrene)-paclitaxel; PTX, Paclitaxel; RECIST, Response Evaluation Criteria in Solid Tumours; RES, Renal reticuloendothelial system; rGO-Au NPs, Reduced-graphene-oxide functionalized with gold nanoparticles; RNA, Ribonucleic acid; ROS, Reactive oxygen species; RP2D, The recommended Phase II dose (in clinical trials); SAB, *Staphylococcus aureus* bacteremia; SAE, Serious adverse event; *s.c.*, Subcutaneously; scFv, Single-chain fragment variable; SD, Stable disease; siRNA, Small interfering RNA; SLNs, solid lipid nanoparticles; SNA, Spherical nucleic acid; SNPs/SiNPs, silica nanoparticles; SPF, Sun protection factor; t1/2, Half-life; TDDSs, targeted drug delivery systems; TFAP2E, transcription factor AP-2 epsilon; TfR 1, transferrin receptor 1; Tmax, The time it takes to reach Cmax; Tri-Ag NPs, Citrate-coated triangular nanoparticles; Tv-Ag NPs, Toxicodendron vernicifluum silver nanoparticles; UPLC-MS/MS, Ultra-performance liquid chromatography–tandem mass spectrometry; UV, Ultraviolet; VAS, Vasectomy; VCAMs, vascular cell adhesion molecules; VEGFs, vascular endothelial growth

factors; Vd, volume of distribution; VOI, volumes of interest; QALYs, quality-adjusted life-years.

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Rozdział 10. Folic-acid-conjugated thermoresponsive polymeric particles for targeted delivery of 5-fluorouracil to CRC cells

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Folic-Acid-Conjugated Thermoresponsive Polymeric Particles for Targeted Delivery of 5-Fluorouracil to CRC Cells

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Abstract: Colorectal cancer is the fourth most common cancer worldwide and the third most frequently diagnosed form of cancer associated with high mortality rates. Recently, targeted drug delivery systems have been under increasing attention owing to advantages such as high therapeutic effectiveness with a significant depletion in adverse events. In this report, we describe the biocompatible and thermoresponsive FA-conjugated PHEA-*b*-PNIPAAm copolymers as nanocarriers for the delivery of 5-FU. The block copolymers were obtained using RAFT (Reversible Addition–Fragmentation chain Transfer) polymerization and were characterized by methods such as SEC (Size Exclusion Chromatography), NMR (Nuclear Magnetic Resonance), UV–Vis (Ultraviolet–Visible), FT-IR (Fourier Transform Infrared) spectroscopy, and TGA (Thermogravimetric Analysis). Nanoparticles were formed from polymers with and without the drug-5-fluorouracil, which was confirmed using DLS (Dynamic Light Scattering), zeta potential measurements, and TEM (Transmission Electron Microscopy) imaging. The cloud points of the polymers were found to be close to the temperature of the human body. Eventually, polymeric carriers were tested as drug delivery systems for the safety, compatibility, and targeting of colorectal cancer cells (CRC). The biological evaluation indicated high compatibility with the representative host cells. Furthermore, it showed that proposed nanosystems might have therapeutic potential as mitigators for 5-FU-induced monocytopenia, cardiotoxicity, and other chemotherapy-associated disorders. Moreover, results show increased cytotoxicity against cancer cells compared to the drug, including a line with a drug resistance phenotype. Additionally, the ability of synthesized carriers to induce apoptosis and necrosis in treated CRC cells has been confirmed. Undoubtedly, the presented aspects of colorectal cancer therapy promise future solutions to overcome the conventional limitations of current treatment regimens for this type of cancer and to improve the quality of life of the patients.

Keywords: folic acid; colorectal cancer; targeted delivery; drug delivery systems; 5-fluorouracil; folate receptors; mitigator; thermoresponsive polymer; polymeric carriers; RAFT polymerization; PHEA-*b*-PNIPAAm; PNIPAAm; NIPAAm

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1. Introduction

Current medical statistics indicate that colorectal cancer (CRC) is a widespread disease with a high mortality rate. Rawla et al. reported, based on the GLOBOCAN database, that CRC is the third most deadly and fourth most frequently detected cancer in the world [1]. The analysis of the risk and occurrence of CRC presented by the Polish National Cancer Registry shows that approximately 1 in 23 men and 1 in 25 women will be diagnosed

with CRC in their lifetime [2]. The most common treatment combines chemotherapy, surgery, radiotherapy, and targeted therapy [3]. The current chemotherapy involves using 5-fluorouracil (5-FU), an effective drug in various solid tumors, including colorectal, neck, head, and breast cancers [4]. Unfortunately, the problem of resistance to this drug is emerging and much effort has been put into improving 5-FU-based therapy in recent years [5]. One of the methods is to use nanosized drug delivery systems (DDS) that transport the drug to cancer cells and minimize the cytotoxic effect against host cells via restriction of drug penetration and retention in healthy tissues [6]. Compared to a free drug, the drug entrapped in the nanocarrier is protected against enzymatic degradation in the digestive system, allowing the drug to penetrate the bloodstream and reach diseased tissues. To date, the most important known delivery systems for 5-fluorouracil are, e.g., solid lipid, chitosan, poly(lactic-*co*-glycolic acid) (PLGA), silica, polymeric, and folic-acid-conjugated nanoparticles. Their main advantages are enhanced cytotoxic effect and cellular uptake, better targeting efficiency, and prolonged drug release [6,7].

Polymeric therapeutics are an important class of drugs in which the polymer performs an integral role, providing an effective encapsulation, controlled delivery, and drug release [8]. Furthermore, polymers are a class of chemical compounds with advantages such as tunable composition, geometry, size, and ease of derivatization. The latter feature allows attachment to the system ligands for targeting tissues or cells [9]. Drug loading methods include the generation of hydrolytic or acid-sensitive linkage, complexation, and ion pair or permanent bond formation, ensuring the release/action of the drug in the target place [10]. One of the advantages of nanoscale drug carriers is their ability to uptake into tumor sites via the enhanced permeability and retention (EPR) effect. The abovementioned effect, discovered by Maeda et al., is based on a universal pathophysiological mechanism in which macromolecular compounds (above 40 kDa) such as polymeric drug carriers might be accumulated in the tumor area. This in turn leads to achieving targeted delivery and retention of the anticancer agent into tumor tissue. It should be emphasized, that EPR strongly depends on factors associated with the tumor location and vascularization degree as well as the physicochemical properties of the carrier and anticancer agent. In clinical practice, using the EPR effect is the main idea in the passive targeting of solid tumors in cancer nanotherapy [11,12].

It was proved that drug delivery technologies can enhance the health of patients. This is due to minimizing off-target drug accumulation, improving the delivery of a drug to the target site, and facilitating patient compliance [13,14]. Furthermore, the controlled release of the active substance through the use of nanoformulation may be helpful for creating a reduction in adverse events. This can result in fewer medical procedures, lead to lower staff costs, and provide a greater chance of remission [15].

It was established that a plethora of tumors overexpressed folate receptors (FR) on their surface [16]. Therefore, FR-targeting has become the basis of many therapeutic, diagnostic, and imaging methods in the treatment of cancer to enhance cancer cells' uptake of drug-loaded vehicles. Known strategies of active targeting rely on folic-acid-modified anticancer drugs, antibodies, or antibody–drug conjugates, as well as drug delivery nanoplatforms. Folic acid was used as a targeting ligand because of its low price, availability, non-toxicity, lack of immunogenicity, and presence of functional groups that facilitate its chemical modification without compromising functionality [16,17]. For example, in our previous study, we showed that new polymeric drug carriers, poly(2-hydroxyethylacrylate)-*b*-poly(*N*-vinylcaprolactam) (PHEA-*b*-PNVCL) conjugated with folic acid, used as part of a combinatory treatment, showed improved cytotoxicity of 5-FU against colon cancer cells, while also playing a protective role for healthy tissues [18].

Thermoresponsive polymeric nanocarriers have been intensively studied over the last two decades. They have all of the advantages of drug delivery systems, such as protection against early drug degradation in the body, extended plasma half-life, and increased solubility of non-hydrophilic drugs. Moreover, they are sensitive to external stimuli, allowing for better drug release control. In aqueous solutions, such polymers may exhibit lower or upper

critical solution temperature (LCST or UCST). Poly(*N*-isopropylacrylamide) (PNIPAAm) is the most investigated thermoresponsive polymer that was found to have many biomedical applications, including tissue engineering [19–21]. As evidence, we recently showed the non-hemolytic nature of polymeric nanoparticles made of polymers containing the cholesterol moiety and PNIPAAm block [22]. Consequently, the biocompatible copolymers consisting of PNIPAAm and poly(2-hydroxyethylacrylate) (PHEA) blocks could provide a high-potential platform for drug delivery [23].

PNIPAAm-based folic-acid-functionalized polymeric drug carriers were demonstrated to be effective in the form of micellar nanoparticles in the delivery of anticancer drugs or model hydrophobic drugs. For instance, Razaei et al. reported that the targeted delivery of paclitaxel via a star-shaped amphiphilic block copolymer (4s[poly(*E*-caprolactone)-*b*-2s(poly(*N*-isopropylacrylamide-*co*-acrylamide)-*b*′-methoxy) poly(ethylene glycol)/poly(ethylene glycol)-folate]) was non-hemolytic and resulted in improved cellular uptake [24–29]. Another example is nanocapsules based on PNIPAAm (PNIPAAm-*co*-PMA) and externally functionalized with folic acid, which are promising carriers for doxorubicin [30,31]. Furthermore, magnetic nanoparticles or nanocomposites functionalized with a temperature-sensitive polymer and a folate ligand were tested as doxorubicin delivery systems [32–34]. Interestingly, in addition to drug transport, folate-conjugated biopolymer was used to deliver an enzyme that converts a non-toxic prodrug to 5-fluorouracil [35]. Dubé et al. described the synthesis of folate-conjugated copolymer based on NIPAAm and amino-*N*′-ethylenedioxy-bis(ethylacrylamide) [36]. Shin et al. proposed poly(*N*-isopropylacrylamide)-pyrrole-folate nanocomposites as doxorubicin carriers triggered by near-infrared light for chemo-photothermal cancer therapy [37].

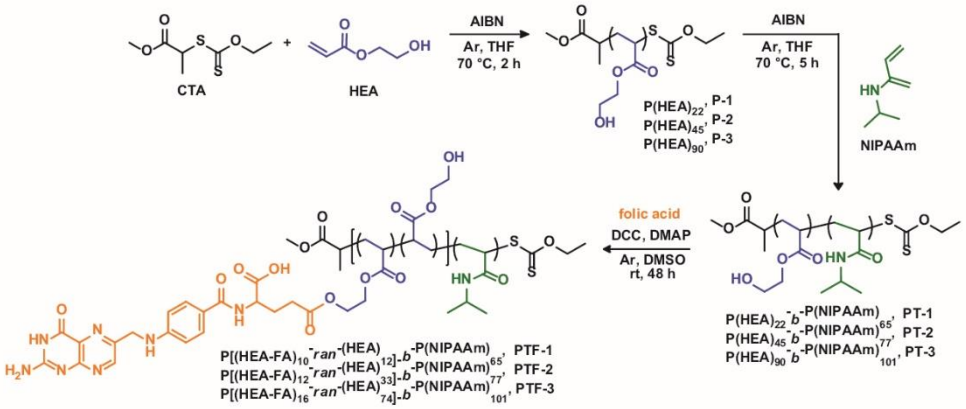
Herein we report the synthesis of the series of biocompatible and thermoresponsive folate-conjugated PHEA-*b*-PNIPAAm block copolymers using reversible addition–fragmentation chain transfer (RAFT) polymerization. The preparation of polymers with different ratios of PHEA to PNIPAAm block and different degrees of functionalization with folic acid allows an understanding of the influence of individual components on the formation of micellar structures, the cloud point values, and the strength of interaction with the drug. In consequence, it provides the design of the best drug delivery platform. A comprehensive physicochemical characterization of block copolymers and micelles with encapsulated 5-FU has been carried out. Furthermore, prepared drug carriers were subjected to biological evaluation. Their biocompatibility with representative host cells–red blood cells, monocytic cells, cardiomyocyte cells, skin, and colorectal fibroblast cells was tested. Moreover, cytotoxic activity against three colon cancer cell lines, including those resistant to chemotherapeutic agents, was investigated.

2. Results and Discussion

2.1. Synthesis and Characterization of PHEA and PHEA-b-PNIPAAm Polymers

PHEA-*b*-PNIPAAm copolymers were obtained from commercially available monomers and simple-chain transfer agent (CTA, methyl 2-((ethoxycarbonothioyl)thio)propanoate) in two steps. First, three poly(2-hydroxyethylacrylate)s (PHEA; P-1, P-2, P-3) of different molecular weights were synthesized by RAFT polymerization using an optimized reaction procedure (Scheme 1, Table S1). Next, chain extensions of each of the PHEA polymers were carried out using RAFT polymerizations of *N*-isopropylacrylamide (NIPAAm) (Scheme 1, Table S2). The obtained thermoresponsive resins (PT-1, PT-2, PT-3) differed in the length of the PHEA block, ensuring a different amount of hydroxyl groups were available for functionalization with folic acid. The polymers and block copolymers were characterized using ^{1}H NMR (Table 1, Figures 1 and S1–S3), FT IR (Figures 2 and S4), UV–Vis (Figures S5 and S6), and SEC (Table 1, Figure S7). The results obtained either from NMR or SEC indicate obtaining PHEA homopolymers with molecular weights close to the theoretical ones, growing in a row **P-1**, **P-2**, **P-3** (Table 1, Figure S7). The differences in the polymeric chain lengths were also confirmed using UV–Vis analysis. As shown in Figure S5, the dithiocarbonate groups possess the absorption band at 280 nm [38], where intensity

decreases as more monomer is incorporated into the structure of the material. An increase in molecular weight values for block copolymers, in comparison to the corresponding homopolymers, confirms successful RAFT copolymerization. For the copolymer of the lowest theoretical number average molecular weight ($M_{n,th}$) (PT-1), good agreement was observed between $M_{n,th}$ and the value obtained from SEC (Figure S7, Table 1). However, in the cases of **PT-2** and **PT-3**, the $M_{n,SEC}$ values differ from the theoretical ones (Table 1). This is most likely due to chain transfer to solvent, which dictates an upper limit in accessible molecular weight [39,40]. The molecular weights of block copolymers could not be determined from the ^{1}H NMR spectra, because the intensity of the dithiocarbonate group signal was at the noise level, making reliable calculations impossible.



Scheme 1. Schematic representation of the synthesis of polymers (considering the theoretical numbers of monomers and folic acid units).

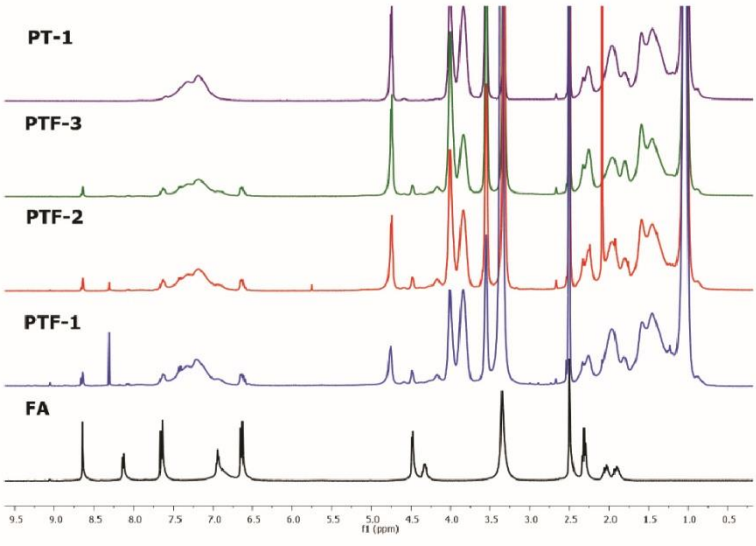


Figure 1. Juxtaposition of ^{1}H NMR spectra of folic acid, folate-conjugated **PTF-1**, **PTF-2**, and **PTF-3** polymers, and an example of non-conjugated polymer **PT-1** in DMSO-d_6.

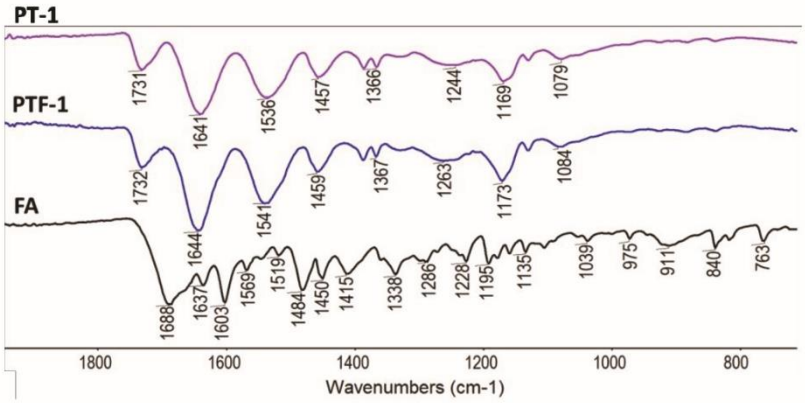


Figure 2. ATR-FT IR spectra of folic acid, **PT-1**, and folic-acid-conjugated **PTF-1** polymer (magnification of the region below 1900 cm^{-1}).

Table 1. Summary of molecular weights of copolymers based on SEC.

Polymer	wt% FA_{th} [a]	wt% FA [b]	$M_{n,th}$ ($g \cdot mol^{-1}$) [c]	$M_{n,NMR}$ ($g \cdot mol^{-1}$) [d]	$M_{n,SEC}$ ($g \cdot mol^{-1}$) [e]	Đ [e]
P-1	-	-	2710	3500	2740	1.87
P-2	-	-	5220	5800	6640	1.55
P-3	-	-	10,210	8700	8300	1.97
PT-1	-	-	9210	-	9070	1.40
PT-2	-	-	12,600	-	9780	1.42
PT-3	-	-	19,610	-	11,040	1.55
PTF-1	33	10	13,700	-	13,040	1.65
PTF-2	29	9	17,840	-	14,700	1.51
PTF-3	26	8	26,500	-	17,040	1.57

[a] wt% FAth = (equivFA × MFA × convFA): Mn,th. [b] Calculated using the calibration curve of FA absorbance at 280 nm in distilled H_2O vs. concentration (Figure S5). [c] Mn,th = MCTA + MHEA × equivHEA × convHEA + MNIPAAm × equivNIPAAm × convNIPAAm + equivFA × MFA × convFA; numbers of equivalents and degrees of conversion are taken from Tables S1–S3; we assume that the conversion of folic acid is quantitative. [d] Measured using ^{1}H NMR. [e] Measured using SEC-RI-MALS.

2.2. Conjugation of Folic Acid

The third step in the synthesis of target polymers was the esterification reaction between the hydroxyl groups of PHEA-*b*-PNIPAAm and the carboxyl group of the folic acid using *N*,*N*′-dicyclohexylcarbodiimide (DCC) and 4-dimethylaminopyridine (DMAP) (Scheme 1, Table S3). We initially performed the coupling of each copolymer with a quantitative amount of folic acid and coupling reagents based on the number of -OH groups, followed by extensive dialysis and freeze-drying. However, the resulting polymers became insoluble in water and phosphate-buffered saline (PBS). Therefore, we carried out the esterification of polymers (PT-(1-3)) with a reduced amount of folic acid, DCC, and DMAP, followed by extensive dialysis leading to the removal of unreacted low molecular weight substrates and by-products. It should be mentioned that two esterification products were probably obtained [41]. We believe that the excess of hydroxyl groups in the system, as well as the appropriate order of adding the reagents, causes only a minimal side reaction between the extreme -NH_2 group and -COOH groups of the folic acid. The resulting folate-conjugated resins, P[(HEA-FA)-*ran*-(HEA)]-*b*-P(NIPAAm) (PTF-1, PTF-2, PTF-3), had folic acid randomly distributed in the structure of the PHEA block. Furthermore, the resins were assumed to have an equal amount of folic acid but differ in the number of non-functionalized -OH groups, ensuring better solubility in water and PBS.

The successful functionalization with folic acid was supported by ^{1}H NMR analysis. The ^{1}H NMR spectrum of the PTF-1 polymer shows characteristic signals in the range of 8.7 to 6.5 nm corresponding to the aromatic protons and the pterine moiety of FA (Figure S3). The signal from the amine group overlaps with the strong signal from the -NH group of NIPAAm. A comparison of the spectra of all folic-acid-functionalized polymers with the spectrum of folic acid and the exemplary unfunctionalized polymer PT-1 in DMSO-d_6 is presented in Figure 1. The presence of folic acid proton signals in the PTF-1, PTF-2, and PTF-3 spectra indicates the successful completion of the coupling reaction.

Furthermore, ATR-FT IR spectroscopy supported the presence of FA in the structure of folate-functionalized polymers. Although signals responding to folic acid or formed ester bonds were overlapped by bands characteristic of functional groups present in the PHEA-b-NIPAAm polymers, some differences may be indicated. Comparing the spectrum showing the enlarged region below 1900 cm^{-1} of the polymer **PT-1** with the spectrum of its folic-acid-conjugated counterpart **PTF-1**, an increase in the signal intensity can be seen at 1644 cm^{-1}, corresponding to C=O stretching of the secondary amine in the folic acid structure, compared to the signal at 1732 cm^{-1} (Figure 2) [42]. This is all the more striking because we expected an increase in the intensity of the second signal mentioned due to the C=O stretching of the newly formed ester bond. The same relationship can be seen when comparing polymers in which the theoretical mass content of folic acid is lower than their counterparts, that is, **PTF-2** with **PT-2** and **PTF-3** with **PT-3** (Figure S4).

UV–Vis spectroscopy was used to support the successful post-functionalization of copolymers with folic acid (Figures S5 and S6). The vitamin folate absorbs ultraviolet light and has absorption peaks at 280 and 350 nm [43]. On the spectra of **PTF-1**, **PTF-2**, and **PTF-3** polymers in deionized water in a concentration of 0.2 mg·mL^{-1}, these two bands can be observed. However, the signal at 350 nm is more reliable as the signal intensity at 280 nm may be slightly overestimated due to absorption by the dithiocarbonate group (Figure S6). The bands are more intense when more FA moieties are present in the polymer structure, providing an expected series of materials. The absorbance values were used to calculate the exact folic acid content of the samples (Table S6). The calibration curve of folic acid in H_2O performed previously was applied [18]. The values obtained are equal to 8–10 wt% (Table 1), about three times lower than the theoretical ones. This is due to the incomplete conversion of the reagents in the esterification reaction, but also due to the intensive dialysis through which some of the functionalized PTF chains were removed from the sample.

The molecular weights of folate-conjugated block copolymers were determined using SEC analysis and are given in Table 1 (and Figure S7). After functionalization of PHEA-*b*-PNIPAAm copolymers with FA, there was a $M_{n,SEC}$ and Đ increase (Table 1), which can be explained by the random presence of FA in the polymeric chains.

The thermal stability of the copolymers and their conjugates with FA were studied using thermogravimetric analysis (Figures S8–S10, Table S4). The tested materials decomposed almost totally, showing one significant weight loss between 300–500 °C. The maximum decomposition rate differed depending on the type of polymer. It was around 440 °C for the PHEA homopolymers, 415 °C for the block copolymers, and 405 °C for the FA-conjugated block copolymers (Table S4). The residues observed on the TG curves of the FA-conjugated samples (PTF-1, PTF-2, PTF-3) at 800 °C were larger compared to the ones determined for the block copolymers (PT-1, PT-2, and PT-3) (Table S4). This confirms the presence of FA, which does not decompose totally in an inert atmosphere in the applied temperature range [18]. Additionally, TG analysis was performed for **PTF-1** micelles containing 5-FU (Figure S8, Table S4). In comparison to the **PTF-1** (without 5-FU), a shift of the maximum degradation rate to a lower temperature and an increase in the amount of residue at 800 °C were observed, confirming modification of the material. The glass transition temperature (T_g) values determined using DSC for PHEA-*b*-PNIPAAm copolymers (82.0, 94.0, 83.2 °C, Table S4) differed from the ones designated for PHEA (<0 °C, Table S4) and known for PNIPAAm homopolymers (>100 °C) [22]. Moreover, the modification of the

block copolymers with FA results created a further increase in T_g (Table S4). This confirms different compositions of the samples.

2.3. Hydrodynamic Diameter and Colloidal Stability Measurements

Dynamic light scattering (DLS) and zeta potential measurements were used to characterize the polymeric structures formed in aqueous media (Figures 3 and S11). The samples were prepared by dropping a concentrated polymer solution in DMF into the water, followed by dialysis and freeze-drying. The measurements were carried out using solutions with a polymer concentration of 0.5 mg·mL^{-1} after stabilization in the dark for 24 h at 25 °C. The block copolymers **PT-(1-2)** form large unorganized aggregates with hydrodynamic sizes between 40–100 nm, while PT-3, with the longest PHEA chain, forms smaller (around 10 nm) and more stable aggregates. The presence of randomly distributed folic acid molecules in the FA-conjugated PHAE-*b*-NIPAAm chains changes the hydrodynamic sizes of the **PTF-2** but does not significantly affect **PTF-1** and **PTF-3**. However, differences in zeta potential values prove that the presence of folic acid results in the formation of more stable structures (Figure 3, Table 2). Furthermore, all six polymers were analyzed in the presence of the anticancer drug, 5-fluorouracil. The samples were prepared by dropping a mixture of polymer and 5-FU in DMF into the water, followed by dialysis and freeze-drying. In the presence of the drug, all polymers form small structures, below 10 nm. It can be supposed that the interaction between the hydrophilic drug and the hydrophilic block, PHEA, resulted in the formation of smaller and more stable aggregates, which was also confirmed by zeta potential results (Figure 3).

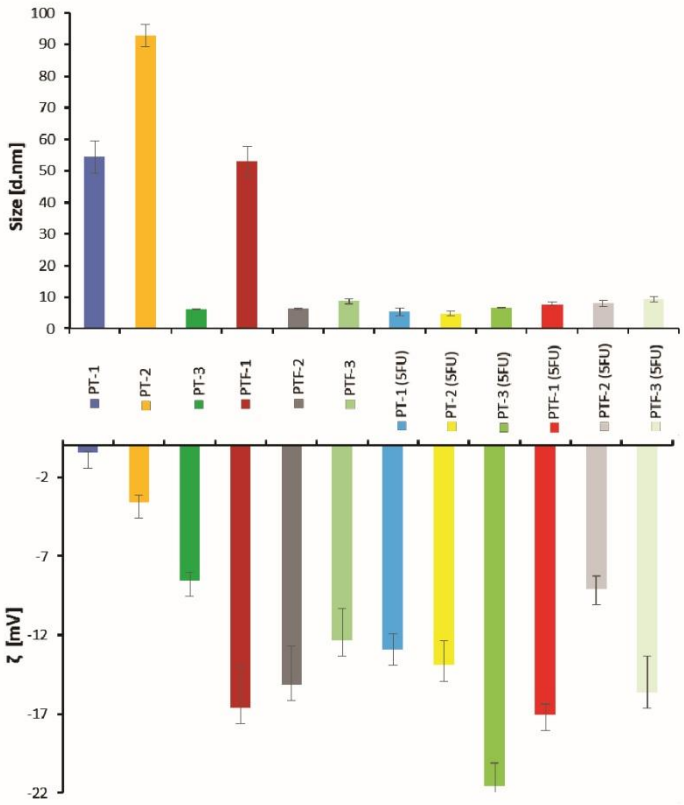


Figure 3. MADLS (size by number) and zeta potential measurements data of all prepared polymers.

Table 2. MADLS (size by number) and zeta potential measurement data of all prepared polymers.

Polymer	Parameters	
	Size (nm)	ζ [mV]
PT-1	54.33 ± 4.98	−0.44 ± 0.07
PT-2	92.83 ± 3.66	−3.56 ± 0.45
PT-3	6.21 ± 0.11	−8.56 ± 0.54
PTF-1	53.10 ± 4.60	−16.61 ± 2.67
PTF-2	6.37 ± 0.28	−15.18 ± 2.51
PTF-3	8.69 ± 0.70	−12.34 ± 2.02
PT-1 (5FU)	5.33 ± 1.27	−12.94 ± 1.02
PT-2 (5FU)	4.73 ± 0.63	−13.91 ± 1.52
PT-3 (5FU)	6.66 ± 0.11	−21.55 ± 1.47
PTF-1 (5FU)	7.77 ± 0.54	−17.02 ± 0.65
PTF-2 (5FU)	8.02 ± 0.91	−9.08 ± 0.84
PTF-3 (5FU)	9.4 ± 0.96	−15.64 ± 2.3

Transmission electron microscopy (TEM) imaging was performed to determine the morphology of the polymeric structures containing 5-FU in the aqueous solution (Figure 4). All analyzed structures formed spherical shapes with round edges. Polymeric structures with diameters from 5 to 7 nm were observed when polymers with encapsulated drug molecules (PTF-1 (5-FU), PTF-2 (5-FU), and PTF-3 (5-FU)) were imagined. These results correspond with the DLS output. The folic-acid-conjugated polymers **PTF-1** and **PTF-2** during the preparation of TEM samples aggregate to form larger objects (20 nm in PTF-2) or fall apart to form thin polymeric film (PTF-1). Only **PTF-3** polymeric structures were well visible during TEM imaging, and separated structures around 6 nm were observed (see SI, Figure S12). This stability of **PTF-3** may be caused by larger participation of the PHEA block in the polymeric chain in comparison to **PTF-1** and **PTF-2**.

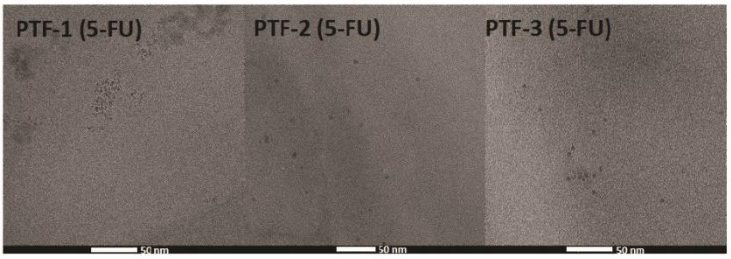


Figure 4. TEM images of polymers with encapsulated 5-fluorouracil.

2.4. Thermoresponsive Behavior

Turbidimetric measurements were carried out to estimate the cloud point temperature (TCP) of prepared polymers (Table 3). Polymeric particles were dissolved in deionized water at a concentration of 1 mg·mL^{-1} and tested in the temperature range from 28 to 40 °C. The change in the light intensity at 550 nm transmitted through the sample was monitored. The phase transition temperature of copolymers **PT-(1-3)** was approximately 35.5 °C, which is about 3.5 °C more than that of pure PNIPAAm possessing sharp cloud point in the range from 31 °C to 33 °C in water, irrespective of the polymer concentration [44]. This is probably due to the long length of the hydrophilic PHEA block in the PHEA-*b*-PNIPAAm polymers, which causes the solubility despite exceeding the phase transition temperature typical for PNIPAAm (around 32 °C). The appearance of folic acid molecules (hydrophilic) in the copolymer structure increased the T_{CP} of **PTF-1** and **PTF-2** polymers by up to 0.5 °C [45]. This can be explained by the formation of hydrogen bonds through its amino and free carboxylic groups. The phase transition temperature of **PTF-3**, which contained the smallest amount of FA, remained unchanged.

Table 3. Turbidimetry data for PHEA-*b*-PNIPAAm and P[(HEA-FA)-*ran*-(HEA)]-*b*-PNIPAAm polymers [a].

Polymer	T_{CP} [°C]	
	Without 5-FU	With 5-FU
PT-1	35.7	36.0
PT-2	35.5	36.0
PT-3	35.4	36.0
PTF-1	36.1	36.5
PTF-2	36.0	36.6
PTF-3	35.6	36.5

[a] Solutions of polymeric micelles in water in conc. 1 mg·mL^{-1} in H_2O at 25 °C.

Subsequently, we examined the T_{CP} of structures formed by dissolving polymers in a solution of the drug in DMF, followed by dialysis and freeze-drying. Polymeric carriers with the encapsulated drug were redissolved in deionized water at a concentration of 1 mg·mL^{-1} and tested as particles without a drug. As predicted, the formation of hydrophobic bonds and π-π interactions of the hydrophilic drug molecule with polymers increased their T_{CP}. In the case of copolymers **PT-(1-3)**, an increase of approx. 0.5 °C was noted. In turn, polymers conjugated with folate **PTF-(1-3)** showed an increase in the phase transition temperature by 0.5 to 0.9 °C. In conclusion, the designed carriers, together with a specific drug, led to the formation of structures whose T_{CP} was close to the temperature of the human body and higher than that of pure PNIPAAm. Furthermore, superior T_{CP} was achieved compared to some known systems consisting of FA and PNIPAAm [24,29].

2.5. Drug Encapsulation and Release

5-Fluorouracil was chosen to investigate the drug loading and controlled release behavior of FA-conjugated thermoresponsive polymers. Drug-loaded micelles were prepared by dissolving the polymer in a DMF solution of 5-FU (to 1 mg 5-FU per 5 mg of drug carrier) and then dropping the solution into vigorously stirred water. Then, dialysis was performed to remove unbound drug and organic solvent, and the content of the dialysis membrane was freeze-dried. Aliquots taken from outside the dialysis membrane were used to indirectly determine the drug loading efficiency (LE) and drug loading content (LC) in the carrier. To increase the drug concentration in the samples, the aliquots were freeze-dried, and the obtained dry residue was redissolved in a small amount of water, followed by HPLC analysis. The encapsulation efficiency calculated on this basis was within the range of 16.8% to 89.4% (Table 4). The yield increased in series from **PT-1** to **PT-3**, in accordance with the growth in the number of hydroxyl and amide groups present in the carrier, indicating the formation of hydrogen bonds between 5-FU and the copolymer. In the case of folic-acid-functionalized copolymers, π-π interactions between FA and the drug may be responsible for the encapsulation of 5-FU. According to this, the LE decreases as the amount of FA in the polymer decreases in the series from **PTF-1** to **PTF-3**. Hydrogen bonds seem to play a minor role, perhaps by obscuring the small units possessing -OH and -NH groups with large FA molecules. The obtained LE values translate directly into drug loading content (LC). They range from 3.3% to 15.2%, with 15.2% being the highest for the compounds with the highest LE, thus 15.2% for **PT-3** and 12.2% for **PTF-1** (Table 4). These are satisfactory results, as most drug carriers are characterized by a low drug content, generally not exceeding 10% [46].

Subsequently, we studied the release of the drug from the 5-FU-loaded polymeric micelles. Carriers prepared in the same way as in the case of studying the efficiency of drug encapsulation were dissolved in imitating-physiological-fluid phosphate-buffered saline. The solution was dialyzed against PBS thermostated at 37 °C for 24 h. The release temperature corresponded to the temperature of the human body in a low-grade fever state. It was aimed at exceeding the T_{CP} of carriers, thus precipitating the polymers and pushing 5-FU out of the polymer chains. Aliquots were taken from outside the dialysis membrane after specified periods of time, and loss was replenished with fresh PBS. The

samples were freeze-dried and dissolved in deionized water for further analysis using HPLC. Unfortunately, the calculated drug content was generally hovering around zero. Only in the case of the polymer with a high drug content **PTF-1** was it possible to determine the total release of the adsorbed 5-FU over 24 h as equal to 4.0% (Table 4). Additionally, the influence of the media on drug release was investigated using two selected polymeric systems: **PT-3** and **PTF-1**. In both cases, the degrees of drug release in water could be determined, but they did not exceed 2.0% (Table 4). It can be concluded that in the tested system, the drug is permanently attached to the polymeric structure. Drug loading methods based on complexation or permanent bond formation have already been used in drug delivery. Some of the systems representing these methods are at the stage of clinical trials [14].

Table 4. Drug encapsulation and release data.

Polymer	LE [%] [a]	LC [%] [b]	Release after 24 h [%]	
			PBS	H_2O
PT-1	16.8	3.3	n.d.	-
PT-2	29.5	5.6	n.d.	-
PT-3	89.4	15.2	n.d.	1.5
PTF-1	69.2	12.2	4.0	2.0
PTF-2	45.5	8.3	n.d.	-
PTF-3	41.7	7.7	n.d.	-

[a] Drug loading efficiency (LE) was determined using HPLC. [b] Drug loading content (LC) was determined using HPLC. n.d.—not detected.

2.6. Biological Studies

Colorectal cancer (CRC) is a disease that is classified as one of the most common cancers globally [5]. CRC is also one of the most significant contributors to high cancer-related mortality rates [47]. In the treatment of colorectal cancer, both in monotherapy and in combination therapy, it is recommended that the first-line therapy is based on 5-fluorouracil (5-FU). In the past, several regimen modulation strategies, including the introduction of 5-FU-based combination schemes, have been developed and tested to improve the anticancer effectiveness and overcome the clinical drug resistance of 5-FU and 5-FU prodrugs [48]. Unfortunately, the 5-year survival rate of patients with early diagnosed CRC (stages I and II) is just over 60%, and well over 50% of patients are diagnosed at stage III or later when distant metastases have already developed. In this case, the overall 5-year survival rate decreases to only 10% [47].

Treatment schemes that are 5-FU-based have several disadvantages, such as unpredictable severe toxicity, which is generally associated with personal changes in dihydropyrimidine dehydrogenase (DPD) expression, short biological half-life, and strong side effects such as cardiotoxicity or myelosuppression. The aim of developing novel drug delivery systems is to overcome all of these limitations. The DDS can facilitate the slow and sustained release of 5-FU. This further prevents its in vivo degradation and reduces its toxicity. Undoubtedly, the advantages mentioned above would significantly improve the length and quality of life of patients with CRC [49].

To develop effective nanosized drug delivery systems, understanding the interaction between nanoparticles and host cells is a crucial step in biological studies [50]. Consequently, the lack of compatibility at this research step might be a considerable limitation in further clinical introduction [51]. To evaluate whether the encapsulation of 5-FU in PHEA-*b*-PNIPAAm-based carriers would prevent toxic effects on host cells (including human RBC, monocytic cells, skin, colon fibroblast cells, and cardiomyocyte cells), hemolysis assay and cytotoxicity evaluation have been performed.

The first step of the study was focused on the interaction of human RBC with polymeric nanosystems applied at the highest concentration (Figure 5A). For this purpose, the hemolytic activity of developed micelles in free form and loaded with 5-FU was examined.

Results showed that both forms of tested carriers (PTF and PTF-FU) did not damage the RBC membrane after 1 h incubation. Consequently, synthesized nanosystems proposed as a carrier for 5-FU meet the criteria foreseen in the pharmaceutical recommendation, where the hemolysis level is classified below 10% [52]. Furthermore, they also conform to the conditions for blood-contacting materials, where the acceptable percentage of hemolysis is below 2% [53]. In effect, proposed carriers might be considered in the further step of the research, including in in vivo evaluation.

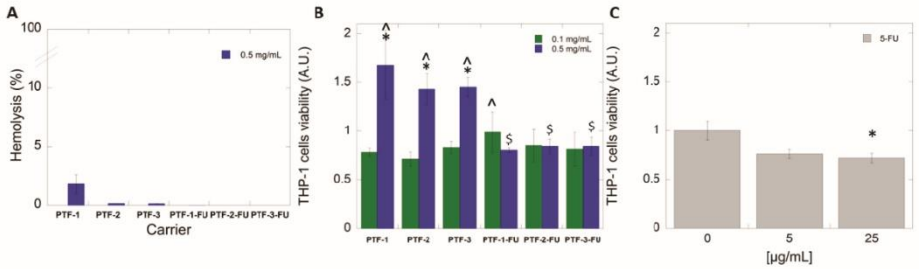


Figure 5. Hemocompatibility of folate-conjugated polymers. Hemolytic activity (panel (**A**)) and viability of monocytic THP-1 cells after the addition of bare folate-conjugated polymers (PTF 1-3) and folate-conjugated polymers with loaded 5-FU (PTF-1-3-FU) (panel (**B**)), and 5-FU (panel (**C**)). Statistical significance for the bare folate-conjugated polymers or folate-conjugated polymers loaded with 5-FU or 5-FU at free form vs. control was marked with (*); comparison of 5-FU concentration at free form vs. polymeric form with or without 5-FU was marked with (^); and comparison of bare folate-conjugated polymers (PTF 1-3) vs. folate-conjugated polymers loaded with 5-FU (PTF-1-3-FU) was marked with ($). $p \leq 0.05$. The data presented constitute average results from three measurements ± SD.

During chemotherapy, many cancer patients experience hematological side effects following 5-FU treatment. 5-FU causes severe myelosuppression, leading to morbidity and mortality in cancer patients. This is due to an immunocompromised state and the possibility of serious infection development [54]. As an effect of 5-FU treatment, significant hematologic toxicity appears. This involves a decrease in all types of peripheral blood cell levels, manifested as neutropenia, monocytopenia, thrombocytopenia, and thrombocytosis. In our study, we focus on evaluating the protective effects of synthesized polymeric particles as mitigators against 5-FU-induced hematologic toxicity. For this purpose, THP-1 cells were tested. They are a suitable model of human monocyte/macrophage cells which reflects their physiological function [55]. The choice of these types of cells is also essential because early monocytopenia after chemotherapy is an important risk factor for neutropenia [56]. Our results showed that after treatment with 5-FU, the monocytic cell exerted a ~30–40% reduction in metabolic activity, which is directly associated with a depletion in proliferation and cell viability (Figure 5C). In turn, after the addition of polymeric micelles to the THP-1 cells, an increased ability with regards to proliferation was observed (Figure 5B). Most importantly, the treatment of representative cells by synthesized micelles after loading 5-FU does not significantly affect their metabolic activity and proliferation, independent of applied concentration. This indicates that the favorable influence of the carriers alleviates the cytotoxic effect of antimetabolite.

It is generally accepted that anticancer drugs are powerful tools for individual or combinatory treatment [57,58]. Their application significantly increases survival and depletes the recurrence rate of cancer. However, their use might be limited by cardiotoxicity. During the therapy, cardiotoxicity can be manifested at an early or late stage of treatment. Importantly, cardiac dysfunction might appear as subtle changes in cardiac structure and function up to irreversible heart failure, which is dangerous to the lives of patients [59].

The search for new methods that could restrict the occurrence of cardiac dysfunction is a challenge for current chemotherapeutic strategies.

Results presented in Figure 6A indicate that empty and loaded-with-5-FU polymeric micelles applied at both tested concentrations did not decrease the viability of cardiomyocytes if compared to the control. However, the treatment of cells with 5-FU in free form causes a statistically marked depletion in cell viability—more than 60% (Figure 6B). It should be emphasized that our 5-FU-loaded carriers exert a statistically significant lower cytotoxic effect than 5-FU applied in free form. This indicates the potential of these carriers to prevent the development of chemotherapy-induced cardiotoxicity in cancer patients.

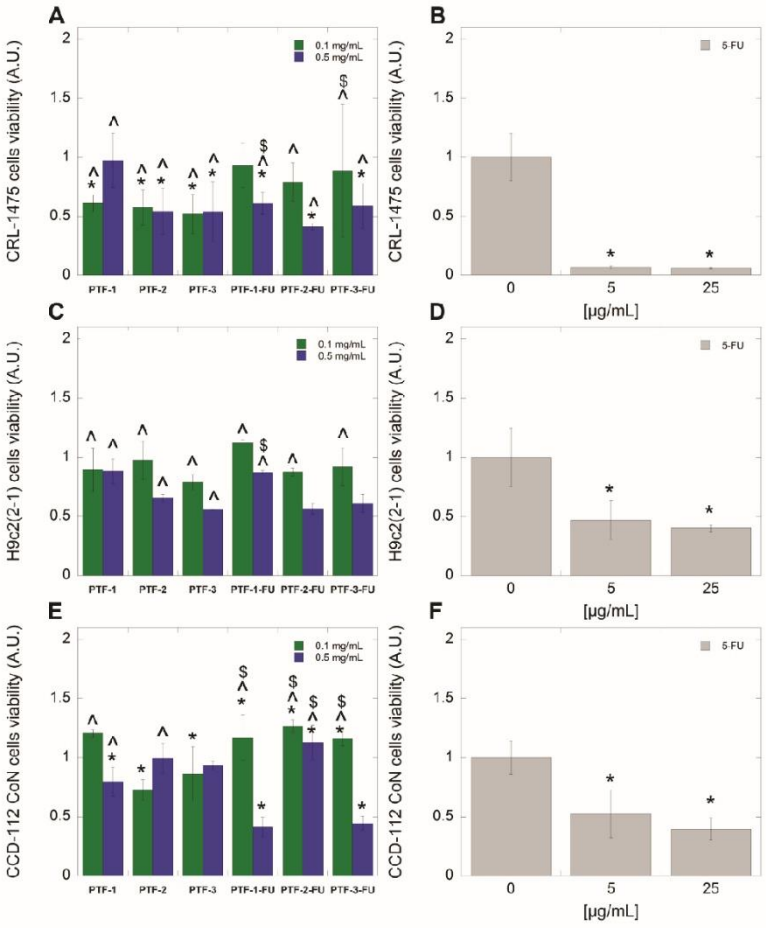


Figure 6. Compatibility of folate-conjugated polymers against representatives of host cells. Viability of cardiomyocyte cells (Panels (**A**,**B**)), skin fibroblast cells (Panels (**C**,**D**)), and colon fibroblast cells (panels (**E**,**F**)) after the addition of bare folate-conjugated polymers (PTF-1-3), folate-conjugated polymers with loaded 5-FU (PTF 1-3-FU), and 5-FU (5 and 25 µg·mL^{-1}). Statistical significance for the bare folate-conjugated polymers or folate-conjugated polymers loaded with 5-FU or 5-FU at free form vs. control was marked with (*); comparison of 5-FU concentration at free form vs. polymeric form with or without 5-FU was marked with (^); and comparison of bare folate-conjugated polymers (PTF 1-3) vs. folate-conjugated polymers loaded with 5-FU (PTF-1-3-FU) was marked with ($), $p \leq 0.05$. The data presented constitute average results from three measurements ± SD.

5-Fluorouracil is a potent inhibitor of the proliferation of fibroblasts, the extracellular matrix cells responsible for conferring strength and resiliency of the skin. The patients undergoing 5-FU-based chemotherapy indicated problems with normal wound healing [60]. This is associated with the fact that cytostatic agents, in a non-specific manner, inhibit the proliferation of normal cells, which are responsible for the regeneration process, and exert a negative impact on the wound healing process. Additionally, the adverse effect of 5-FU chemotherapy is recurrent and causes chronic intestinal mucositis. This is also related to the increased production of pro-inflammatory cytokines and the suppression of efficient healing abilities of the mucosa [61]. In our study, we examined the effect of FA-modified micellar nanocarriers as a delivery system of 5-FU on the viability of skin and colon fibroblast cells (Figure 6C–F). Results showed that the treatment of fibroblast cells with 5-FU applied in free form caused a significant reduction in viable cells below 10% in the case of skin fibroblast and ~40% in the case of colon fibroblast (Figure 6D,F). However, using the synthesized carriers with FA moiety resulted in a statistically significant increased viability of treated cells, simultaneously causing a depletion in 5-FU cytotoxicity. In the case of skin fibroblast, the viability was 5-fold higher when cells had been treated with a micellar form of 5-FU. In turn, the treatment of colon fibroblast caused a 2-fold increase in cells' viability after treatment using folic-acid-modified micelles with loaded 5-FU at concentration 0.1 $mg \cdot mL^{-1}$. The obtained results suggest that the use of a micellar carrier for cytostatic delivery could be helpful in reducing the side and toxic effects related to anticancer treatment.

In the last step of the research, the anticancer potential of synthesized polymeric particles as drug carriers of 5-FU against three different CRC cell lines was investigated. For this purpose, DLD-1, CaCo-2, and HT-29 were chosen for testing sensitivity to 5-FU applied in free form and incorporated form (Figure 7A–F). Tested cells were incubated with empty and loaded nanoparticles at two concentrations (0.1 and 0.5 $mg \cdot mL^{-1}$) and with 5-FU added in the free form at two concentrations (5 and 25 $\mu g \cdot mL^{-1}$) for 24 h. As shown in Figure 7B,D,F, treatment of all kinds of cancerous cell lines with 5-FU in the free form resulted in a lack of statistically significant depletion in survival if the drug was applied at a concentration of 5 $\mu g \cdot mL^{-1}$; at the highest tested dose of 5-FU, i.e., 25 $\mu g \cdot mL^{-1}$, the cell viability of DLD-1 (Figure 7B) and HT-29 (Figure 7F) cells was reduced to 80% and 60%, respectively.

Interestingly, the viability of CaCo-2 cells in the presence of 5-FU has remained unchanged, independent of the applied concentration (Figure 7D). However, the incubation of CRC cells with synthesized carriers indicated divergent effects (Figure 7A,C,E). In the case of DLD-1 cells and CaCo-2 cells, known in the literature as being resistant to 5-FU, our results showed that encapsulation of 5-FU inside the synthesized carriers caused a statistically significant depletion in cell growth and viability by 40% in comparison to the untreated control. The increase in the cytotoxic effect of 5-FU added in the encapsulated form was also observed in the case of HT-29 cells, known as intermediately sensitive to 5-FU. In effect, it could be concluded that evaluated cancer cells were considerably more sensitive to particle-mediated treatment than the free drug.

It is established that the major mechanisms responsible for the development of a 5-FU resistance phenotype in cancer cells are associated with impaired drug uptake and target alterations. Based on antimetabolites, the mode of action induces cytotoxicity via interfering with the biosynthesis of nucleic acids RNA and DNA. We suggested that the application of synthesized FA-functionalized polymeric particles might lead to overcoming 5-FU resistance [62]. The abovementioned suggestion could be accomplished via interaction with FA receptors which provide drug uptake and then inhibition of proliferation, and finally induction of cell death. In order to better explain the proposed mode of action, two CRC cell lines of DLD-1 and HT-29 cells have been engaged [63,64]. The chosen CRC cells possess different molecular and genetic profiles including RER status and TGFbIIR mutations, as well as the expression level of the folate receptors [65]. To evaluate the impact of the synthesized carriers on cancer cells' metabolic activity and proliferation,

a resazurin assay has been performed. Results presented in Figure 8A–D indicate that FA-functionalized polymeric particles significantly inhibit cell division if compared to the control. In the case of DLD-1 cells, after treatment with 5-FU-loaded carriers, more than 50% of cells possessed markedly decreased metabolic activity, while the treatment of cells with 5-FU at free form indicated a lack of impact on cell proliferation. In turn, in the case of HT-29 cells, similar activity has been observed for both forms of the drug (free and encapsulated). It should be emphasized that in both treated cell's lines, an inhibition of cell proliferation has been noted after the treatment of cells with empty carriers, which suggests that some cytotoxic modes of action might be characteristic for carriers alone. In another set of experiments, to elucidate the mode of action, bioluminescent- and fluorometric-based assays were used. After 24 h exposition of CRC cells to synthesized carriers, a bioluminescent assay that measures the exposure of phosphatidylserine (PS) on the outer leaflet of the cell membrane during the apoptotic process was performed. Results indicated that, in the case of DLD-1 cells, loaded carriers caused apoptosis at a level similar to 5-FU applied at a concentration of 20 $\mu g \cdot mL^{-1}$at free form. In turn, the treatment of HT-29 cells with 5-FU-loaded polymeric carriers caused a 2-fold increased apoptosis if compared to drugs applied at free form in a concentration of 20 $\mu g \cdot mL^{-1}$. To investigate more accurately the molecular mechanism involved in observed killing activity, we investigated whether the necrosis pathway might be involved in this process. For this purpose, a fluorescent-based technique was applied. Upon the loss of membrane integrity, the dye enters the cell, then binds to DNA, and generates a fluorescent signal. As demonstrated in Figure 8I–L, synthesized carriers, both empty and loaded with 5-FU, are able to induce cell death more effectively than a drug in free form, independent of applied concentration (20 or 100 $\mu g \cdot mL^{-1}$). This suggests that the main mechanism of action of the proposed carriers is the induction of necrosis in treated cells. The abovementioned suggestion might explain the cytotoxic effect (decreased viability and inhibition of proliferation) observed for empty carriers.

Using 5-FU (or its derivatives) encapsulated in polymeric carriers to treat CRC has been reported recently. Moodley and Singh [66] synthesized polymeric mesoporous silica nanoparticles (MSNs) functionalized with biocompatible polymers, chitosan, and poly(ethylene glycol) for the delivery of 5-FU. The results showed better controlled release profiles (15–65%) over 72 h and cell-specific cytotoxicity against cancer cells. After the in vitro assessment, they also found that these formulations are safe and efficient delivery systems with great potential for in vivo applications. Öztürk et al. [67] have examined the 5-fluorouracil-loaded PCL nanoparticles. The obtained formulations showed high encapsulation efficiency of about 93%. Cytotoxicity studies revealed that PCL nanoparticles containing 5-FU exhibited a higher antiproliferative effect than free form 5-FU on the Caco-2 cell line. In turn, Wang et al. [68] designed PLGA nanoparticles conjugated with folic acid. They found lower LC_{50} for encapsulated 5-FU against HT-29 cancer cells if compared to the free form of the drug. The authors indicated that the presence of the folic acid moiety on the surface of the nanoparticles is responsible for the rapid uptake of the nanoparticle into the cell. Those mentioned above and the currently presented results agree with our previously published study. We introduced the synthesis and biological activity of well-defined polymeric drug delivery systems based on PHEA-PNVCL with folic acid moiety. These studies showed that appropriately created carriers could be used in 5-fluorouracil complexation and combinatory treatment. The statement was established based on cytotoxic results, in which a significant decrease in the viability of the representatives of CRC cells has been noted [18].

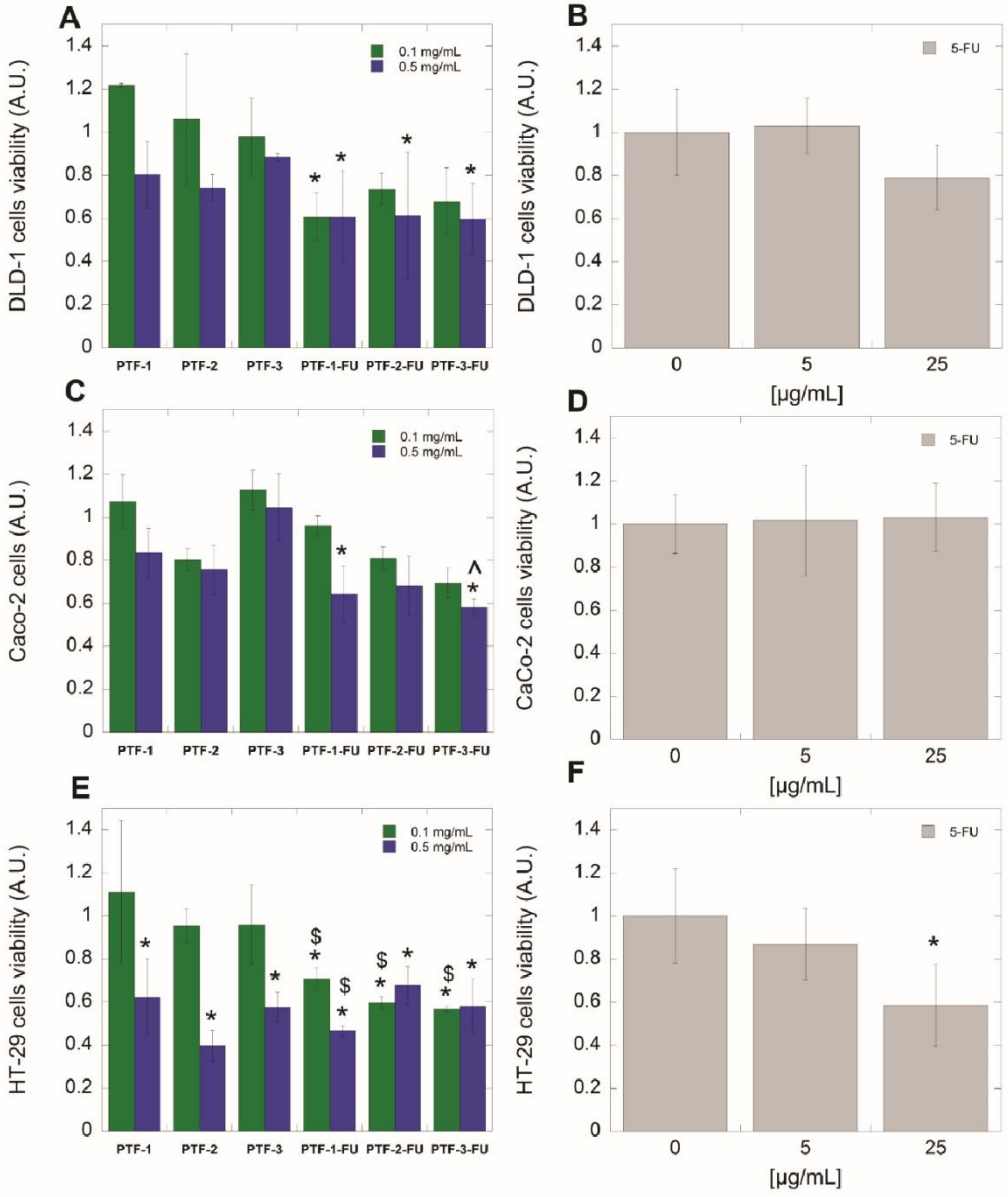


Figure 7. Cytotoxic effect of folate-conjugated polymers against CRC cells. Viability of DLD-1 (panels (**A**,**B**)), CaCo-2 (panels (**C**,**D**)), and HT-29 (panels (**E**,**F**)) cells after the addition of bare folate-conjugated polymers (PTF 1-3), folate-conjugated polymers loaded with 5-FU (PTF-1-3-FU), and 5-FU. Statistical significance for the bare folate-conjugated polymers and folate-conjugated polymers loaded with 5-FU or 5-FU at free form vs. control was marked with (*); comparison of 5-FU concentration at free form vs. polymeric form with or without 5-FU was marked with (^), and comparison of bare folate-conjugated polymers (PTF 1-3) vs. folate-conjugated polymers loaded with 5-FU (PTF-1-3-FU) was marked with ($), $p \leq 0.05$. The data presented constitute average results from three measurements ± SD.

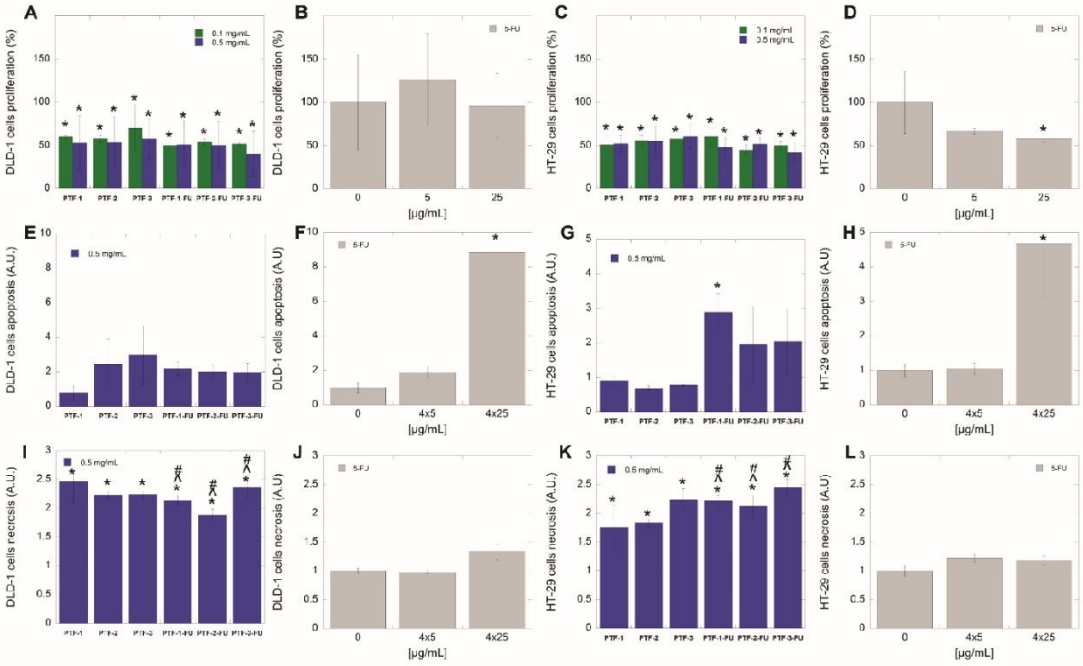


Figure 8. Inhibition of proliferation and mode of action of folate-conjugated polymers against CRC cells. Proliferation of DLD-1 (panels (**A**,**B**)) and HT-29 (panels (**C**,**D**)) cells after the addition of bare folate-conjugated polymers (PTF 1-3), folate-conjugated polymers loaded with 5-FU (PTF-1-3-FU) and 5-FU. Induction of apoptosis in DLD-1 (panels (**E**,**F**)) and HT-29 (panels (**G**,**H**)) cells after the addition of bare folate-conjugated polymers (PTF 1-3), folate-conjugated polymers loaded with 5-FU (PTF-1-3-FU), and 5-FU. Induction of necrosis in DLD-1 (panels (**I**,**J**)) and HT-29 (panels (**K**,**L**)) cells after the addition of bare folate-conjugated polymers (PTF 1-3), folate-conjugated polymers loaded with 5-FU (PTF-1-3-FU), and 5-FU. Statistical significance for the bare folate-conjugated polymers or folate-conjugated polymers loaded with 5-FU or 5-FU at free form vs. control was marked with (*); comparison of 5-FU concentration (5 or 5 × 4 μg·mL^{-1}) at free form vs. polymeric form with or without 5-FU was marked with (^); and comparison of 5-FU concentration (25 or 4 × 25 μg·mL^{-1}) at free form vs. polymeric form with or without 5-FU was marked with (#). $p \leq 0.05$. The data presented constitute average results from three measurements ± SD.

3. Experimental Section

3.1. Materials

The initiator, 2,2′-azobis(2-methylpropionitrile) (AIBN, 98%, MERCK, Darmstadt, Germany) was recrystallized from methanol. Monomer, *N*-isopropylacrylamide (NIPAAm, 99%, ACROS, Geel, Belgium) was recrystallized from toluene–hexane (60:40, *v*/*v*), before use. 2-Hydroxyethyl acrylate (96%, Aldrich, Burlington, MA, USA), folic acid (FA, pure, Alfa Aesar, Kandel, Germany), *N*,*N*′-dicyclohexylcarbodiimide (DCC, 98%, Fluorochem, Pune, India), 4-dimethylaminopyridine (DMAP, ≥99%, Aldrich), 5-fluorouracil (5-FU, Ebewe Pharma, Unterach am Attersee, Austria), pyrene (Py, ≥99%, Aldrich), phosphate-buffered solution (PBS, pH = 7.4, GIBCO, Monza, Italy), and Dulbecco's Modified Eagle Medium (DMEM, GIBCO) were used as received. Potassium *O*-ethylcarbonodithioate [21] and methyl 2-((ethoxycarbonothioyl)thio)propanoate [69] were synthesized according to previously reported procedures. Organic solvents THF, EtOH, and DMF were purchased from Avantor Performance Materials, Poland, S.A., DMSO from Chempur, and MeOH for HPLC from Merck. DMSO and THF were dried over activated molecular sieves 4 Å and stored under

argon. The deuterated solvent was purchased from Armar Chemicals (DMSO-d_6). For all experiments, glassware was dried in a laboratory oven at 120 °C for 20 h.

3.2. Methods

3.2.1. Nuclear Magnetic Resonance (NMR)

1H NMR spectra were recorded on a Bruker Avance II 400 spectrometer at 400 MHz. Chemical shifts δ are given in ppm, referenced to the solvent peak of $CDCl_3$, defined at δ = 7.26, or DMSO-d_6, defined at δ = 2.50.

3.2.2. Attenuated Total Reflectance Fourier Transform Infrared Spectroscopy (ATR-FTIR)

ATR-FTIR spectra were taken on a Thermo Scientific Nicolet 6700 FTIR spectrophotometer, possessing an ATR accessory. Spectra were collected in the wavenumber range from 4000 to 500 cm^{-1} by adding 64 scans with a resolution of 4 cm^{-1}.

3.2.3. Size Exclusion Chromatography (SEC)

The polymers, copolymers, and their conjugates with folic acid were characterized using size exclusion chromatography (SEC). DMF/LiBr (10 mM) solution was used as eluent at 55 °C with a flow rate of 1.0 mL min^{-1}. Prior to injections, the samples were dissolved in the eluent to a concentration of 5 mg mL^{-1} and filtered through 0.45 μm PTFE filters. The samples were analyzed using two column sets, KF-804L and KF-805L Shodex (homopolymers), or TSKgel Alpha-2500 and Alpha-3000 Tosh Bioscience (block copolymers and their FA conjugates), coupled with a three-detector system: a refractometer thermostated at 35 °C (Optilab Rex, Wyatt technology, Santa Barbara, CA, USA), a UV detector (Prostar, Varian, Palo Alto, CA, USA) set at 254 nm, and a multi-angle laser light scattering (MALS) detector (Mini Dawn, 3 angles, Wyatt technology). The dn/dc of PHEA (0.076 $mL{\cdot}g^{-1}$) was taken from the literature [70]. The dn/dc value of block copolymers and their FA conjugates was assumed to be equal to one of the PNIPAAm homopolymers (0.087 $mL{\cdot}g^{-1}$) [71].

3.2.4. Dynamic Light Scattering (DLS) and ζ Potential

The colloidal stability of the polymeric systems was examined using Zetasizer Ultra (Malvern Panalytical Ltd., Malvern, UK) equipped with a 10 mW helium/neon laser (l = 633 nm) at 25 °C. The instrument settings were optimized automatically using the ZS XPLORER software (Malvern Panalytical Ltd., Malvern, UK). All measurements were carried out using a Multiangle Dynamic Light Scattering (MADLS) detection system. The measurements of polymer samples (prepared according to procedures given in Sections 3.3.4 and 3.3.5) with conc. 0.5 $mg{\cdot}mL^{-1}$ in reversible osmosis water were completed at 25 °C after stabilization in the dark for 24 h. The analyses were repeated five times, two extreme results were rejected, and the remaining mean of three results was taken.

3.2.5. Ultraviolet–Visible Spectroscopy (UV–Vis)

UV–Vis spectra were collected using a Jasco V-670 Spectrophotometer at a wavelength range of 250–460 nm. PHEA-*b*-PNIPAAm polymers and folate-conjugated PHEA-*b*-PNIPAAm polymers in conc. 0.2 $mg{\cdot}mL^{-1}$ in deionized water at 25 °C were analyzed.

3.2.6. Turbidimetry

Thermo-regulated UV–Vis spectroscopy was applied to determine the cloud points of the samples. Particle suspensions were prepared using deionized water at a concentration of 1 $mg{\cdot}mL^{-1}$. A Jasco V-670 Spectrophotometer was used to record spectra at a wavelength of 550 nm in the absorbance mode with a heating rate of 0.5 °C per minute in the temperature range of 28–40 °C. The temperature at which optical transmittance was starting to drop sharply was considered T_{CP}.

3.2.7. Thermogravimetric Analysis (TGA)

Thermogravimetric analyses were conducted using a Mettler Toledo Star TGA/DSC unit. Polymeric samples weighing 2–3 mg placed in aluminum oxide crucibles were heated from 50 °C to 900 °C. The heating rate was equal to 10 $K \cdot min^{-1}$, and the argon flow rate was 40 $mL \cdot min^{-1}$. An empty pan was used as a reference.

3.2.8. Differential Scanning Calorimetry (DSC)

Differential scanning calorimetry (DSC) was performed using a Mettler Toledo Star DSC unit. A sample weighing 2–3 mg was placed in an aluminum crucible and sealed. A sample was first heated from 25 to 200 °C at a rate of 10 $°C \cdot min^{-1}$, held isothermally for 5 min, and then cooled to −50 °C at a rate of −20 $°C \cdot min^{-1}$. Two heating/cooling cycles under an argon flow rate of 200 $mL \cdot min^{-1}$ were performed, and an empty pan was used as a reference. The glass transition temperature (T_g) was taken as the midpoint of the heat capacity change in the second heating run.

3.2.9. Transmission Electron Microscope

In order to be analyzed with a Transmission Electron Microscope (TEM), plunge-freeze-drying was applied to prepare samples. Briefly, solutions of polymers (pretreated according to Sections 3.3.4 and 3.3.5) with a concentration of 0.1 $mg \cdot mL^{-1}$ in deionized water were prepared. Next, 3 µL of the sample was placed on a holey carbon copper grid (200 mesh copper, SPI Supplies, West Chester, PA, USA) and excess of the solution was taken away with a tissue; this was repeated twice. Treated grids were frozen with liquid nitrogen (LN2) and dried overnight on a vacuum pump. A Tecnai G2 X-Twin microscope was used to take images that were taken at the accelerating 200 kV voltage applying LN2 cryotrap for the microscope column.

3.2.10. Freeze-Drying

Samples were freeze-dried on Christ Alpha 1-2 LDplus apparatus equipped with a double chamber. Solutions of polymers in distilled water were frozen with liquid N_2, followed by freeze-drying under 0.013 mbar pressure for 24 h.

3.2.11. Dialysis

Dialysis of folate-conjugated polymers was performed against DMSO or distilled water using dialysis membrane Spectra/Por® 6 (MWCO 1000, surface width 18 mm) at 25 °C. Dialysis of polymeric micelles was performed against distilled water using dialysis membrane ZelluTrans/ROTH T1 (MWCO 3500, surface width 46 mm) at 25 °C.

3.3. Synthetic Procedures

3.3.1. General Procedure for RAFT Polymerization of HEA (P-(1-3))

Methyl 2-((ethoxycarbonothioyl)thio)propanoate (CTA) and 2-hydroxyethyl acrylate (HEA) were dissolved in dry THF under argon. The mixture was immersed in an oil bath and thermostated at 70 °C while stirring. AIBN as a solution in dry THF was added, and polymerization proceeded for 2 h at 70 °C until monomer conversion was high (96–98%). After cooling, the polymer was isolated and purified via precipitation in cold Et_2O, collected, and dried under reduced pressure, affording the product (PHEA, P-(1-3)) as a colorless sticky resin. The HEA unit numbers and molecular weights ($M_{n,NMR}$) were calculated by integrating the ^1H NMR signals of -CH_2CH_3 and -CH_2CH_2OH protons and are given in Table S1. **P-1**: ^1H NMR (DMSO-d_6, 400 MHz): δ 4.80–4.70 (m, OH), 4.61 (q, J = 7.2 Hz, 2H, -CH_2CH_3), 4.05–3.90 (m, -CH_2CH_2OH), 3.60–3.50 (m, -CH_2CH_2OH), 2.45–2.15 (m, -$CH_2CHC(O)$-), 1.90–1.40 (m, -$CH_2CHC(O)$-), 1.37–1.31 (m, 3H, -$CHCH_3$), 1.08–1.03 (m, 3H, -CH_2CH_3). FTIR (neat): 3396 (OH), 2949, 2882, 1724 (C=O), 1448, 1394, 1331, 1238, 1159, 1073, 1022, 889, 842, 757 cm^{-1}.

3.3.2. General Procedure for RAFT Polymerization of PHEA with NIPAAm (PT-(1-3))

Polymer PHEA and *N*-isopropylacrylamide were dissolved in anhydrous THF under argon. The mixture was immersed in an oil bath and thermostated at 70 °C while stirring. AIBN as a solution in anhydrous THF was added, and polymerization proceeded for 5 h at 70 °C. After cooling, the polymer was isolated and purified via double precipitation in cold Et_2O, collected, and dried under reduced pressure, affording the product as (PHEA-*b*-PNIPAAm, PT-(1-3)) a white powder. **PT-1**: ^{1}H NMR (DMSO-d_6, 400 MHz): δ 6.90–7.60 (m, -NH-), 4.85–4.70 (m, OH), 4.10–3.95 (m, -OCH_2CH_2OH), 3.95–3.70 (m, -$CH(CH_3)_2$), 3.60–3.50 (m, -OCH_2CH_2OH), 2.40–2.15 (m, -$CH_2CHC(O)O$-), 2.15–1.15 (m, -$CH_2CHC(O)O$-, -$CH_2CHC(O)NH$-), 1.15–0.85 (m, $CH(CH_3)_2$). FTIR (neat): 3282 (OH), 3075, 2970, 2933, 2875, 1732 (C=O), 1641, 1536, 1457, 1386, 1366, 1329, 1244, 1169, 1130, 1079 cm^{-1}.

3.3.3. General Procedure for Conjugation of PHEA-*b*-PNIPAAm with Folic Acid (PTF-(1-3))

Polymer PHEA-*b*-PNIPAAm was freeze-dried right before the reaction. Folic acid was dissolved in anhydrous DMSO under argon for several hours. The folic acid solution was added to the polymer, DCC, and DMAP, and the reaction proceeded for 48 h at room temperature under argon. Then, the reaction mixture was filtered through a cotton pad and dialyzed against DMSO for 48 h and subsequently against water for 96 h. The aqueous solution of the polymer was freeze-dried, affording the product (PTF-(1-3)) as a yellow light solid. **PTF-1**: ^{1}H NMR (DMSO-d_6, 400 MHz) characteristic signals: δ 8.68–8.58 (m, Ar-H, FA), 7.68–7.58 (m, 2 x -Ar-H, FA), 7.55–6.75 (m, -$NHCH(CH_3)_2$), 6.68–6.56 (m, 2 x -Ar-H, FA), 4.85–4.65 (m, OH), 4.52–4.43 (-CH_2NH-, FA), 4.10–3.95 (m, -OCH_2CH_2OH, -OCH_2CH_2OFA), 3.95–3.70 (m, -$CH(CH_3)_2$), 3.60–3.50 (m, -OCH_2CH_2OH), 2.40–2.15 (m, -$CH_2CHC(O)O$-), 2.15–1.15 (m, -$CH_2CHC(O)O$-, -$CH_2CHC(O)NH$-), 1.15–0.90 (m, $CH(CH_3)_2$). FTIR (neat): 3291 (OH), 2971, 2933, 2874, 1732 (C=O), 1644, 1541, 1458, 1387, 1367, 1263, 1171, 1130, 1080 cm^{-1}.

3.3.4. Polymeric Micelles Formation

15 mg of the polymer were dissolved in 1.5 mL of DMF and added dropwise to 15 mL of distilled water with constant stirring. Next, the sample was dialyzed against 1 L of distilled water for 24 h, changing the water in which the membrane was immersed twice (after 1 h and 3 h). Finally, the polymeric micelles solution (content of the membrane) was freeze-dried and stored in the dark.

3.3.5. Drug Encapsulation

Drug-encapsulated polymer particles were formed as follows: 15 mg of the polymer was dissolved in 1.5 mL solution of 5-FU in DMF (c = 2 mg·mL^{-1}) and added dropwise to 15 mL of distilled water with constant stirring. Next, it was dialyzed against 1 L of distilled water for 24 h, changing the water twice (after 1 h and 3 h). The membrane content (polymeric particles with encapsulated drug) was freeze-dried and further analyzed.

The encapsulation efficiency was measured indirectly by measuring the content of the non-encapsulated drug for each sample as follows: aliquots outside the membrane (30 mL) from each 1 L of water used in dialysis were taken, combined (90 mL in total), and freeze-dried. Next, samples were dissolved in 3 mL of deionized water, filtered using 0.45 μm PTFE filters, and analyzed using high-performance liquid chromatography (HPLC). First, a calibration curve of 5-FU (1.0–0.01 mg·mL^{-1}) aqueous solutions was made using HPLC with a UV detector. A ThermoScientific Hypersilil GOLD 25005-254630 RP column (4.6 × 250 mm, 5 μm) was used for separation, maintaining the column temperature at 20 °C. The standards and samples were analyzed using a mobile phase of water and methanol (90:10, *v/v*) at a flow rate of 1.0 mg·mL^{-1}. The volume of injection was 20 μL. The peak of 5-FU was detected at the wavelength of 265 nm.

Drug loading efficiency (LE) and drug loading content (LC) were calculated as follows:

$$LE = \frac{\text{mass of 5FU in carriers}}{\text{mass of 5FU feed}}$$

$$LC = \frac{\text{mass of 5FU in carriers}}{\text{mass of 5FU loaded carriers}}$$

3.3.6. Drug Release

5 mg of polymeric micelles with the encapsulated drug were dissolved in 5 mL of PBS and dialyzed against 100 mL of PBS thermostated at 37 °C while stirring. Aliquots (10 mL) were taken from outside the membrane after 15 min, 30 min, 45 min, 60 min, 90 min, 2 h, 4 h, 6 h, 8 h, and 24 h while adding PBS each time to maintain the total PBS volume of 100 mL. Aliquots were freeze-dried, dissolved in 1 mL of deionized water, and further analyzed using HPLC. First, a calibration curve of 5-FU solutions in PBS (0.1–0.001 mg·mL^{-1}) was made using HPLC with a UV detector. The same conditions and RP column were used for drug release analysis, as for drug encapsulation. As a result, the drug content in the samples oscillated at around zero. For this reason, only the highest value for all samples after 24 h, calculated without the effect of dilution, is given in Table 4. Drug release from selected samples (PT-3 and PTF-1) was also performed in distilled water using an analogous procedure to investigate the effect of the medium on 5-fluorouracil released from micelles.

3.4. Biological Studies

3.4.1. Hemolysis Assay

The hemolytic activity was tested using human red blood cells (RBCs) suspended in phosphate-buffered saline (PBS) (hematocrit ~5%) with a high applied concentration of empty or 5-FU-entrapped polymeric agents −0.5 mg·mL^{-1}. Briefly, the RBCs were incubated with tested agents for 1 h at 37 °C. After the incubation, samples were centrifuged, and absorbance was measured at 540 nm. Hemolysis was calculated according to the following equation: Hemolysis (%) = [(As − An)/(Ap − An) × 100)], where Ap, As, and An are the absorbance of the positive control, test sample, and negative control, respectively. The positive control was the RBCs lysed with 1% Triton X-100, and the negative control was the human red blood cell suspension treated with PBS.

3.4.2. Cell Culture

Human colorectal adenocarcinoma cell lines (DLD-1, CaCo-2, and HT-29), human colorectal fibroblasts (CCD-112CoN), skin fibroblasts (CRL-1475), human monocytic cell line (THP-1), and rat embryonic cardiomyocytes (H9c2(2-1)) were obtained from the American Type Culture Collection (ATCC). The DLD-1, CaCo-2, CCD-112 CoN, and CRL-1475 cells were grown in RPMI 1640 medium, line HT-29 in McCoy's 5a medium supplemented with 10% fetal bovine serum (FBS) and 1% penicillin/streptomycin, at 37 °C and 5% CO_2. The THP-1 cells were grown in RPMI 1640 medium supplemented with 10% heat-inactivated FBS. The 1% penicillin/streptomycin and 2-mercaptoethanol were added at 37 °C in a 5% CO_2 atmosphere, so the final concentration was 0.05 mM. The H9c2(2-1) cells were grown in Dulbecco's modified Eagle's Medium supplemented with 10% FBS. 1% penicillin/streptomycin was added, and cells were cultured at 37 °C in 5% CO_2-air.

3.4.3. Cytotoxicity Assay

In vitro cytotoxicity was evaluated by performing neutral red assay of the representatives of the healthy host (cardiomyocyte, skin fibroblast, and colon fibroblast cells) and colorectal cancer cells. In brief, polymeric carriers in free form or encapsulated with 5-FU form at concentrations of 0.1 and 0.5 mg·mL^{-1} were added to treated cells and incubated for 24 h. Simultaneously, the cells were treated with 5-FU applied at concentrations of 5 and 25 µg·mL^{-1}. After exposure, the percentage of viable cells was measured using spectrophotometric methods. For this purpose,10 µL of 0.33% neutral red solution was

added to each well and incubated. Following this, neutral red was removed after 2–3 h, and then to the cells, neutral red assay fixative (100 µL) was thoroughly added. As a final step, the fixative solution was removed, and the incorporated dye was then solubilized in an adequate volume of solubilization solution (100 µL). After that, the absorbance was measured at 540 nm, and the results were normalized to the control.

In another set of experiments the proliferation and metabolic activity of representatives of CRC cells–DLD-1 and HT-29 cells were determined using resazurin-based assay. For this purpose, the cells were treated with the tested polymeric carriers, 5-FU-loaded polymeric carriers, and free 5-FU, and applied at concentrations of 5 and 25 $\mu g \cdot mL^{-1}$. After 24 h of exposure, 10 µL of resazurin reagent was added to each well. Then, the cells were incubated for 2 h in the dark at 37 °C in a 5% CO_2 atmosphere. Absorbance was recorded at 570 nm using a microplate reader. The results were normalized to the control.

During the next series of experiments, the viability and metabolic activity of THP-1 monocytic cells were evaluated after treatment with the tested polymeric micelles applied as free or encapsulated with 5-FU form using the MTS assay. After 24 h of exposure, 20 µL of MTS reagent was added to each well. Then, the cells were incubated for 2 h in the dark at 37 °C in a 5% CO_2 atmosphere. Absorbance was measured at 490 nm using a microplate reader. The results were normalized to the control.

3.4.4. Mode of Action–Apoptosis and Necrosis Detection

To examine the ability of the synthesized carriers to induce apoptosis and necrosis, multiplex assay including bioluminescent annexin v assay and fluorometric assay–necrosis assay were performed. In brief, polymeric carriers in free form or encapsulated with 5-FU form at concentrations of 0.5 $mg \cdot mL^{-1}$ were added to treated cells and incubated for 24 h. Simultaneously, the cells were treated with 5-FU applied at concentrations 4×5 and 4×25 $\mu g \cdot mL^{-1}$. Then, an equal volume (100 µL) of 2× Detection Reagent was added. After 24 h of exposure, luminescence and fluorescence ex 485 nm em 530 nm signal were collected. The results were normalized to the untreated control.

4. Statistical Analysis

Statistical analyses were performed using Statistica 13.3 software (StatSoft Inc., Tulsa, OK, USA). Three replicates were measured per time point in each cell experiment. The results were normalized to the control and reported as mean ± standard deviation. For statistical calculations, a one-way analysis of variance (ANOVA) with Dunnett's correction was used. Statistical significance was accepted at $p < 0.05$.

5. Conclusions

Current cancer treatment regimens help to increase survival rates and prognosis, which undoubtedly results in a pharmacoeconomic aspect as a reduction in costs is associated with the treatment process. Despite this, the challenge of fighting against cancer is not over, especially in the case of CRC, considered one of the most aggressive types of cancer.

Created carriers loaded with 5-FU showed significantly lower cytotoxicity against representative host cells and a notable reduction in the viability of CRC cells, including those with a resistance phenotype. Using multiplex-based assay, the ability of synthesized carriers to induce apoptosis and necrosis in treated CRC cells has been confirmed.

The presented polymeric nanosystem may function as a mitigator of 5-FU-induced cytotoxicity against normal cells and a potent drug delivery system against cancer cells. Additionally, it could simplify the therapeutic approaches and overcome the problems of the current 5-FU delivery strategies.

Finally, it could be concluded that the obtained results have underlined the potential of using folate-conjugated polymeric carriers, which could play an essential role in the treatment of colorectal cancer.

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Institutional Review Board Statement: The hemolytic activity of the tested agents was evaluated in blood samples from healthy adult volunteers under IRB approval: R-I-002/254/2019. This study was approved by the Institutional Review Board (IRB) of The Medical University of Bialystok. All subjects provided informed written consent, and the collected samples were anonymous.

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Abbreviations

5-FU: 5-Fluorouracil; ATCC: American Type Culture Collection; ATR-FTIR: attenuated total reflectance Fourier transform infrared spectroscopy; CP: cloud point; CRC: colorectal cancer; CTA: chain transfer agent; D: dispersity; DCC: N.N′-dicyclohexylcarbodiimide; DDS: drug delivery systems; DLS: dynamic light scattering; DMAP: 4-dimethylaminopyridine; FA: folic acid; FBS: fetal bovine serum; FR: folic receptor; FT-IR: Fourier-transform infrared spectroscopy; HPLC: high-performance liquid chromatography; LC: drug loading content; LCST: lower critical solution temperature; LEE: drug loading efficiency; LN2: liquid nitrogen; Mn: molecular weight; NMR: nuclear magnetic resonance; PBS: phosphate-buffered saline; PHEA: Poly(2-hydroxyethyl acrylate); PHEA-b-PNVCL: Poly(2-hydroxyethylacrylate)-b-poly(N-vinylcaprolactam); PLGA: Poly(lactic-co-glycolic acid); PNIPAAm: Poly(N-Isopropylacrylamide); RAFT/MADIX: reversible addition–fragmentation chain transfer/macromolecular design via the Interchange of Xanthates; RBCs: red blood cells; SEC: size exclusion chromatography; TEM: transmission electron microscope; TGA: thermogravimetric analysis; UCST: upper critical solution temperature; UV–Vis: ultraviolet–visible spectroscopy.

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Rozdział 11. Oświadczenie autora rozprawy doktorskiej

Białystok, 24.05.2023 r.

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Oświadczenie autora

Oświadczam, iż mój udział w przygotowaniu publikacji:

1. Sylwia Milewska, Katarzyna Niemirowicz-Laskowska, Gabriela Siemiaszko, Piotr Nowicki, Agnieszka Zofia Wilczewska, Halina Car: Current Trends and Challenges in Pharmacoeconomic Aspects of Nanocarriers as Drug Delivery Systems for Cancer Treatment. International Journal of Nanomedicine, 2021: 16, s. 6593-6644, DOI: 10.2147/IJN.S323831, IF: 7.033, MNiSW: 140.0.

wchodzącej w skład mojej rozprawy doktorskiej polegał na współtworzeniu koncepcji i planu pracy, zebraniu i analizie literatury dotyczącej podjętej tematyki badawczej, przygotowaniu tabel, rycin oraz edycji manuskryptu, co określam jako **75%** udziału w przygotowaniu wyżej wymienionej publikacji.

2. Sylwia Milewska, Gabriela Siemiaszko, Agnieszka Zofia Wilczewska, Iwona Misztalewska-Turkowicz, Karolina Halina Markiewicz, Dawid Szymczuk, Diana Sawicka, Halina Car, Ryszard Lazny, Katarzyna Niemirowicz-Laskowska: Folic-acid-conjugated thermoresponsive polymeric particles for targeted delivery of 5-fluorouracil to CRC cells. International Journal of Molecular Sciences, 2023: 24(2), 1364, s. 1-25, DOI: 10.3390/ijms24021364, IF: 6.208, MNiSW: 140.0.

wchodzącej w skład mojej rozprawy doktorskiej polegał na współtworzeniu koncepcji pracy i hipotez badawczych, przeprowadzeniu badań biologicznych na hodowli *in vitro*, analizie i interpretacji wyników uzyskanych w toku pracy, jak również współudziale w ich analizie statystycznej, przygotowaniu tabel oraz edycji artykułu, co określam jako **60%** udziału w przygotowaniu wyżej wymienionej publikacji.

Sylwia Milewska
Podpis autora rozprawy doktorskiej (czytelny)

Katarzyna Niemirowicz-Laskowska
Podpis promotora (czytelny)

Iwona Misztalewska-Turkowicz
Podpis promotora pomocniczego (czytelny)

Rozdział 12. Oświadczenia współautorów rozprawy doktorskiej

Białystok, 24.05.2023 r.

Halina Car
Imiona i nazwisko współautora

Zakład Farmakologii Doświadczalnej
Wydział Nauk o Zdrowiu
Uniwersytet Medyczny w Białymstoku
Miejsce pracy/afiliacja

Oświadczenie współautora

Oświadczam, iż mój udział w przygotowaniu publikacji:

1. *Sylwia Milewska, Katarzyna Niemirowicz-Laskowska, Gabriela Siemiaszko, Piotr Nowicki, Agnieszka Z Wilczewska, Halina Car: Current Trends and Challenges in Pharmacoeconomic Aspects of Nanocarriers as Drug Delivery Systems for Cancer Treatment. International Journal of Nanomedicine, 2021: 16, s. 6593-6644, DOI: 10.2147/IJN.S323831, IF: 7.033, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na współudziale w tworzeniu koncepcji pracy, nadzorze merytorycznym nad publikacją oraz udziale w przygotowaniu finalnej wersji manuskryptu.

2. *Milewska Sylwia, Siemiaszko Gabriela, Wilczewska Agnieszka Zofia, Misztalewska-Turkowicz Iwona, Markiewicz Karolina Halina, Szymczuk Dawid, Sawicka Diana, Car Halina, Lazny Ryszard, Niemirowicz-Laskowska Katarzyna: Folic-acid-conjugated thermoresponsive polymeric particles for targeted delivery of 5-fluorouracil to CRC cells. International Journal of Molecular Sciences, 2023: 24(2), 1364, s. 1-25, DOI: 10.3390/ijms24021364, IF: 6.208, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na nadzorze merytorycznym oraz przygotowaniu finalnej wersji manuskryptu.

Jednocześnie wyrażam zgodę na przedłożenie wyżej wymienionych prac przez Panią mgr Sylwię Milewską jako część rozprawy doktorskiej w formie spójnego tematycznie cyklu prac opublikowanych w czasopismach naukowych.

Halina Car
.....
Podpis (czytelny)

Białystok, 24.05.2023 r.

Katarzyna Niemirowicz-Laskowska
Imiona i nazwisko współautora

Zakład Farmakologii Doświadczalnej
Wydział Nauk o Zdrowiu
Uniwersytet Medyczny w Białymstoku
Miejsce pracy/afiliacja

Oświadczenie współautora

Oświadczam, iż mój udział w przygotowaniu publikacji:

1. *Sylwia Milewska, Katarzyna Niemirowicz-Laskowska, Gabriela Siemiaszko, Piotr Nowicki, Agnieszka Z Wilczewska, Halina Car: Current Trends and Challenges in Pharmacoeconomic Aspects of Nanocarriers as Drug Delivery Systems for Cancer Treatment. International Journal of Nanomedicine, 2021: 16, s. 6593-6644, DOI: 10.2147/IJN.S323831, IF: 7.033, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na współudziale w tworzeniu koncepcji pracy, nadzorze merytorycznym nad publikacją oraz udziale w przygotowaniu finalnej wersji manuskryptu

2. *Milewska Sylwia, Siemiaszko Gabriela, Wilczewska Agnieszka Zofia, Misztalewska-Turkowicz Iwona, Markiewicz Karolina Halina, Szymczuk Dawid, Sawicka Diana, Car Halina, Lazny Ryszard, Niemirowicz-Laskowska Katarzyna: Folic-acid-conjugated thermoresponsive polymeric particles for targeted delivery of 5-fluorouracil to CRC cells. International Journal of Molecular Sciences, 2023: 24(2), 1364, s. 1-25, DOI: 10.3390/ijms24021364, IF: 6.208, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na współtworzeniu koncepcji pracy, nadzorze merytorycznym dotyczącym części biologicznej publikacji oraz udziale w edycji i przygotowaniu finalnej wersji manuskryptu.

Jednocześnie wyrażam zgodę na przedłożenie wyżej wymienionych prac przez Panią mgr Sylwię Milewską jako część rozprawy doktorskiej w formie spójnego tematycznie cyklu prac opublikowanych w czasopismach naukowych.

Katarzyna Niemirowicz-Laskowska

Podpis (czytelny)

Białystok, 24.05.2023 r.

Diana Sawicka
Imiona i nazwisko współautora

Zakład Farmakologii Doświadczalnej
Wydział Nauk o Zdrowiu
Uniwersytet Medyczny w Białymstoku
Miejsce pracy/afiliacja

Oświadczenie współautora

Oświadczam, iż mój udział w przygotowaniu publikacji:

1. *Milewska Sylwia, Siemiaszko Gabriela, Wilczewska Agnieszka Zofia, Misztalewska-Turkowicz Iwona, Markiewicz Karolina Halina, Szymczuk Dawid, Sawicka Diana, Car Halina, Lazny Ryszard, Niemirowicz-Laskowska Katarzyna: Folic-acid-conjugated thermoresponsive polymeric particles for targeted delivery of 5-fluorouracil to CRC cells. International Journal of Molecular Sciences, 2023: 24(2), 1364, s. 1-25, DOI: 10.3390/ijms24021364, IF: 6.208, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na współudziale w wykonaniu analizy statystycznej otrzymanych wyników badań.

Jednocześnie wyrażam zgodę na przedłożenie wyżej wymienionych prac przez Panią mgr Sylwię Milewską jako część rozprawy doktorskiej w formie spójnego tematycznie cyklu prac opublikowanych w czasopismach naukowych.

Diana Sawicka
Podpis (czytelny)

Białystok, 24.05.2023 r.

Piotr Nowicki
Imiona i nazwisko współautora

Zakład Farmakologii Doświadczalnej
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Miejsce pracy/afiliacja

Oświadczenie współautora

Oświadczam, iż mój udział w przygotowaniu publikacji:

1. *Sylwia Milewska, Katarzyna Niemirowicz-Laskowska, Gabriela Siemiaszko, Piotr Nowicki, Agnieszka Z Wilczewska, Halina Car: Current Trends and Challenges in Pharmacoeconomic Aspects of Nanocarriers as Drug Delivery Systems for Cancer Treatment. International Journal of Nanomedicine, 2021: 16, s. 6593-6644, DOI: 10.2147/IJN.S323831, IF: 7.033, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na konsultacji merytorycznej dotyczącej badań klinicznych oraz udziale w przygotowaniu finalnej wersji manuskryptu.

Jednocześnie wyrażam zgodę na przedłożenie wyżej wymienionej pracy przez Panią mgr Sylwię Milewską jako część rozprawy doktorskiej w formie spójnego tematycznie cyklu prac opublikowanych w czasopismach naukowych.

Piotr Nowicki
.....
Podpis (czytelny)

Białystok, 24.05.2023 r.

Agnieszka Zofia Wilczewska
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Zakład Polimerów i Syntezy Organicznej
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Miejsce pracy/afiliacja

Oświadczenie współautora

Oświadczam, iż mój udział w przygotowaniu publikacji:

1. *Sylwia Milewska, Katarzyna Niemirowicz-Laskowska, Gabriela Siemiaszko, Piotr Nowicki, Agnieszka Z Wilczewska, Halina Car: Current Trends and Challenges in Pharmacoeconomic Aspects of Nanocarriers as Drug Delivery Systems for Cancer Treatment. International Journal of Nanomedicine, 2021: 16, s. 6593-6644, DOI: 10.2147/IJN.S323831, IF: 7.033, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na współudziale w tworzeniu koncepcji pracy, nadzorze merytorycznym nad publikacją oraz udziale w przygotowaniu finalnej wersji manuskryptu w tym na współudziale w wykonaniu rycin.

2. *Milewska Sylwia, Siemiaszko Gabriela, Wilczewska Agnieszka Zofia, Misztalewska-Turkowicz Iwona, Markiewicz Karolina Halina, Szymczuk Dawid, Sawicka Diana, Car Halina, Lazny Ryszard, Niemirowicz-Laskowska Katarzyna: Folic-acid-conjugated thermoresponsive polymeric particles for targeted delivery of 5-fluorouracil to CRC cells. International Journal of Molecular Sciences, 2023: 24(2), 1364, s. 1-25, DOI: 10.3390/ijms24021364, IF: 6.208, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na współtworzeniu koncepcji pracy, nadzorze merytorycznym dotyczącym części chemicznej publikacji, udziale w edycji i przygotowaniu finalnej wersji manuskryptu.

Jednocześnie wyrażam zgodę na przedłożenie wyżej wymienionych prac przez Panią mgr Sylwię Milewską jako część rozprawy doktorskiej w formie spójnego tematycznie cyklu prac opublikowanych w czasopismach naukowych.

Agnieszka Wilczewska
.....
Podpis (czytelny)

Białystok, 24.05.2023 r.

Ryszard Łaźny
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Oświadczenie współautora

Oświadczam, iż mój udział w przygotowaniu publikacji:

1. *Milewska Sylwia, Siemiaszko Gabriela, Wilczewska Agnieszka Zofia, Misztalewska-Turkowicz Iwona, Markiewicz Karolina Halina, Szymczuk Dawid, Sawicka Diana, Car Halina, Lazny Ryszard, Niemirowicz-Laskowska Katarzyna: Folic-acid-conjugated thermoresponsive polymeric particles for targeted delivery of 5-fluorouracil to CRC cells. International Journal of Molecular Sciences, 2023: 24(2), 1364, s. 1-25, DOI: 10.3390/ijms24021364, IF: 6.208, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na udziale w walidacji uzyskanych wyników oraz współudziale w edycji ostatecznej wersji manuskryptu.

Jednocześnie wyrażam zgodę na przedłożenie wyżej wymienionej pracy przez Panią mgr Sylwię Milewską jako część rozprawy doktorskiej w formie spójnego tematycznie cyklu prac opublikowanych w czasopismach naukowych.

R. Łaźny
.....
Podpis (czytelny)

Białystok, 24.05.2023 r.

Gabriela Siemiaszko
Imiona i nazwisko współautora

Zakład Chemii Organicznej
Wydział Farmaceutyczny z Oddziałem
Medycyny Laboratoryjnej
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Miejsce pracy/afiliacja

Oświadczenie współautora

Oświadczam, iż mój udział w przygotowaniu publikacji:

1. *Sylwia Milewska, Katarzyna Niemirowicz-Laskowska, Gabriela Siemiaszko, Piotr Nowicki, Agnieszka Z Wilczewska, Halina Car: Current Trends and Challenges in Pharmacoeconomic Aspects of Nanocarriers as Drug Delivery Systems for Cancer Treatment. International Journal of Nanomedicine, 2021: 16, s. 6593-6644, DOI: 10.2147/IJN.S323831, IF: 7.033, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na współudziale w tworzeniu koncepcji pracy oraz współudziale w opracowaniu części dotyczących chemicznych aspektów nośników leków.

2. *Milewska Sylwia, Siemiaszko Gabriela, Wilczewska Agnieszka Zofia, Misztalewska-Turkowicz Iwona, Markiewicz Karolina Halina, Szymczuk Dawid, Sawicka Diana, Car Halina, Lazny Ryszard, Niemirowicz-Laskowska Katarzyna: Folic-acid-conjugated thermoresponsive polymeric particles for targeted delivery of 5-fluorouracil to CRC cells. International Journal of Molecular Sciences, 2023: 24(2), 1364, s. 1-25, DOI: 10.3390/ijms24021364, IF: 6.208, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na wykonaniu badań z zakresu syntezy i analizy fizykochemicznej nośników leków, udziału w przygotowaniu i edycji finalnej wersji manuskryptu

Jednocześnie wyrażam zgodę na przedłożenie wyżej wymienionych prac przez Panią mgr Sylwię Milewską jako część rozprawy doktorskiej w formie spójnego tematycznie cyklu prac opublikowanych w czasopismach naukowych.

Gabriela Siemiaszko
.....
Podpis (czytelny)

Białystok, 24.05.2023 r.

Karolina Halina Markiewicz
Imiona i nazwisko współautora

Zakład Polimerów i Syntezy Organicznej
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Miejsce pracy/afiliacja

Oświadczenie współautora

Oświadczam, iż mój udział w przygotowaniu publikacji:

1. *Milewska Sylwia, Siemiaszko Gabriela, Wilczewska Agnieszka Zofia, Misztalewska-Turkowicz Iwona, Markiewicz Karolina Halina, Szymczuk Dawid, Sawicka Diana, Car Halina, Lazny Ryszard, Niemirowicz-Laskowska Katarzyna: Folic-acid-conjugated thermoresponsive polymeric particles for targeted delivery of 5-fluorouracil to CRC cells. International Journal of Molecular Sciences, 2023: 24(2), 1364, s. 1-25, DOI: 10.3390/ijms24021364, IF: 6.208, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na współudziale w wykonaniu oraz opisaniu badań fizykochemicznych (TGA, DSC) w manuskrypcie oraz edycji finalnej wersji manuskryptu.

Jednocześnie wyrażam zgodę na przedłożenie wyżej wymienionych prac przez Panią mgr Sylwię Milewską jako część rozprawy doktorskiej w formie spójnego tematycznie cyklu prac opublikowanych w czasopismach naukowych.

Karolina H. Markiewicz
.....
Podpis (czytelny)

Białystok, 24.05.2023 r.

Iwona Misztalewska-Turkowicz
Imiona i nazwisko współautora

Zakład Polimerów i Syntezy Organicznej
Wydział Chemii
Uniwersytet w Białymstoku
Miejsce pracy/afiliacja

Oświadczenie współautora

Oświadczam, iż mój udział w przygotowaniu publikacji:

1. *Milewska Sylwia, Siemiaszko Gabriela, Wilczewska Agnieszka Zofia, Misztalewska-Turkowicz Iwona, Markiewicz Karolina Halina, Szymczuk Dawid, Sawicka Diana, Car Halina, Lazny Ryszard, Niemirowicz-Laskowska Katarzyna: Folic-acid-conjugated thermoresponsive polymeric particles for targeted delivery of 5-fluorouracil to CRC cells. International Journal of Molecular Sciences, 2023: 24(2), 1364, s. 1-25, DOI: 10.3390/ijms24021364, IF: 6.208, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na współudziale w tworzeniu koncepcji pracy, nadzorze merytorycznym oraz udziale w badaniach fizykochemicznych.

Jednocześnie wyrażam zgodę na przedłożenie wyżej wymienionych prac przez Panią mgr Sylwię Milewską jako część rozprawy doktorskiej w formie spójnego tematycznie cyklu prac opublikowanych w czasopismach naukowych.

Podpis (czytelny)

Białystok, 24.05.2023 r.

Dawid Szymczuk
Imiona i nazwisko współautora

Zakład Polimerów i Syntezy Organicznej
Wydział Chemii
Uniwersytet w Białymstoku
Miejsce pracy/afiliacja

Oświadczenie współautora

Oświadczam, iż mój udział w przygotowaniu publikacji:

1. *Milewska Sylwia, Siemiaszko Gabriela, Wilczewska Agnieszka Zofia, Misztalewska-Turkowicz Iwona, Markiewicz Karolina Halina, Szymczuk Dawid, Sawicka Diana, Car Halina, Lazny Ryszard, Niemirowicz-Laskowska Katarzyna: Folic-acid-conjugated thermoresponsive polymeric particles for targeted delivery of 5-fluorouracil to CRC cells. International Journal of Molecular Sciences, 2023: 24(2), 1364, s. 1-25, DOI: 10.3390/ijms24021364, IF: 6.208, MNiSW: 140.0.*

wchodzącej w skład rozprawy doktorskiej Pani mgr Sylwii Milewskiej polegał na współudziale w wykonaniu analiz fizykochemicznych DLS.

Jednocześnie wyrażam zgodę na przedłożenie wyżej wymienionej pracy przez Panią mgr Sylwię Milewską jako część rozprawy doktorskiej w formie spójnego tematycznie cyklu prac opublikowanych w czasopismach naukowych.

Dawid Szymczuk
.....
Podpis (czytelny)

Rozdział 13. Dorobek naukowy

13.1. Wykaz publikacji stanowiących rozprawę doktorską

Publikacja I

Milewska Sylwia, Niemirowicz-Laskowska Katarzyna, Siemiaszko Gabriela, Nowicki Piotr, Wilczewska Agnieszka Zofia, Car Halina: Current Trends and Challenges in Pharmacoeconomic Aspects of Nanocarriers as Drug Delivery Systems for Cancer Treatment. International Journal of Nanomedicine, 2021: 16, s. 6593-6644, DOI: 10.2147/IJN.S323831, IF: **7.033**, MNiSW/MEiN: **140.00**. Cytowania: **10/12** (Web of Science/Scopus).

Publikacja II

Milewska Sylwia, Siemiaszko Gabriela, Wilczewska Agnieszka Zofia, Misztalewska-Turkowicz Iwona, Markiewicz Karolina Halina, Szymczuk Dawid, Sawicka Diana, Car Halina, Lazny Ryszard, Niemirowicz-Laskowska Katarzyna: Folic-acid-conjugated thermoresponsive polymeric particles for targeted delivery of 5-fluorouracil to CRC cells. International Journal of Molecular Sciences, 2023: 24(2), 1364, s. 1-25, DOI: 10.3390/ijms24021364, IF: **6.208**, MNiSW/MEiN: **140.00**.

Sumaryczny Impact Factor (IF) dla cyklu publikacji: **13.241**
Łączna liczba punktów według listy czasopism punktowanych Ministerstwa Edukacji i Nauki (MEiN): **280 punktów**.
h-index: **3**

13.2. Wykaz innych publikacji naukowych

1. **Milewska Sylwia**, Nowicki Piotr, Car Halina, Niemirowicz-Laskowska Katarzyna: Zastosowanie nanonośników 5-fluorouracylu w terapii raka jelita grubego. W: Badania i osiągnięcia z zakresu nauk przyrodniczych, pod. red. Maciąg M., Maciąg K., Lublin 2021, s. 53-67, p-ISBN: 978-83-66489-69-1, IF: **0**, punktacja MNiSW/MEiN: **20.00**.

2. Markiewicz Karolina Halina, Niemirowicz-Laskowska Katarzyna, Szymczuk Dawid, Makarewicz Kacper, Misztalewska-Turkowicz Iwona, Wielgat Przemysław, Majcher-Fitas Anna Małgorzata, **Milewska Sylwia**, Car Halina, Wilczewska Agnieszka Zofia: Magnetic Particles with Polymeric Shells Bearing Cholesterol Moieties Sensitize

Breast Cancer Cells to Low Doses of Doxorubicin, International Journal of Molecular Sciences, DOI: 10.3390/ijms22094898, IF: **6.208**, punktacja MNiSW/MEiN: **140.00**.

3. Siemiaszko Gabriela, Niemirowicz-Laskowska Katarzyna, Markiewicz Karolina Halina, Misztalewska-Turkowicz Iwona, Dudź Ewelina, **Milewska Sylwia**, Misiak Paweł, Kurowska Izabela, Sadowska Anna, Car Halina, Wilczewska Agnieszka Zofia: Synergistic effect of folate-conjugated thermosensitive polymers and 5-fluorouracil in the treatment of colon cancer, Journal of Nanobiotechnology, DOI: 10.1186/s12645-021-00104-9, IF: **7.917**, punktacja MNiSW/MEiN: **100.00**.

4. Car Halina, **Milewska Sylwia**, Niemirowicz-Laskowska Katarzyna: Politerapia i geriatryczny zespół jatrogenny W: Wielkie zespoły geriatryczne. Red. nauk. Mateusz Cybulski, Elżbieta Krajewska-Kułak., Wrocław: Edra Urban & Partner, 2021, s. 4964, p-ISBN: 978-83-66548-75-6, IF: **0**, punktacja MNiSW/MEiN: **20.00**.

Sumaryczny Impact Factor (IF) dla powyższych publikacji: **14.125**
Łączna liczba punktów według listy czasopism punktowanych Ministerstwa Edukacji i Nauki (MEiN): **280 punktów**.
h-index: **3**

13.3. Wykaz doniesień zjazdowych

1. **Milewska Sylwia**, Niemirowicz-Laskowska Katarzyna, Nowicki Piotr, Siemiaszko Gabriela, Misztalewska-Turkowicz Iwona, Wilczewska Agnieszka Zofia, Car Halina: Leczenie raka jelita grubego oparte na nanotechnologii – kosztochłonne czy kosztoefektywne? W: X Jubileuszowa Ogólnopolska Konferencja Naukowo-Szkoleniowa" Farmakoekonomika szansą na zbilansowanie wydatków systemu opieki zdrowotnej w Polsce", Poznań, 17-18.11.2022. Konferencja w formie online.

2. Niemirowicz-Laskowska Katarzyna, **Milewska Sylwia**, Siemiaszko Gabriela, Misztalewska-Turkowicz Iwona, Sadowska Anna, Car Halina, Wilczewska Agnieszka Zofia: Zastosowanie polimerów skoniugowanych z kwasem foliowym w terapii nowotworów jelita grubego. W: VII Konferencja Związki Biologicznie Czynne - Aktywność, Struktura, Synteza, Wydział Chemii Uniwersytetu w Białymstoku,

Białostocki Oddział Polskiego Towarzystwa Chemicznego, Białystok, 24-25 czerwca 2022.

3. **Milewska Sylwia**, Siemiaszko Gabriela, Sadowska Anna, Sawicka Diana, Nowicki Piotr, Misztalewska-Turkowicz Iwona, Wilczewska Agnieszka Zofia, Car Halina, Niemirowicz-Laskowska Katarzyna: The useof folic acid in the targeted treatment of colorectal cancer. W: National Scientific Conference "e-Factory of Science" - VII edition, 09.04.2022 r. Online Conference.

4. Szymczuk Dawid, Markiewicz Karolina Halina, Niemirowicz-Laskowska Katarzyna, Misztalewska-Turkowicz Iwona, Wielgat Przemysław, **Milewska Sylwia**, Car Halina, Wilczewska Agnieszka Zofia: Biological application of magnetic particles with polymeric shells containing cholesterol moieties. W: The Silesian Meetings on Polymer Materials POLYMAT 2022 in memory of Prof. Andrzej Dworak. Zabrze, 17th of March, 2022.

5. **Milewska Sylwia**, Misiak Paweł, Kurowska Izabela, Rogowski Karol: Cholesterol end-capped poly(N-isopropylacrylamide)s as a first step of creation of new drug delivery carriers. W: 15th Bialystok International Medical Congress for Young Scientists, Białystok, 21-22 May, 2021.

6. **Milewska Sylwia**, Niemirowicz-Laskowska Katarzyna, Nowicki Piotr, Siemiaszko Gabriela, Misztalewska-Turkowicz Iwona, Wilczewska Agnieszka Zofia, Car Halina: Farmakoekonomiczne aspekty nanotechnologii. W: IX Ogólnopolska Konferencja Naukowo-Szkoleniowa "Farmakoekonomika szansą na zbilansowanie wydatków systemu opieki zdrowotnej w Polsce", on-line, 19 listopada 2021.

7. **Milewska Sylwia**, Siemiaszko Gabriela, Sadowska Anna, Misztalewska-Turkowicz Iwona, Niemirowicz-Laskowska Katarzyna, Car Halina, Wilczewska Agnieszka Zofia: Zastosowanie nośników polimerowych jako komponentów terapii synergistycznej z 5-fluorouracylem w terapii raka jelita grubego - aspekty doświadczalne i farmakoekonomiczne. W: Ogólnopolska Konferencja Naukowa "Kierunek NANO - badania i osiągnięcia z obszaru nanotechnologii", 27 listopada 2020 r.

13.4. Wykaz innych aktywności naukowych

Aktywność dydaktyczna:

1. Prowadzenie zajęć dydaktycznych dla studentów poniższych kierunków:

- Pielęgniarstwo I° – przedmiot Farmakologia w roku akademickim 2019/2020, 2020/2021, 2021/2022, 2022/2023,

- Położnictwo I° – przedmiot Farmakologia w roku akademickim 2019/2020, 2020/2021, 2021/2022,

- Położnictwo II° – przedmiot Farmakologia i ordynowanie produktów leczniczych w roku akademickim 2022/2023,

- Dietetyka I° – przedmiot Farmakologia i farmakoterapia żywieniowa oraz interakcja leków z żywnością w roku akademickim 2019/2020, 2020/2021, 2021/2022,

- Ratownictwo medyczne – przedmioty: Farmakologia, Farmakologia i toksykologia kliniczna w roku akademickim 2021/2022,

- Zdrowie publiczne – przedmiot Farmakoekonomika w roku akademickim 2020/2021, 2021/2022, 2022/2023.